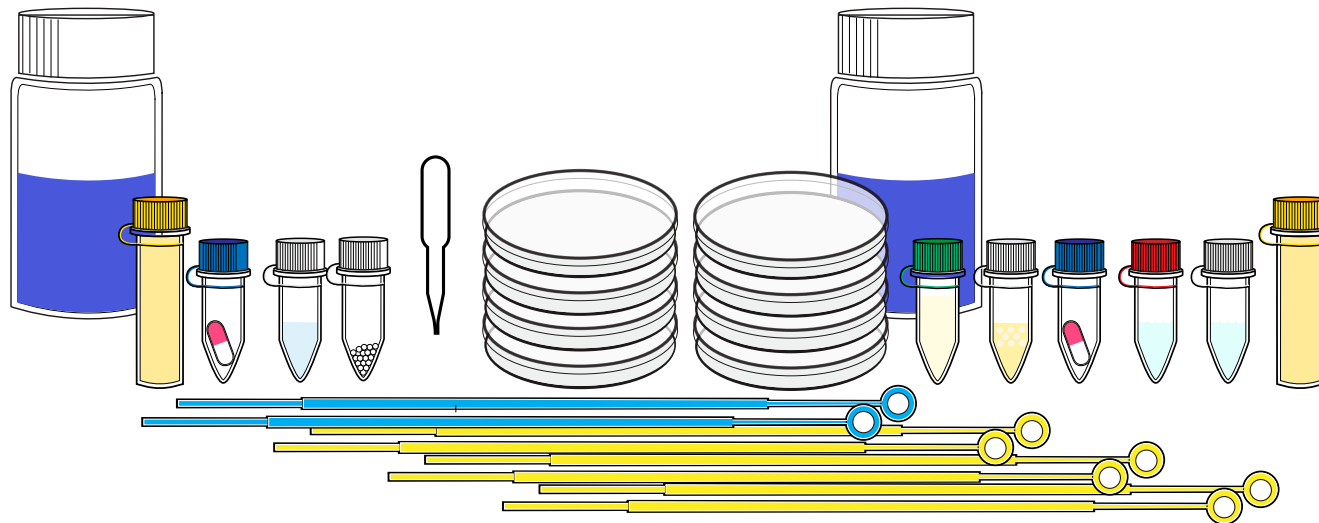


GENETIC ENGINEERING WITH

INDUCE-IT KIT™

— User Manual —

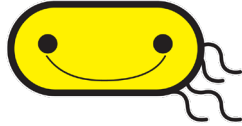


INDUCE-IT KIT™

User Manual

| | | | |
|--|-----------|---|----|
| Welcome! Let's get started | 03 | Checkpoint | 17 |
| Practicing Safe Science | 04 | 3. Creating competent blank cells | 19 |
| How will I learn? | 05 | Checkpoint | 20 |
| Discover your Induce-it Kit | 06 | 4. Transformation | 21 |
| What's in the kit ? | 08 | 5. Plating cells | 22 |
| Storing your kit | 10 | Checkpoint | 23 |
| Technical specifications | 10 | 6. Create fresh selective LB plates | 24 |
| Things you will need | 11 | 7. Streak your cells | 25 |
| Implementation timeline | 12 | 8. Induce your cells and see your results | 26 |
| | | Congratulations | 27 |
| | | Storage, disposal and clean up | 28 |
| Your experiment (Experiment protocol) | | Glossary | 29 |
| 1. Creating LB agar plates | 14 | Troubleshooting | 31 |
| Checkpoint | 15 | More information | 32 |
| 2. Growing blank cells | 16 | | |

Welcome! Let's get started



This User Guide was created to help you get the most out of your Amino Labs Experience. Even if you are familiar with genetic engineering, science or other Amino Labs™ products, please take the necessary time to read through this guide. This will ensure you practice safe science, store, use and get the most out of your Kit and know what to do in case of a spill or accident.

In the first section, you will learn about your kit's components, how to store them before and during your experiment, as well as a few tips on activities to complete before you get your hands wet. The second section is procedural -- these are the step by step instructions on how to run your experiment. Make sure to follow our tips to ensure your best success! The third section covers "what's next"; how to keep your creations, store or dispose of any leftover ingredients and general clean up instructions. And, the final section is there to help you -- a glossary, troubleshooting, and our contact information.

Amino Labs is excited to welcome you to the world of advanced genetic engineering with the Induce-it Kit™ and our entire ecosystem of easy-to-use, easy-to-succeed at products!

Following this guide will help ensure that you are getting the most out of your current and future experiences to keep on making new creations with DNA. Have fun!

Practicing Safe Science

Genetic engineering and life sciences are safe activities when you follow simple guidelines. Read on to ensure you adopt safe practices.

The kit in your hands contains only non-pathogenic ingredients. These are part of the biosafety Risk Group 1 (RG1) (Biosafety Level 1). This is the most benign level and therefore the safest: with these kits, no special containment or training is required in North America. But you must follow these safety guidelines for your safety and the success of your experiment(s)!

We recommend the system and kits for ages 12+, under adult supervision, and 14+ with or without supervision. We recommend that an adult empties the discard container. The cleaning instructions must be strictly followed for safety and experiment success. Make sure to store the kit per the instructions found in this booklet.

- **Do not eat or drink near your experiments.** Keep your experiment at least 10 feet from food, drinks, etc. Under no circumstances should you eat any of the kit's content.
- **Immunocompromised persons:** While the ingredients in these kits are non-pathogenic, some persons, such as immunocompromised persons, can be affected by large numbers of bacteria and should talk to their doctor before doing any experiment.
- **Wash your hands before and after** manipulating your experiment, or the hardware.

- **Wear gloves**, even when cleaning your station or handling the kit contents (petri plates, loops, etc). This will protect you from your experiment, and your experiment from you. Any latex, nitrile, or general purpose gloves you can find at the pharmacy will do. After you put your gloves on, be aware of what you touch. Try not to touch your face, scratch itches with your gloved fingers!
- **If using the DNA Playground™ or BioExplorer™ place it on a stable work surface.** Keep it level at all times.
- **Clean up your station, spills and work surface before and after use.** Use a 10% solution of chlorinated bleach generously sprayed onto a paper towel and rub onto any contaminated surfaces. (Careful! This can discolor your clothes). A chlorinated spray cleaner also works.
- **Find a container to hold the inactivation bag where you will discard used consumables.** An old 1L yogurt container, large plastic cup or the like will do. Used consumables will be loops, any tube or used petri dish.
- **Eye-wear is not provided but can be worn.**

You can download a biosafety poster for your space from www.amino.bio/biosafetyinaction and complete a short safety quiz at www.amino.bio/biosafety-quiz

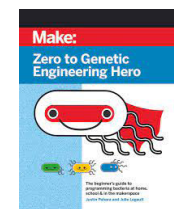
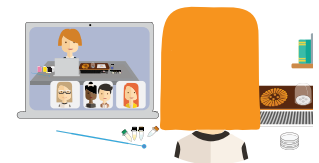
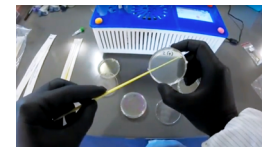
If you would like to do a short Online lab safety course for your edification, we recommend a Government of Canada course: www.amino.bio/biosafety

Find more safe science resources at amino.bio/blogs/news/practice-safe-science

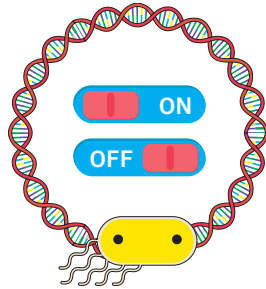
How will I learn?

Learning and prototyping with genetic engineering and cells is becoming accessible to newcomers of all ages and backgrounds thanks to dedicated scientists and kits such as the one you are about to use. One of the easiest way to learn a new science, hobby or topic is by trying it, hands-on. Everything you need to complete the science is included; each ingredient in the kit is pre-measured and labeled for a stress-free experience and our minilabs decrease setup time, mess, guesswork and the need to collect and calibrate multiple machines. These instructions aimed to be easy-to-follow for everyone but may contain some new terms. Therefore, we have added a glossary at the end. We also have additional resources to help you go further in your learning:

- An essential addition to our ecosystem is the free **Virtual Bioengineer™ simulation** developed with the educators at the Biobuilder Educational Foundation. A 20 minutes guided experience that makes it easy to practice using your DNA Playground™ and Induce-it Kit™ beforehand. It also includes additional information on the manipulations and a more in-depth look into DNA and genetic engineering. We recommend it strongly! Complete it online at www.amino.bio/vbioengineer.
- View **real-time tutorials**, including video tutorials for each day of the genetic engineering experience on our Youtube channel. The genetic engineering days are under 'Engineer-it Kit,' the same procedures as the first 3 days of your Induce-it Kit experience . Subscribe! youtube.com/c/AminoLabs.
- Would you like for an Amino Labs team member to guide you through your journey? Try the **Virtual Tutoring or Teacher Professional Development sessions**, a multi-day experience via video www.amino.bio/collections/virtual-sessions
- Are you interested in the theory behind the experiment and in going deeper on the science, learning pro-tips and completing more advance genetic engineering?The **Zero to Genetic Engineering Hero book** is for you. Find out more at www.amino.bio/book

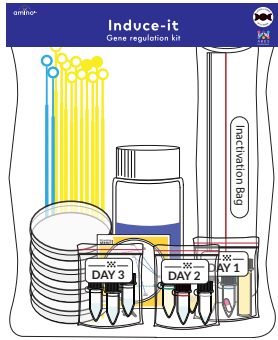


Discover your Induce-it Kit™



The Induce-it Kit™ has everything needed to insert a DNA Program (plasmid) into bacteria, and with your help, have them produce what the DNA “tells” them. In this case, a colored protein that won’t appear until you ‘turn it on’ using a chemical. That is because, in the plasmid, there is a ‘genetic switch’ that is ‘turned off’. That’s right! Just like in computer programming, behaviors in the cells can be turned ‘on’ and ‘off’ using chemicals, or even environmental conditions like temperature and light. If you’ve used one of our Engineer-it kits before, you’ll have engineered cells to produce colored proteins which you can see as colored colonies on the petri dish. In that case, the colored proteins were created automatically because the genetic switch was in the ‘on’ position. With the Induce-it kit, a similar plasmid will be inserted into cells. Once grown, you will then turn the genetic switch ‘on’ using a chemical called Isopropyl β -D-1-thiogalactopyranoside or IPTG for short. IPTG is a common reagent used in laboratory experiments! You can learn more about it, and about genes, genetic switches, and how to use chemicals, light, and temperature to activate DNA programs by reading Chapter 7 of the *Zero to Genetic Engineering Hero* book, www.amino.bio/book. You can also have a look at the Heat-it Kit™ and the RGB kit™ in our store to explore our other genetic switch experiments. www.amino.bio

Note! Amino Labs kits contain the same or similar ingredients in most of its kits but it is important to remember that the bacteria, the antibiotics, DNA, and the different buffers are made to fit perfectly together. Make sure you keep kit components separated!



