

**Product Name:** 2X Universe Taq Mix

**Catalog No:** VN2000M, VN2001M

**Packing Size:** 500 x 25 µl rxns and 250 x 25 µl rxns

**Shipping Condition:** Ice pack

**Storage Condition:** -20°C

**Product Description:**

**Universe Taq Mix** is a ready-to-use 2X mixture of Universe Taq polymerase, salts, magnesium and dNTPs for setting up a trouble-free PCR reaction. All you have to do is to add template, specific primers and water, thereby save time, effort and minimize pipetting error.

**Universe Taq Mix** is recommended for all standard PCR applications. The mix is comprised of **Universe Taq** DNA polymerase and an innovative buffer system that deliver very high yield PCR amplification over a wide range of PCR templates, enables universal annealing temperature by isostabilizing primer-template duplex structures. The engineered Taq enzyme confers fast cycling, resistance to common inhibitors and host-start which enable superior specificity and sensitivity. It can work in both purified DNA and crude samples.

**Protocol:**

**PCR setting for a 25 µl reaction**

Reagent	Volume	Final Concentration
2X Universe Taq Mix	12.5 µl	1x
Left Primer	Variable	0.2-0.4 µM
Right Primer	Variable	0.2-0.4 µM
DNA Template <sup>†</sup>	Variable	0.1-100 ng
De-ionized Distilled H <sub>2</sub> O	Adjust final volume to 25 µl	-

<sup>†</sup>DNA amount depends mostly on genome size and target gene copy number.

**Typical Cycling Parameters**

	Three-step		Two-step <sup>1</sup>	
Initial denaturation	95°C	2 min (DNA) 5-10 min (crude samples)	95°C	2 min (DNA) 5 min (crude samples)
25-40 cycles				
Denaturation	94°C	15 to 20 sec	98°C	5 sec
Annealing <sup>2</sup>	60°C	15 to 20 sec	60°C	20 sec
Extension	68°C	15-30 sec / 1kb target		
Hold	4°C			4°C

**Troubleshooting guide**

No PCR products	Please check your PCR settings to ensure that all necessary components are included in the PCR mix. It's also recommended to perform a gradient annealing temperature experiment to determine the optimal temperature for your specific target. This can help to improve PCR efficiency and specificity, resulting in more reliable and consistent results.
The bands in agarose gel are smear	It's important to check for DNA and primer degradation as degraded DNA or primers can also negatively impact the PCR results.
Low yield of products	If you are using crude samples, it's important to initially denature the samples for 5-10 minutes to ensure proper release of DNA. Additionally, you can try the following strategies to improve PCR performance: <ul style="list-style-type: none"> <li>• Conduct an enzyme titration experiment to optimize the enzyme concentration for your specific target.</li> <li>• Increase the extension time to ensure complete amplification of the target DNA.</li> <li>• Try a gradient annealing temperature experiment to identify the optimal annealing temperature for your primers.</li> <li>• Consider redesigning your primers if the amplification is not specific or efficient enough.</li> </ul>
Non-specific products are observed	If you are observing non-specific products in your PCR reaction, there are several strategies you can try: <ul style="list-style-type: none"> <li>• Perform a gradient annealing temperature experiment to identify the optimal temperature for your primers, which can help to reduce non-specific products.</li> <li>• Check the GC content of your target sequence. If it's above 65%, it may be necessary to use PCR enhancers.</li> <li>• Evaluate your primers and redesign them if necessary to improve specificity and reduce non-specific products.</li> </ul>

**Note: This product is for R&D use only**