Individual size

The Individual kit size will let you complete the experiment in full, one time. This kit can be used alone, (with parent supervision if necessary) or in a small group.

Group size

The Group kit size contains:

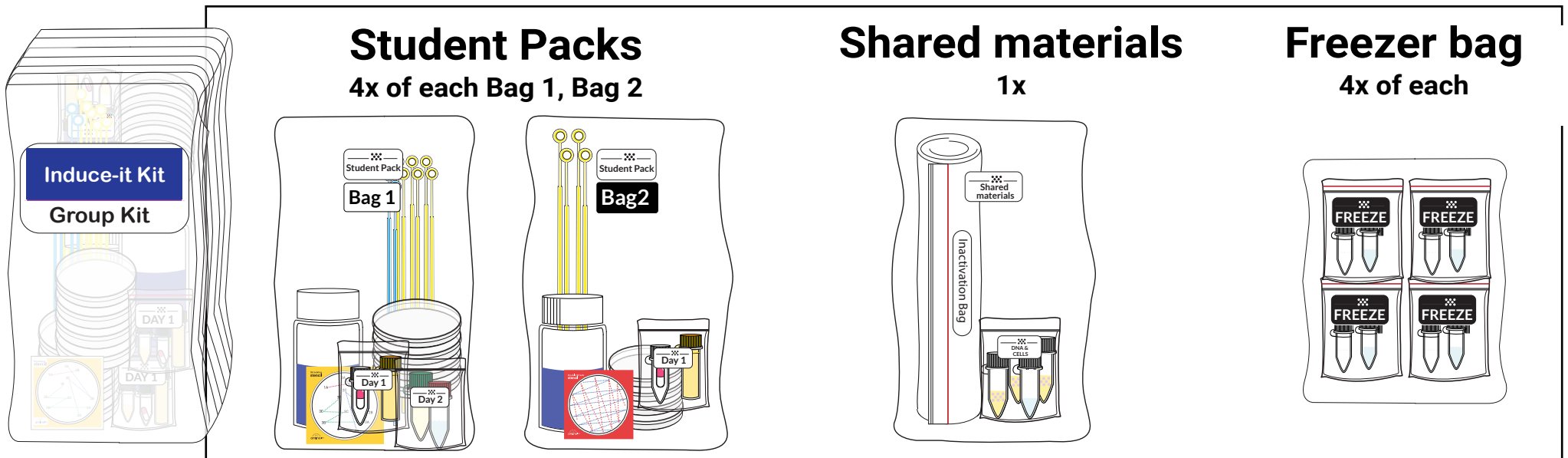
4 individually-wrapped student packs bag 1,

4 individually-wrapped student packs bag 2

1 shared materials bag that contains blank cells, positive control cells, DNA program, and the inactivation bags to be shared by everyone.

1 "Freeze upon arrival" bag with 4 individually-wrapped freezer components, one for each of your student team.

With the group kit you will have everything else you need for the experiment to be done 4 times by student teams/lab groups.



Kit Components

Bag 1



Transformation Buffer: A proprietary transformation buffer used in the bacteria transformation procedure to yield high efficiencies. When you adhere strictly to the protocol, this buffer rivals other available competent cells & procedures.¹



Recovery Media: Amino Labs' recovery media is used after the heat shock during the transformation protocol. This nutrient broth aids the cells in recovering and has a proprietary recipe that further boosts the cells ability to survive the transformation and begin dividing.¹



Agar Powder: This LB agar powder is industry standard. Each tube can make 50 mL of molten LB agar (3.5% w/v). Agar is the surface the bacteria grow on and the food they eat to grow.¹



Cells *: A standard K12 strain of E. Coli, non-pathogenic and the typical strain used by thousands of labs around the world. This strain comes as a "stab," in a small tube of agar.¹



+ Cells * : This stab of E. coli K12 is non-pathogenic and allows you to create a positive control sample by growing it on a plate to test your selective agar and compare to your experiment result.



- Cells *: This stab of E. coli K12 is non-pathogenic and allows you to create a negative control sample by growing it on a plate to to compare to your experimental sample.



DNA *: A DNA plasmid to program your bacteria.



Antibiotics for Transformation: Amino Labs' proprietary antibiotic delivery system helps stabilize antibiotics for shipping and long-term storage. Each capsule has a measured amount of antibiotics for 50 mL of molten LB agar. In such small quantities, these antibiotics are very safe, even if ingested by accident. Do not ingest them, however!¹



Sterile Water: Sterility is critical when genetic engineering. This Sterile water bottle contains distilled water sterilized in an autoclave to ensure there are no contaminating organisms present. This 50 mL volume, when used with LB agar powder is enough to make 5 LB agar plates.¹



Blue Loops: Small inoculating loops are used for transferring 1 uL of liquid and other tasks. These replace costly traditional pipettes.



Yellow Loops: Large inoculating loops are used for transferring 10 uL of liquid and other tasks. Yellow loops are great for spreading out bacteria after a transformation.



Petri Dish / Plate: 6cm Petri dishes are large enough for typical lab experiments and help save on the cost of reagents as well as reduce waste.



Inactivation Bag *: A heavy duty bag to put all of the kit waste in. After your experiment, add bleach and water to the bag to inactivate all the samples and practice safe science.



Plate Streaking Stencil: To help you grow the bacteria into separated, fast-growing colonies for engineering

Bag 2



Agar Powder, Antibiotics for Transformation, Sterile Water, Yellow loops, Petri Dishes: Just like Bag 1, Bag 2 has these necessary items to help you grow cells.



Plate Streaking Stencil: To help you grow the maximum amount of bacteria on your petri dish.

Freezer bag



IPTG (Freezer bag): A chemical that mimics the natural functions of lactose. Used to induce ('turn on') gene expression of a DNA program using a genetic switch called the lac repressor.



Dissolving buffer (Freezer bag): A chemical to dissolve the IPTG powder.



Pipet (Freezer bag): Used to transfer the IPTG from the tube to the petri dishes.

¹ For education purposes only.

* If you have the group kit version, all cells, DNA and inactivation bags will be in the 'Shared materials' bag to be shared by students

Unpacking and Storing your kit

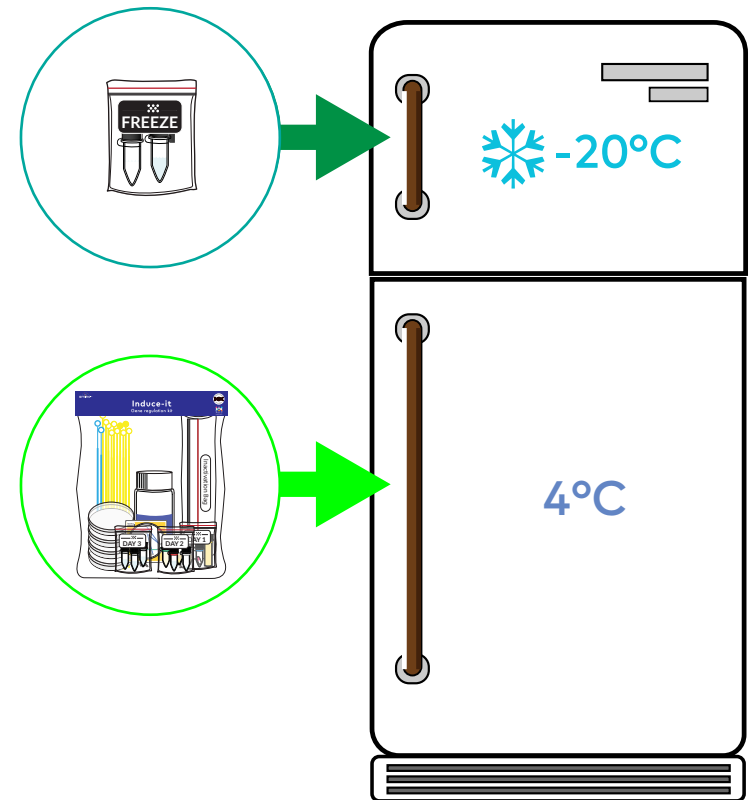
For a better shelf life and successful experiments:

- place your Kit (Bag 1 & Bag 2) in a standard refrigerator at around 4°C.
- place the smaller Freezer bag in a freezer.

Once refrigerated, your kit will be good until the "Best by" date on the sticker on the outer packaging.

Do Not Freeze your kit bags!

Only freeze the small bag with the Freeze sticker.



Technical Specs

DNA plasmid 250 ng
Selection/Antibiotic: variable
Cells /+Cells: K12 E. coli stab

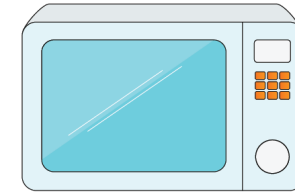
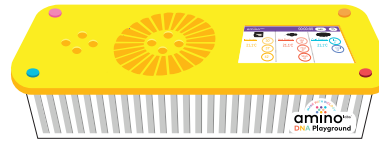
Transformation Buffer: 50 uL tubes
Recovery media: 350 uL tubes
IPTG: 1x100 uL (once dissolved)

Growth plates: 6 cm petri dishes
Growth media:
LB agar powder (1.6 g)
50 mL sterile water

Necessary Equipment

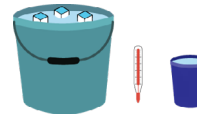
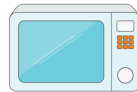
For Best results:

- DNA Playground™
- Microwave



Alternative solution:

- **Microwave**
- **Thermometer** (for 42°C)
- **Timer**



- **Ice bucket or bowl and ice:** This will become your "**Cold station**" "Ice" for the experiment. Make sure to keep the ice from melting too much during the experiment. You may need a fresh replacement during the experiment if it is warm where you are.
- **Hot water bath or bowl with hot water:** This will become your **Hot station** set to "**Shock/42**" for the experiment. Heat the water to 42°C and try to keep it as stable as possible while you heatshock.
- **Incubator:** This will replace the **Incubator** set to "**37**". If you do not have an incubator (biology or egg one, as long as they set to 37°C), you can create one using an online tutorial Search for DIY incubator on our youtube chanel - Youtube.com/aminolabs - or go to this direct link: <https://www.youtube.com/watch?v=LEsv0Qvbczs>

If you are using this solution, our online Udemy course will be an excellent resource for you - in this video series, Dr. Pahara completes an Engineer-it Kit using this alternative set up and shows how to use a light bulb and Tupperware as a DIY incubator. The Engineer-it kit experiment is exactly the same as what you will do with Bag 1 of your Blue-it Kit. <https://udemy.com/handsonbiology/>

Necessary Safety Supplies



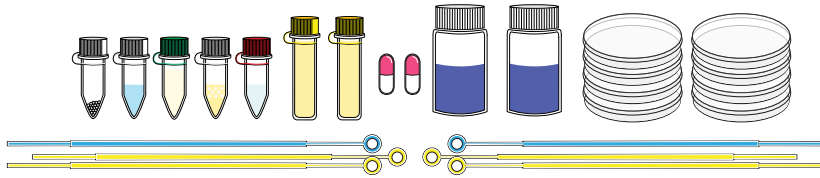
Disposable container 500ml-1L
to hold tubes, loops and other contaminated waste (e.g., yogurt container, plastic cup).

Latex or nitrile gloves
like the ones found at a pharmacy.
at least 10 pairs

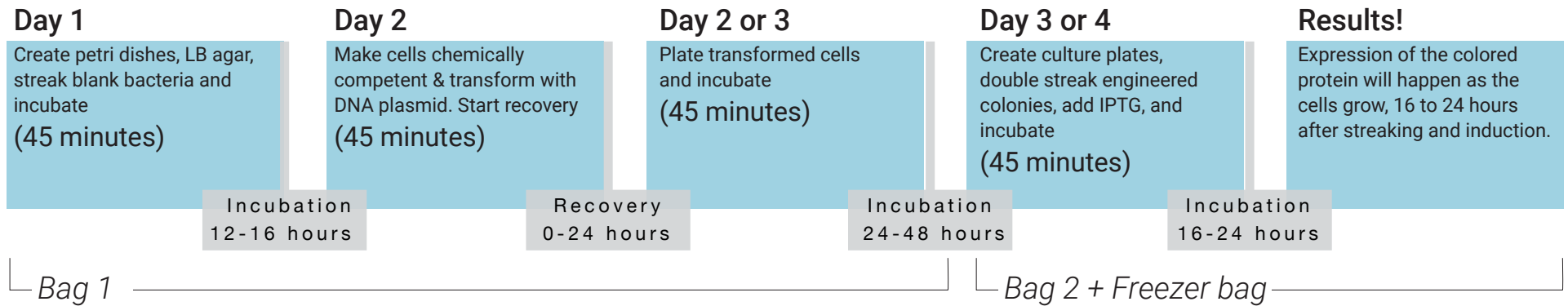
Chlorinated bleach spray
1 regular bottle (or you can mix a 10% solution: 1 part bleach to 9 parts water in a spray bottle)

Bleach ~250 mL
to inactivate all the experiment materials at the end of the experiment.

Timeline



Induce-it Kit



Experiment Protocol



An Experiment Protocol is a scientific way to talk about your instructions for completing the exercises. These will not include any theory or background information on the why of each step. You can find that in the Virtual Bioengineer Simulator, the *Zero to Genetic Engineering Hero* book and the tutorial videos.

In the next pages are detailed, step by step instructions to complete the experiment and genetically engineer your bacteria with DNA. **Please make sure to read all the steps in the section before starting the hands-on manipulation;** some steps will be done in rapid sequences.

Remember that the **Virtual Bioengineer Simulator** available on amino.bio is free to use, and will allow practicing the engineering steps you are about to complete. Also available is a series of **real-time video tutorials** covering the engineering experiment part of your kit. Look for the Engineer-it follow along experiment video!

Find it and more on our youtube channel: youtube.com/c/AminoLabs

Experiment Protocol

1. Creating LB Agar Plates Day 1, 25 minutes

Goal Create non-selective and selective LB agar plates.

Materials from your kit - BAG 1

(1) 50 mL sterile water (1) antibiotic pill (1) Sharpie marker)
(1) LB agar powder (4) 6 cm petri dishes



Prepare

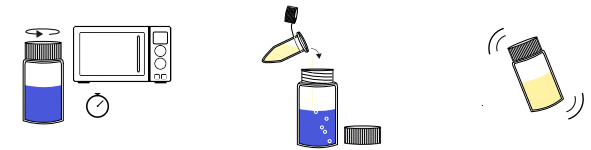
1.1 Label each petri dish with a sharpie-type pen. Label the bottom of the petri dishes (*the side with the star-shaped ring*): **1x** N.S. [your initials] **3x** S. [your initials] of these three, label one **+**, one **-**, and one with an **"e"**



Mix the Agar

1.2 Unscrew the lid from the sterile water bottle and keep it loosely on top of the bottle to prevent any contaminants from entering the water, but allowing air to escape. This will prevent pressure build-ups.

1.3 Place the bottle in the microwave and heat the water **until you see it boil**. You can use 45 seconds as your starting time but you have to see a rolling boil where many bubbles are rising constantly before you continue to the next step. Careful, the bottle will be hot! **!! If the water does not boil, the agar powder will not dissolve and your plates will not solidify !!**



1.4 Add the tube of agar powder to the boiling water. Careful, the water is hot! Some agar powder may "clump" around the lip of the tube due to the water evaporation. This is okay, we have accounted for this possible loss.

1.5 Microwave the water and agar powder in 4 seconds intervals until you see it boil again. Instead of a rolling boil, you will see more of a foam forming above the molten LB agar liquid. **Careful, the liquid will boil over if you microwave in more than 4 sec. increments.** After you see the liquid foaming, swirl to mix for 10 seconds.



Make non-selective (N.S.) Plate

1.6 Pour molten LB agar in the bottom half of your N.S. Petri dish. Enough to fill the petri dish half-full. Swirl the plate to make sure the molten LB agar fills the bottom. If the agar does not cover all the bottom, gently tilt it. Place the lid 3/4 of the way back on so that the agar can cool and dry (solidify).



Make selective (S.) plates

1.6 Add the antibiotic pill to the bottle of agar and gently swirl for a few minutes until the contents of the pill have dissolved. Do not introduce bubbles into the LB agar: don't swirl too vigorously. The gelatin capsule may not fully dissolve. The important thing is that the contents of the capsule do dissolve.

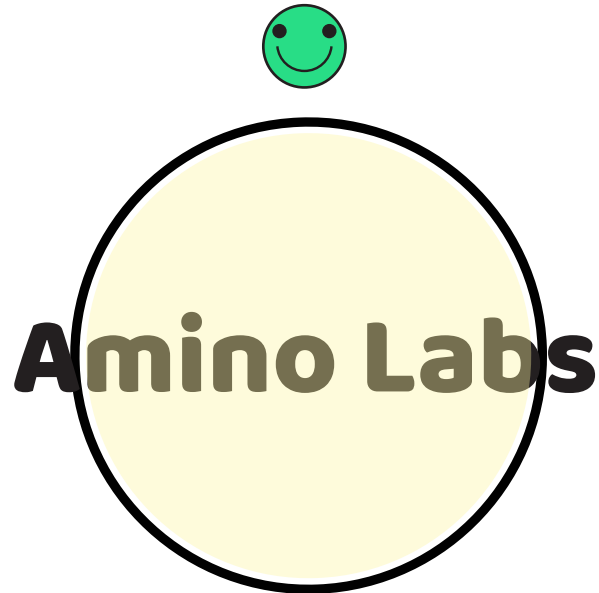
1.7 Once the antibiotic pill is dissolved, pour the molten LB agar into the bottom half of the 3 remaining petri dishes. Place the lids 3/4 of the way back on so that the agar can cool and dry (solidify).



1.8 Let the LB agar harden. The non-selective plate and the "-" selective plate are used in the next step. Put the remaining selective plates in their original ziploc bag for later use, and store in a refrigerator.

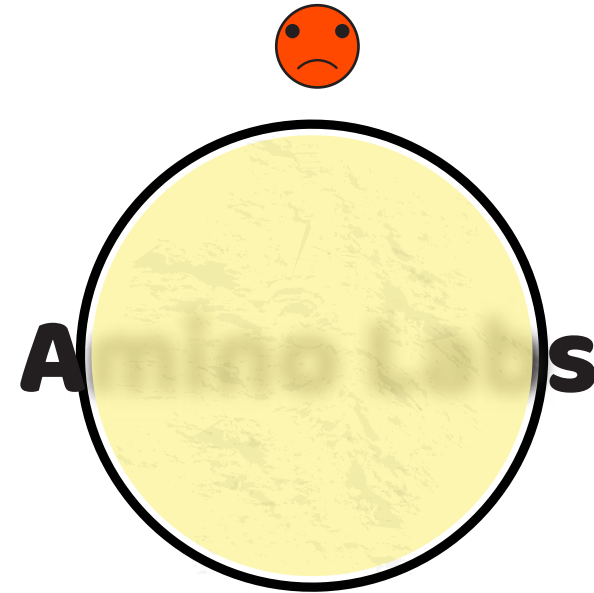
Checkpoint - Agar Plates

Use this guide to check if you are ready to move onto the next step.



A perfect Agar plate is completely clear and solid - if you set it 4" above some image or text, you should be able to read it / see it clearly.

Move on to the next step!



An agar plate that is cloudy and/or bumpy and/or soft is not ideal - if you set your plate 4" above some text or image and cannot see clearly through it, it means you needed more boiling or mixing.

Troubleshooting tip

If your plates do not solidify after 30 minutes it is very likely that the water was not boiled enough to dissolve the agar powder. As a 'hack', you can pour all of the petri dish content back into the water bottle and microwave until you see it boil. Swirl to mix and re-pour your plates.

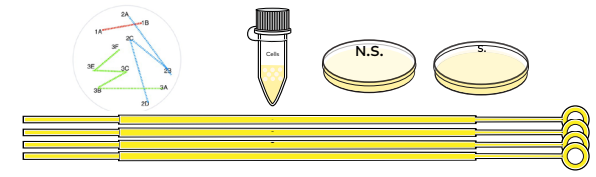
Unfortunately, if the agar does not solidify, this means you need to halt your experiment and complete the troubleshooting guide and follow the instructions at www.amino.bio/troubleshoot

2. Growing Blank Cells Day 1, 20-45 minutes + 16-24 hours wait time

Goal Streak petri dish with provided cells to get fast-growing, well-separated colonies that will be engineered.
Fast growing cells take up DNA the best!

Materials from your kit - BAG 1

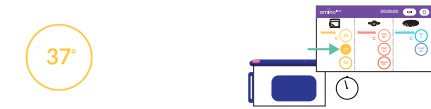
Non-selective & "-" selective plate (4) Yellow Inoculation Loops
(1) Plate streaking stencil (1) Stab of cells (**Not Cells +**)



Prepare

2.0 Inspect your N-S and S- plates for water droplets on the surface. If some are present, take the lid half-off and let the water evaporate before using.

2.1 Turn on your Incubator to 37°C



Streak your Plate

2.2 Place your N.S. Petri dish on top of the zigzag pattern on the Plate streaking stencil. Take one yellow loop and dip it into the stab of cells. Inspect your loop to make sure it appears wet to confirm you have collected cells.

2.3 Trace the line 1 of the stencil with this loop. Discard the Loop in your Inactivation Bag.

2.4 Using a new yellow loop, trace line 2. Discard the loop.

2.5 Using a new yellow loop, trace line 3. Discard the loop. Close your petri dish and set aside. Keep your stencil.

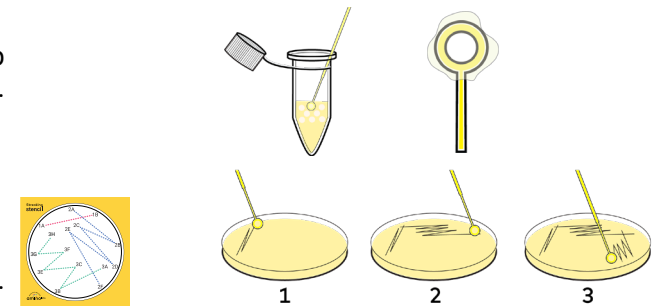
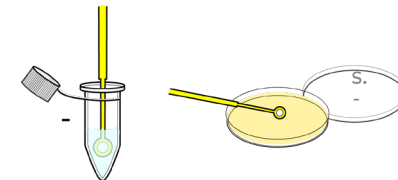


Plate your negative control ("-") plate)

2.6 Take your Selective LB Agar plate labeled "-". Using a single yellow loop, dip into the same tube of cells and spread them across your agar plate in any pattern of your choosing. On this negative control "-" plate, cells should not grow.

2.7 Close your tube of bacteria and place it back in the fridge in a zip-lock bag if you want to keep them, discard them in the Inactivation bag if you do not. You no longer need them for this experiment.



Incubate Overnight

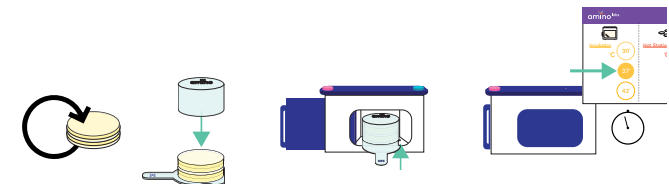
2.8 Flip your streaked N.S. & S. "-" plates upside down and stack them in a pile in an incubator set to 37°C. If you are using the DNA Playground, put your stack of plates on top of the incubator paddle, set the incubator humidity chamber on top and slide in the incubator. Close the incubator door and lock it using the incubator key.

If you are using The DNA Playground or a commercially-made incubator at 37°C:

It is important to do the next steps in 12 to 24 hours so that your cells will be in their optimal growth phase.

If you are using a homemade incubator:

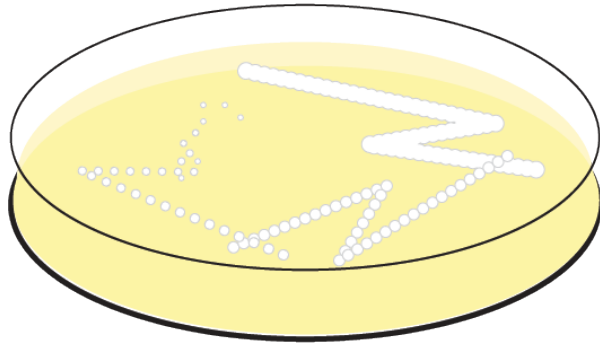
It must incubate between 35°C and 37°C. If the cells are below 37°C, it will take longer for them to incubate - you may have to wait up to 48 hrs until you see small colonies on your plate.



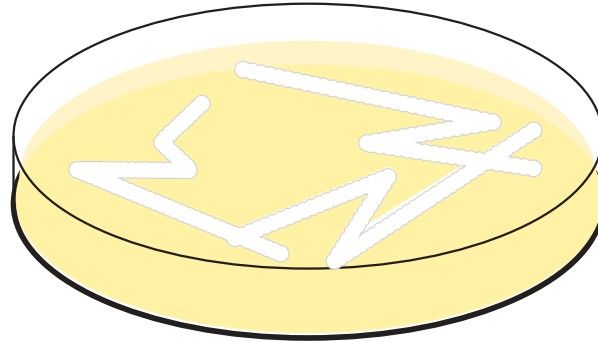
Teacher Tip! If you are in a classroom setting and your class is 24 hours-30 hours apart or more, you can initially grow your cells at 30 C overnight and then increase the temperature to 37 the day of making competent cells.

Checkpoint - Non-selective plate & blank cells

Use this guide to check if you are ready to move onto the next step.

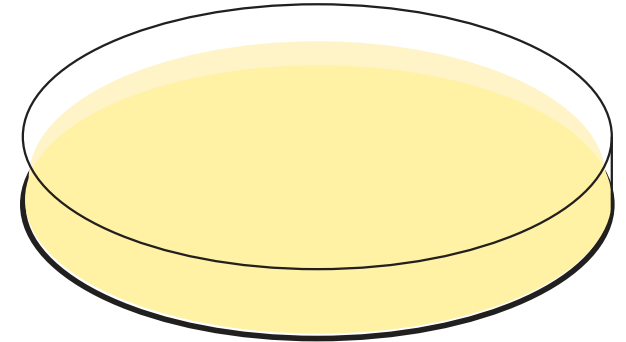


A perfect N.S. plate has lots of small individual white colonies (dots) of fast-growing bacteria after incubation. Proceed to the next page.



An okay N.S. plate has bacteria growth but few or no individual colonies (dots) after incubation. Continue with the experiment by selecting the colonies that are on the edges of the dense lines of bacteria.

Note that your results may not be as good as when you start with a perfect plate. Don't worry though, practice makes perfect, and our Success Guarantee means you will have the chance to be successful if this try doesn't work.

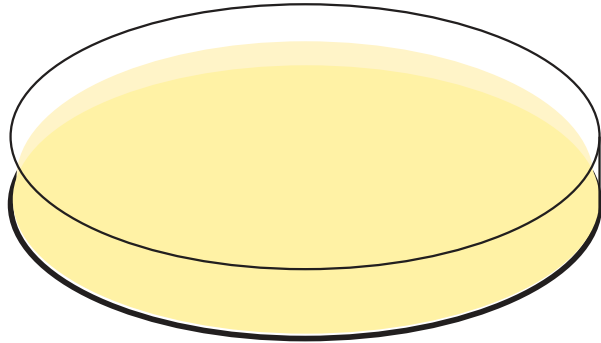


If you see no growth on your N.S. plate:

1. If your incubator was not at 37°C or is homemade, incubate for another 24 hrs.
2. If you are certain you incubated at 37°C, or incubated for 48 hrs and still have no colonies, you might not have had cells on your loop when you streaked. Repeat Step 2: Growing Blank cells on this plate.
3. If you still have no colonies after repeating Step 2, complete the guide at www.amino.bio/troubleshoot

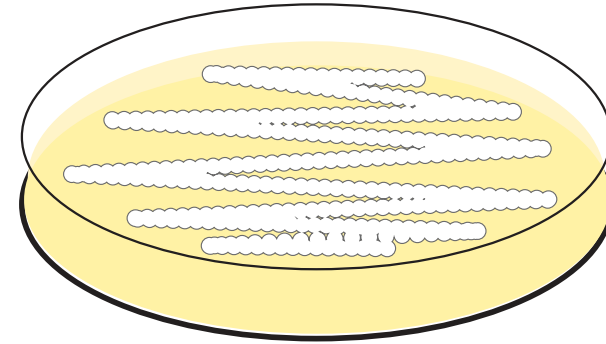
Checkpoint - Selective plate and “-” control

Use this guide to check if you are ready to move onto the next step.



A perfect “-” plate has no growth on it. This is because the antibiotics do not allow the blank cells to grow. This means that you made your selective plates properly.

Move on to the next step.



If you see growth on your “-” plate, this suggests that there is not enough antibiotics.

Continue the exercise with your other selective plates. Note that your results may not be as good as if you had the right amount of antibiotics. Don't worry though, practice makes perfect, and our Success Guarantee means you will have the chance to be successful if this try doesn't work.

3. Making Chemically Competent Cells Day 2, 10-15 minutes

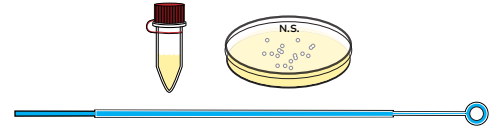
Goal Pick small colonies, and suspend the bacteria in cold transformation buffer, enabling bacteria to better take up DNA.

Materials from your kit

(1) Streaked N.S. Plate

(1) Blue Loop

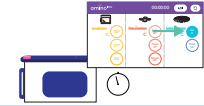
(1) T. Buffer Tube



Prepare

3.1 Make sure you have completed the Non-Selective Plate & Blank cell + Selective Plate and "-" control Checkpoints. You can discard the S. "-" plate in your inactivation bag.

3.2 Turn on the cold station to the "Ice" (4°C) setting on your Amino Lab's Minilab™ or get your Ice bucket.

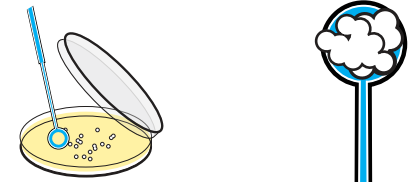


Mix the cells and T. Buffer

3.3 Take your T. Buffer tube and make sure all the liquid is in the bottom of the tube by briefly putting it in your microcentrifuge (don't forget to put a balance tube!), or by tapping it gently on a surface. You should have 5mm of liquid in the bottom of the tube. Set it on your cold station set to "ice" or in real crushed ice for 2 minutes so that it cools down.

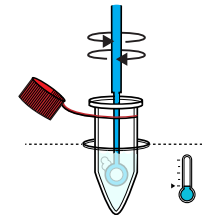
3.4 Take a blue inoculating loop and gently scrape it over small, well-separated colonies on your N.S. plate. Colonies that are ~1mm in diameter work the best. You want to collect ~10 or 20 of these colonies on your loop, enough so that you can see that the center of the loop is full.

Tip: A colony is one of the white "dots" or "mounds" you see on your N.S. LB agar. The separated colonies are those that look like individual dots, not streaks or solid lines of white.



3.5 Immerse the loop with the bacteria in the cold Transformation Buffer without touching the sides of the tubes.

3.6 While keeping the T. Buffer tube on the cold station, twist the loop like a blender (or like you are trying to start a fire) to mix and suspend the cells in the liquid while keeping the tube in the cold station. You don't have to twist too vigorously as this could break off the loop.

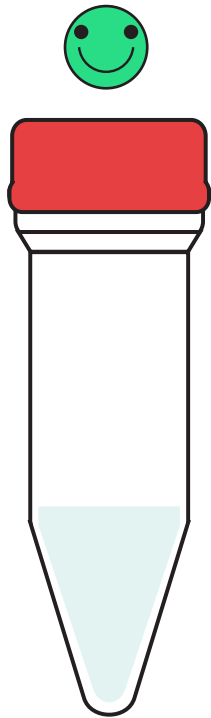


When successful, the solution should be slightly cloudy, and there should be no clumps floating in the solution. You may have to mix vigorously for up to 45 seconds. If you see clumps, keep blending. You can lift the tube out of the cold station to see whether the T. Buffer is cloudy and the clumps are gone, but replace it quickly into the cold station to keep it cool. Have a look on the next page to see what your tube should look like. When you have successfully mixed in your cells to the T. Buffer, discard the loop in the inactivation bag.

Without delay, move on to the next step to add your DNA! **Ideally you should add your DNA less than 1 minute after making your competent cells.**

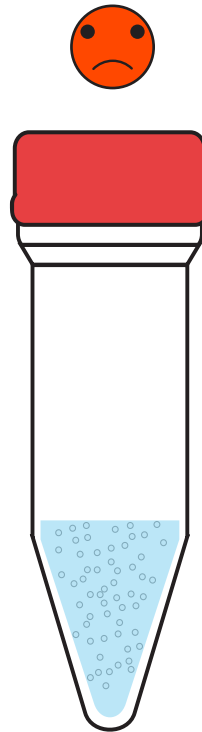
Checkpoint - Competent cells

Use this guide to check if you are ready to move onto the next step.

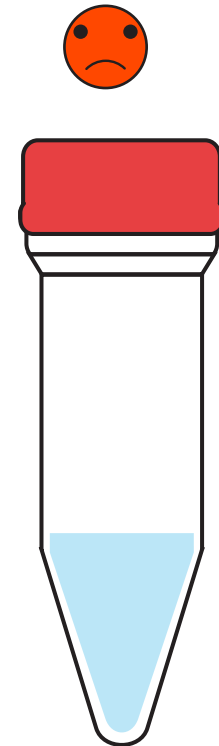


Cloudy liquid - Perfect!
The cells can now take up DNA.

Move on to the next step.



You can still see clumps... Keep
mixing until liquid is cloudy like
the image on the left.



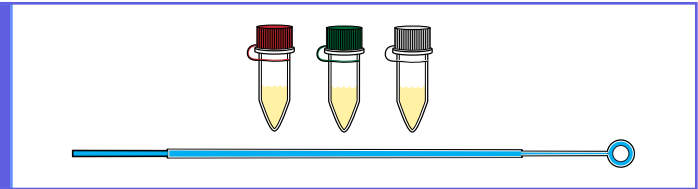
Clear liquid? Add more cells and
mix until the liquid is cloudy like
the image on the left.

4. Transformation Day 2, 45-60 minutes

Goal Introduce a DNA plasmid into competent bacteria and recover the cells.

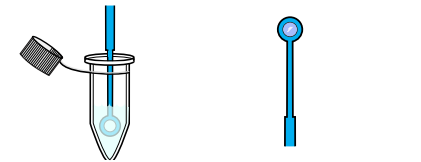
Materials from your kit - BAG 1

- (1) DNA plasmid tube
- (1) Blue Loop
- (1) Competent Cells {from prior step}
- (1) Recovery Media tube



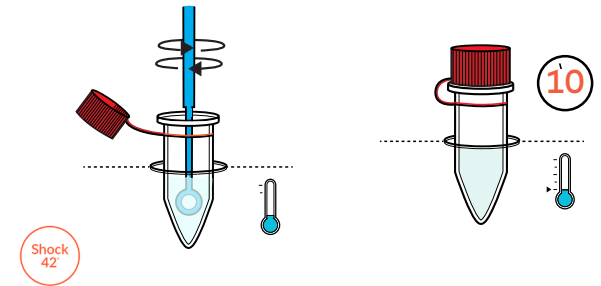
Take the DNA

4.1 Tap your DNA tube on the table or briefly microcentrifuge to make sure most of the DNA is at the bottom. Dip a Blue inoculating loop into the DNA tube and twist a few times. When you pull the loop out of the DNA tube, the hole of the loop should have liquid in it. Each tiny droplet of the liquid contains thousands of DNA plasmids, small circles of DNA programs that you will use to engineer your cells. Make sure you can see liquid in the loop!



Mix the competent cells & the DNA

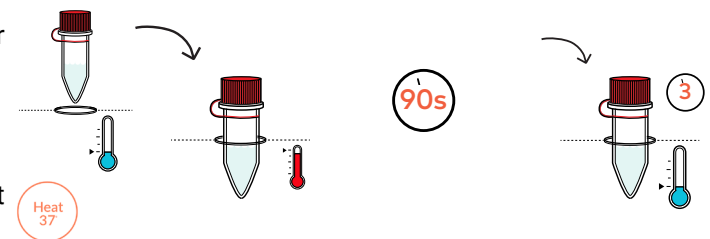
4.2 Slowly dip and spin the inoculating loop containing DNA into the competent cells you made in the previous step. Stir/swirl for 5 seconds to fully mix. Do this while the tube is on Ice / Cold Station. Discard the loop. * Do not reuse the inoculating loop! *



4.3 You will incubate your tube for 5 minutes on Ice / Cold Station. While this is happening, turn on "Shock 42°C" on your Minilab or set your water bath to 42°C. You can close and put the DNA tube back in the fridge. If stored in a fridge, and you have not accidentally contaminated the DNA, it should last a month or more and can be used again in a future engineering experiment.

HeatShock

4.4 It is now time to get the DNA into the cells. Heat shock your cells, T. Buffer, and DNA solution by moving your tube to the Hot Station (42°C) / Water Bath (42°C) for 90 seconds

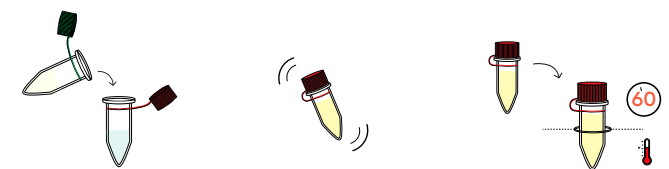


4.5 After 90 seconds, immediately place your tube back on Ice / Cold Station for 2 minutes.

4.6 The next step is recovery which will happen at 37°C. Turn on your Minilab Hot station to Heat 37°C or adjust your water bath temperature to 37°C.

Recovery

4.7 Tap the tube of Recovery Media on the table to ensure that most of the liquid is at the bottom of the tube and pour into your tube of cells + T. Buffer + DNA solution. Mix gently by inverting 10 times. Some liquid will stay in the recovery tube. That is acceptable.



4.8 Place your tube of solution in the Hot station set to "Heat 37°C" or your water bath for 30-45 minutes to allow the cells to recover and start expressing their antibiotic resistance proteins. Mix the cells by inverting them every 15 minutes. * Make sure the liquid is in the bottom of the tube when you place it in the tube heaters! * See www.amino.bio/whip-it to learn the Whip-it method for moving liquid inside a tube.

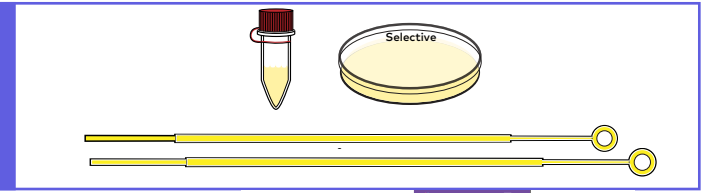
After the initial 30 minutes of recovery, you can wait up to 24 hours before moving onto the next step.

5. Plating Cells Day 2 or 3, 10-15 minutes + 24-72 hours wait

Goal Spread your transformed bacteria ("e"), positive control cells ("+") on selective LB agar plates

Materials from your kit - **BAG 1**

- | | |
|-----------------------------------|---|
| (1) Selective Plate {from step 1} | (1) Transformed Cells {from prior step} |
| (2) Yellow Loop | (1) "+ Cell" Positive Control tube |



Prepare

5.1 Turn on your Incubator at 37°C.



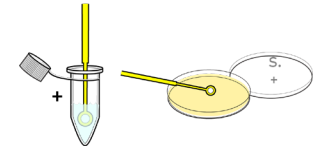
Plate your transformed cells ("e" plate)

5.2 On the Selective LB Agar plate labeled "e", pour 1/2 of your Transformed cells solution from the previous step. Spread with a yellow inoculation loop so that the liquid covers the entire plate. Be gentle as not to puncture the agar! Discard the Loop. Leave the lid of the plate partially off to allow for evaporation. You will be ready to incubate when there is no pooling of liquid on the plate. Close the tube with your transformed cells and keep them just in case you do not get colonies since you can always pour the remaining recovered cells onto the selective plate and incubate them after your step 6. incubation.



Plate your positive control ("+" plate)

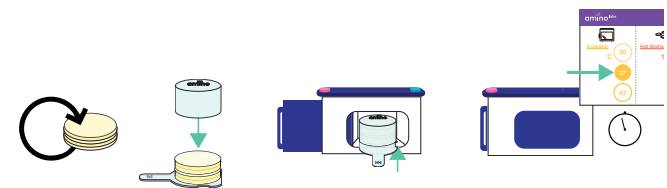
5.3 Get the stab of "+" cells" and your Selective LB Agar plate labeled "+". Using a single yellow loop, dip into the + cells stab and spread them across your agar plate in any pattern of your choosing.



Incubate

5.4 Once there is no liquid pooling on any of the plates, place the plates upside down in the Incubator. If you are using a Minilab incubator, make sure to use the humidity chamber as well. Inverting the plates ensures that water vapor in the plates keeps the LB agar surface moist.

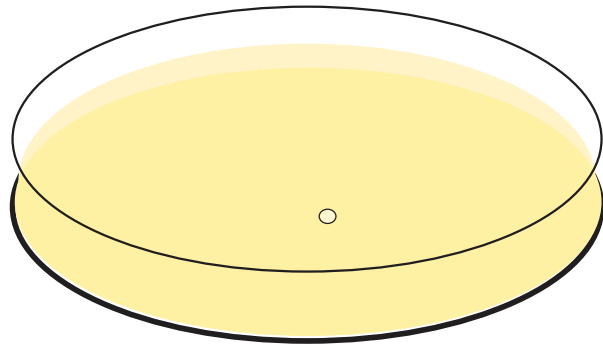
5.5 Incubate for 24-48 hours until you see one or more colonies.



The cells will grow in colonies and start producing their new DNA program in the next hours. The DNA program can take up to 48 hours to develop. Keep an eye out, and your camera ready to document!

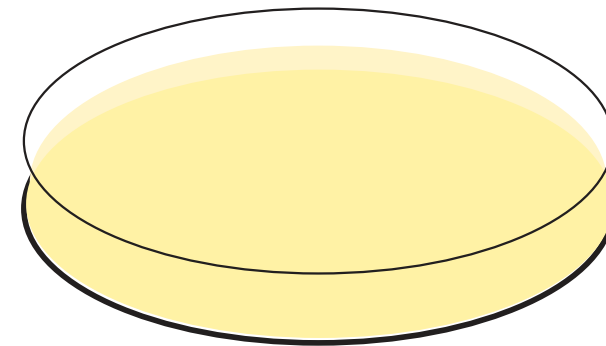
Checkpoint - Did your engineered cells grow?

Use this guide to check if you are ready to move onto the next step.



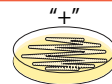
A perfect “e” plate has one or more colonies on it. Remember, even a single colony is a success. One colony is all you need to move onto the next step. Congratulations!

Move on to the next step.



If you see no growth on your “e” plate, this means that your engineering may not have worked, or that you haven’t incubated long enough.

If you cannot see any growing cells after 48 hours, you can pour the rest of your engineered, recovered cells onto the “e” plate and continue incubator for another 24 hours. If you still don’t see any growth after this, your experiment may have failed. Don’t be discouraged. In science, failure is a chance to learn more. Complete the troubleshooting guide at amino.bio/troubleshoot to claim your free kit to try again, as part of our *Success Guarantee*.



You can continue your experiment with a colony from your “+” plate if the + cells grew. Try to select a single, isolated colony if you can.

6. Create Selective LB Agar Plates Day 3 or 4, 25 min.

Goal Create a new batch of selective LB agar plates to amplify (culture) your engineered cells.

Materials from your kit - BAG 2

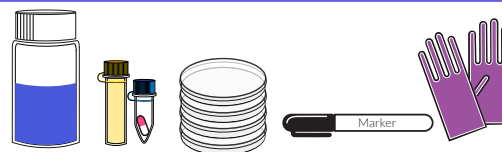
(1) 50 mL sterile water

(1) LB agar powder

(1) antibiotic pill

(4) 6 cm petri dishes

(1) Sharpie marker



Prepare

1.1 Using a sharpie-type pen, label the bottom of the petri dishes as follows: **4x S.** (for selective) + Add [your initials] if doing this in groups with multiple kits. (*The bottom is the side with little tabs*)



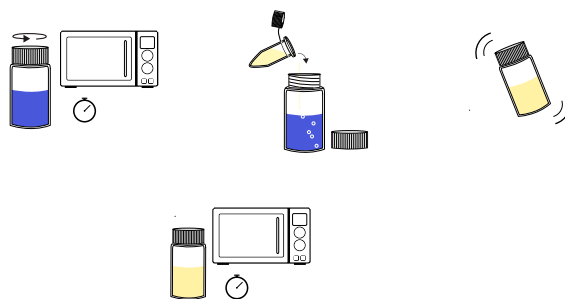
Mix the Agar

1.2 Unscrew the lid from the sterile water bottle and keep it loosely on top of the bottle to prevent any contaminants from entering the water, but allowing air to escape. This will prevent pressure build-ups.

1.3 Place the bottle in the microwave and heat the water **until you see it boil**. You should see a rolling boil where many bubbles are rising constantly. Careful, the bottle will be hot!

1.4 Add the tube of Agar powder to the boiling water. Careful, the water is hot! Some agar powder may “clump” around the lip of the agar tube. This is due to the water evaporation coming into contact with the agar powder as you pour it in. This is okay, we have accounted for this loss of powder.

1.5 Microwave the water and agar powder in 4 seconds intervals until you see it boil again. Instead of a rolling boil, you will see more of a foam forming above the molten LB agar liquid. **Careful, the liquid will boil over if you microwave in more than 4 sec. increments.** After you see the liquid foaming, swirl to mix for 10 seconds.



Note that you will not be making a non-selective plate. All four plates will be selective agar for this part of the experiment..

Make selective (S.) plates

1.6 Add the antibiotic pill to the bottle of agar and gently swirl for a few minutes until the contents of the pill have dissolved. Do not introduce bubbles into the LB agar, which means don't swirl too vigorously. The gelatin capsule of the pill may not fully dissolve. The important thing is that the contents of the capsule do dissolve.

1.7 Once the antibiotic pill is dissolved, pour the molten LB agar into the 4 petri dishes. Place the lids back on.

1.8 Let the LB agar harden. You will use all 4 plates in the next step.



7. Amplify (Culture) engineered cells Day 3 or 4, 10minutes

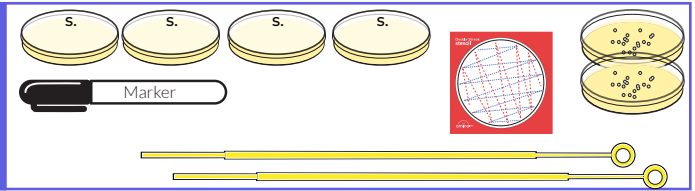
Goal Grow a large quantity of cells on plates to induce after growth.

Materials from your kit - BAG 2

Agar plates from previous step
Your engineered cells

Positive control cells
(2) Yellow Loops

Double streak stencil
Marker

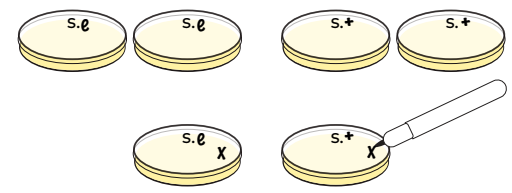


Prepare

7.1 Turn on your incubator to 37°C.



7.2 Identify two of your selective plates with “e cells” for your engineered cells and the other two with “+ cells” for your positive control cells. Remember to always identify plates on the bottom of the plates!



7.3 On one “e cells” plate and one “+ cells” plate, make a small x where you will add the inducer. We recommend the side of the plate since it makes it easier to see the difference between induced and non-induced cells.

Streak two petri dishes with engineered cells

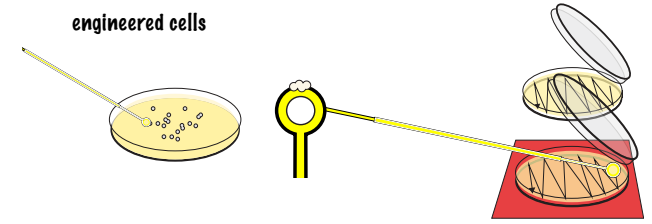
7.4 Take your petri dish of engineered cells from the previous incubation. Place one of your new “e cells” petri dish on top of the double zigzag pattern stencil.

7.5 Using one yellow loop, pick one or more colonies of engineered cells from the incubated plate. You pick a colony by touching the end of the loop to it, gently rubbing it.

7.6 With your picked colonies on your loop, trace one of the zig zag across the new “e cells” selective agar plate.

7.7 Using the same yellow loop, trace the second zigzag, which is at 90° of the first. This will ensure you will have lots of cells growing across your plate. Keep your yellow loop in your hand.

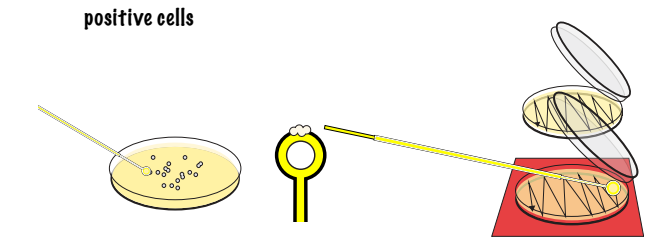
7.8 Take your second new “e cells” petri dish, and place it on top of the stencil. With the same yellow loop, repeat the steps above to streak the same colonies on your new plate. Discard your loop and close your plates.



Streak two petri dishes with positive cells

7.9 Take your petri dish of positive cells from the previous incubation. Place one of your new “+ cells” petri dish on top of the double zigzag pattern stencil.

7.10 Using one yellow loop, pick one or more colonies of positive + cells from the incubated plate. Streak these colonies on your two “+ cells” plates by repeating the steps above.

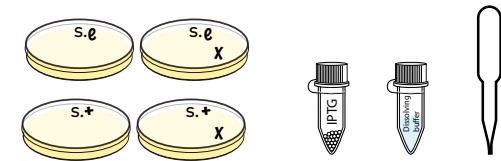


8. Induce your cells & see your results! Day 3 or 4, 10 mins + 16-24 hrs wait

Goal Induce your incubated cells and see the colored protein expressed!

Materials from your kit

Your 4 plates from the previous step (1) IPTG tube
(1) Pipet (1) Dissolving buffer



Prepare your IPTG

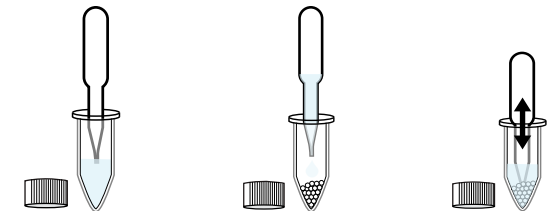
8.1 If you have a microcentrifuge, you can place your IPTG tube in your centrifuge and spin at max speed for 10-30 seconds. This will ensure that all the IPTG powder is at the bottom. If you do not have a microcentrifuge, 'whip' the tube so that all the powder is at the bottom of the tube. Use the whip-it technique: amino.bio/whip-it.



Dissolve your IPTG

8.2 Take the tube of Dissolving buffer and pipet its entire content into the IPTG tube. To mix the buffer and IPTG powder, use your pipet and suck the content up and down 10 times to mix together.

You will be re-using the pipet in the next steps: If you have a DNA Playground or tube rack, you can set your tube of dissolved IPTG with the pipet resting inside on one of the stations set to 'OFF'. If not, close the tube and set your pipet on a clean surface like a paper towel.

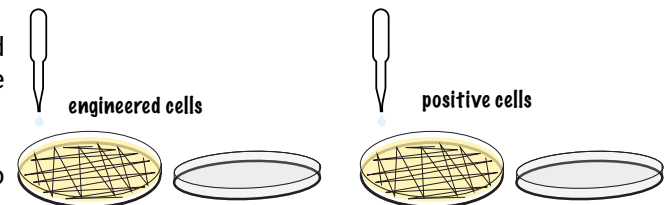


Induce your cells

8.3 Using the plastic pipet, add one drop of dissolved inducer on the surface of the agar, on top of your streaked cells, where you marked the x. Only add the inducer to the two plates with the "x". The other two plates will be your negative controls.

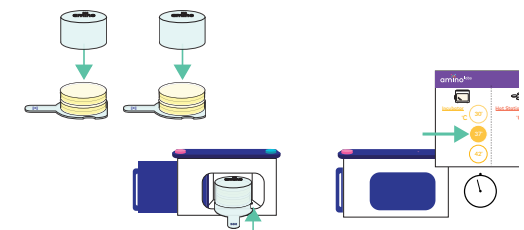
8.4 Once you've added the drop of inducer to one "e cells" and one "+ cells" plate, leave the lid off the plates to let the inducer liquid diffuse into the agar and the liquid evaporate for ~5 minutes.

8.5 Incubate your 4 plates at 37°C for the next 16 to 24 hours. The plates with the IPTG should be incubated right-side up, whereas you can incubate the other 2 plates upside down as usual.



See your results

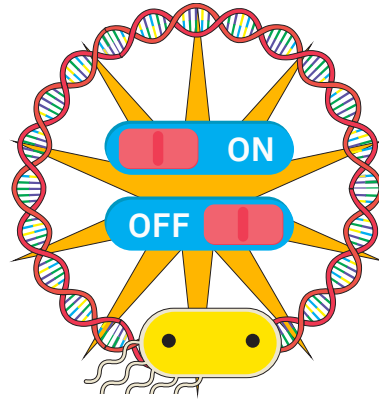
The inducer will be at its highest concentration at the point where you've dropped it. However, it will dissolve outwards in a circular pattern through the LB agar. The higher the inducer concentration, the more the gene will be induced, activating the color production in the bacteria! The induction of the genetic switch relies both on time and concentration of IPTG. Check back on your plates after 16 hours and onwards. You can take photos to see the color-change happen over time. You've manually turned on a genetic switch! Congratulations!



Note

If you cannot see any color change 24 hours after adding the inducer, you may not have fully dissolved the IPTG powder. Check the IPTG tube for remaining powder. You can try to re-dissolve it with distilled water and adding it to the plates. If nothing happens, your experiment might have failed. Don't be discouraged. In science especially, failure is a chance to learn more. Complete the troubleshooting guide at www.amino.bio/troubleshoot

Congratulations!



Using your Induce-it Kit™, you inserted a DNA Program inside single-celled organisms, bacteria, and had the bacteria execute that DNA Program to produce something for you, which you then controlled the expression of using a chemical. Just like scientist and industries do every day inside their large laboratories! To learn more about genetic switches, IPTG, and inducible plasmids, look at Chapter 7 of the *Zero to Genetic Engineering Hero* book. We hope you enjoyed the experience and will continue to experiment in the field of genetic engineering.

Show off your results with your friends, our community and us too on Twitter, Instagram, Facebook, and youtube (@aminobiolab)

For now, let's make sure you dispose of and store your remaining material correctly.

Storage, Disposal, Clean Up

After you see your results, all experiment Petri dishes, tubes of cells and loops should be in the inactivation bag in your discard container. Disposing of experiment materials is an integral part of the experiment. **Always wear gloves for cleanup!**

A. Preserving Petri dishes: If you want to preserve the living paintings or experiment results in Petri dishes instead of disposing of them, use one of our Keep-it kits. This will help you maintain the petri dish by pouring a special resin on top. If you do not have Keep-it Kits on hand but will be getting one soon, keep the Petri dishes you want to preserve in a ziploc bag in a cool area and out of sunlight in the meantime. You can refrigerate it to keep it “fresh” for up to a month.

B. Reusable materials: If you have DNA in your kit, it can last up to 6 months when stored in a refrigerator. If you wish to keep it, store it in a ziploc bag inside a sealed plastic container in a refrigerator away from food items. If you do not wish to keep it, add to an inactivation bag. Make sure the lids are separate from the tubes so that the inactivating liquid can get inside. If you see any mold or unknown bacteria growing on any material at any point, immediately inactivate them by using a solution of bleach water. Follow the inactivation instructions below. If you are out of inactivation bags, use a sturdy ziploc type bag or disposable container with a lid. Always wear gloves when handling experiment materials and cleaners!

C. Unused ingredients: If you did not use all the agar Petri dishes you poured, store these for later use. Store them in their ziploc bag within a sealed container in the refrigerator for up to a few months. Keep them away from food items. If you see any mold or unknown bacteria growing inside, then you should always immediately inactivate the Petri dishes.

D. Inactivation: Make sure all bacteria, agar, tubes, loops, paintbrushes, Petri dishes, contaminated gloves, and other non-paper material you are not keeping are in the inactivation bag. Remember that any paper packaging like loop wrappers, plastic bags, and gloves that have not touched bacteria go in the regular garbage or recycling.

Make sure all the tubes have their lids off once in the inactivation bag and add a solution of 1 part bleach to 4 to 6 parts water to the inactivation bag. Close the bag and let sit for 24 to 48 hours before discarding the liquid in the toilet and the solids & bags in the garbage. Step-by-step instructions are on the inactivation bag and in an Inactivation video on youtube; [youtube.com/c/AminoLabs](https://www.youtube.com/c/AminoLabs).

Spray some chlorinated bleach cleaner in the discard container once emptied if it has become contaminated by experiment materials. Let it sit for an hour before wiping down. You can wait to wipe it down until you empty out your inactivation bags the next day.

E. Clean your workspace: Use a chlorinated spray cleaner, wipes, or a solution of 1 part chlorinated bleach to 9 parts water to wipe down your work area and equipment. You can wipe down the minilabs with this solution and follow it with an eyeglass or window cleaner to remove the inevitable streaking from the bleach cleaner. Never use rubbing alcohol (isopropyl alcohol) on the DNA Playgrounds.

Glossary

Agar: is a Jello-like substance that serves as a growth media for bacteria. It is mixed with our bacteria's favorite food: Lysogeny broth (LB). LB is made up of yeast, vitamins, and minerals. LB can also be found in liquid-form.

Antibiotics: When you transform bacteria, they will become resistant to a type of antibiotics no longer used in hospitals. This antibiotic will be mixed in with the agar and LB so that, as you incubate your culture, only transformed bacteria will grow. This is called a "selection marker".

Autoclave: An autoclave is a machine used to carry out industrial and scientific processes requiring elevated temperature and pressure in relation to ambient pressure/temperature. In life science, autoclaves are used to sterilize equipment and supplies by subjecting them to pressurized saturated steam at high temperatures (around 250 °F) for several minutes, up to an hour. Autoclaves are similar to some baby bottle sterilizers which you might be familiar with.

Buffers: Buffers are saline solutions that help, in this case, open up the cell membranes so that they may take up new DNA.

Cells: Cells are tiny, living units that function like mini-factories. Bacteria are single-celled organisms (unicellular) microorganisms. They are different from plant and animal cells because they don't have a distinct, membrane-enclosed nucleus containing genetic material. Instead, their DNA floats in a tangle inside the cell. Individual bacteria can only be seen with a microscope, but they reproduce so rapidly that they often form colonies that we can see. Bacteria reproduce when one cell splits into two cells through a process called binary fission. Fission occurs rapidly, in as little as 20 minutes.

Competent Cells: Since DNA is a very hydrophilic molecule, it won't normally pass through a bacterial cell's membrane. In order to make bacteria take in the DNA plasmid, the cells must first be made "competent" to take up DNA. This is done by creating small holes in the bacterial cells by suspending them in a solution with a high concentration of calcium (the transformation buffer). DNA can then be forced into the cells by incubating the cells and the DNA together on ice, placing them briefly at 42°C (heat shock), and then putting them back on ice. This causes the bacteria to take in the DNA and is called "Transformation".

DNA: The DNA is the set of instructions that tell the cell how to function like a computer program tells your computer what to do. DNA stands for **D**eoxyribo**n**ucleic acid.

DNA plasmid: A plasmid is a small circular piece of DNA (about 2,000 to 10,000 base pairs) that contains essential genetic information for the growth of bacteria. Bacteria share vital information by passing it among themselves in the form of genes in plasmids. By inserting a new plasmid in our bacteria, we can get them to produce things for us, can get them to produce things for us, like mini-factories. In this case, we have a plasmid that encodes for the creation of colorful pigments.

Genome: a genome is all genetic material of an organism. It consists of DNA. Learn more about genomes in the *What is DNA?* simulator on amino.bio

Heatshock: is when the cells are moved from ice-cold to warm temperature, typically 42°C, to take in DNA plasmids more efficiently.

Inoculation: is when you introduce bacteria into a medium suitable for its growth.

Inoculating Loops: are used to transfer liquids, cells, and DNA from one vial to the next instead of tradi-

tional lab pipettes, making your job easier, and less costly. They come in different pre-calibrated sizes, so you do not need to worry about minuscule liquid volumes. They are also used to spread bacteria on an agar surface without puncturing the soft agar.

Non-Selective: A non-selective plate means that any cells/bacteria put on this agar will grow as long as they are oxygen-loving organisms (called aerobic bacteria).

Plates (or Petri dish): A petri dish is a small plastic container used to culture (grow) bacteria in a controlled environment.

Recovery period: is the period after the heat shock in which the cells develop their antibiotics resistance and start dividing.

Selective: A selective plate means it contains antibiotics. When you insert a new DNA program into cells to make them create pigments, or anything else, you also put a “selective marker” (antibiotics resistance) inside the code. This means that only the cells that have taken up the new program will be able to grow on a plate that has the antibiotics mixed in. You only get the cells you transformed!

Transformation: See competent cells.

Troubleshooting

Here are some possible common issues:

Your agar is too wet/ doesn't solidify:

When done correctly, the agar will be the consistency of Jell-O. If it is not:

1. You likely did not heat (boil) the water before, or after adding the LB agar powder
2. You might not have added all the powder from the tube, resulting in too much water vs. LB agar powder.
3. You may not have fully dissolved the powder, meaning it cannot turn into a gel and will look cloudy. You can practice by making Jell-O! Next time heat and swirl longer to ensure the powder is fully dissolved.

You don't have any colonies and its been 24+ hours:

Don't worry, every scientist has experienced this, and it can take some practice before success.

1. Double check that your incubator is on at 37°C. If it is not, or if you are growing at room temperature, then it can take much longer to see the bacteria colonies. Keep waiting!

If you kept the second half of your recovered cells, you can pour them on your plate after 48 hours of seeing no engineered colonies grow and keep incubating.

2. You may need to try again to hone your skills. See our YouTube videos for tips and tricks on how to improve your chances of success.

Your colonies of bacteria grew, but they are the wrong color or there is mold on your petri dish:

Danger! If at the end of, or during, the incubation period your resulting bacteria/plate is: a)not the right color; b)is black when it shouldn't be, this is a sign that your culture is NOT YOUR ENGINEERED BACTERIA. You should immediately inactivate it and clean your space and unit.

To inactivate it, either add it to the inactivation bag or pour 100% chlorinated bleach into the dish, put the lid on and let it sit for 24 hours before throwing it out: The strong oxidizing environment degrades any living organisms. After 24 hours, if there are still organisms present add more concentrated bleach until it is almost full, and let stand for a further 24 hours.

There may be mold in your environment. We recommend, getting a small air purifier with a HEPA filter for the room.

Always be aware that concentrated bleach is a strong oxidizing agent and if poured on the skin can cause irritation, and on clothes remove color. Follow the safety and handling protocol on the manufacturer's label.

Find an interactive troubleshooter online at

amino.bio/troubleshoot. We recommend using it for tips, tricks and to claim your Success Guarantee Kit if you need of one.

If anything else causes you issues, please contact us : help@amino.bio

More Information



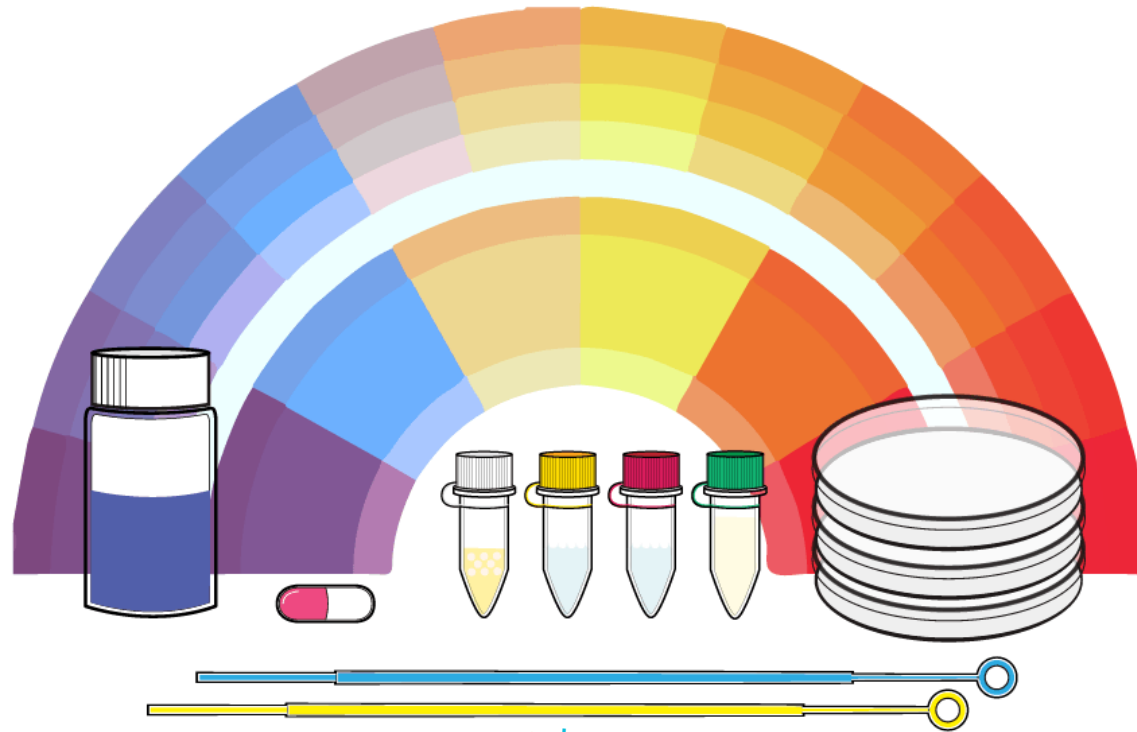
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