IBI SCIENTIFIC

For Research Use Only

INSTRUCTION MANUAL

MAXI SELECT-D TE, G-25

IB06030 – 50/pack

Product Number	Size
IB06030	50

Physical Specifications

Package Quantity	50/pack
Maximum Sample Volume	100 µl
Centrifuge Type Required	Table Top/Swing Bucket Rotor
Recoveries	Pd(N)12 >60% Pd(N)19-24 >80% Large DNA >90%
Retention	Retention Unincorporated NTPs >90%

Internal Specifications

NOTE: Forceps may be needed to remove the column and collection tubes from the swinging bucket or carrier centrifuge tubes utilized.
Each DNASE- and RNASE-free column is prepackaged with Sephadex G-25 in sterile TE buffer (10mM Tris HCL, 1mM EDTA, pH 8.0). Two nuclease-free collection tubes (autoclavable) are supplied for each column. Proper precautions should be taken to avoid contamination of the column, column contents, collection tubes and samples with exogenous RNASE. The supplied columns and collection tubes are sterile and nuclease-free.
The optimal sample loading volume is 50µl (100µl Max.) with a maximum of 100µg of nucleic acid per column. Sample should not be viscous prior to loading.
For best results use a centrifuge with a swinging bucket or horizontal rotor, clinical tabletop centrifuges are also suitable.

Recommended Use

The MAXI SELECT-D TE, G-25 spin columns are intended for use in desalting, recovering DNA fragment (>12mer), and removing unincorporated radiolabeled deoxynucleoside triphosphates (dNTPs) from small volume 5′ end-labeling reactions and fill-in labeling reactions utilizing a DNA polymerase. After brief centrifugation, the purified nucleic acid is recovered from the column without significant change in volume.

Quality Assurance

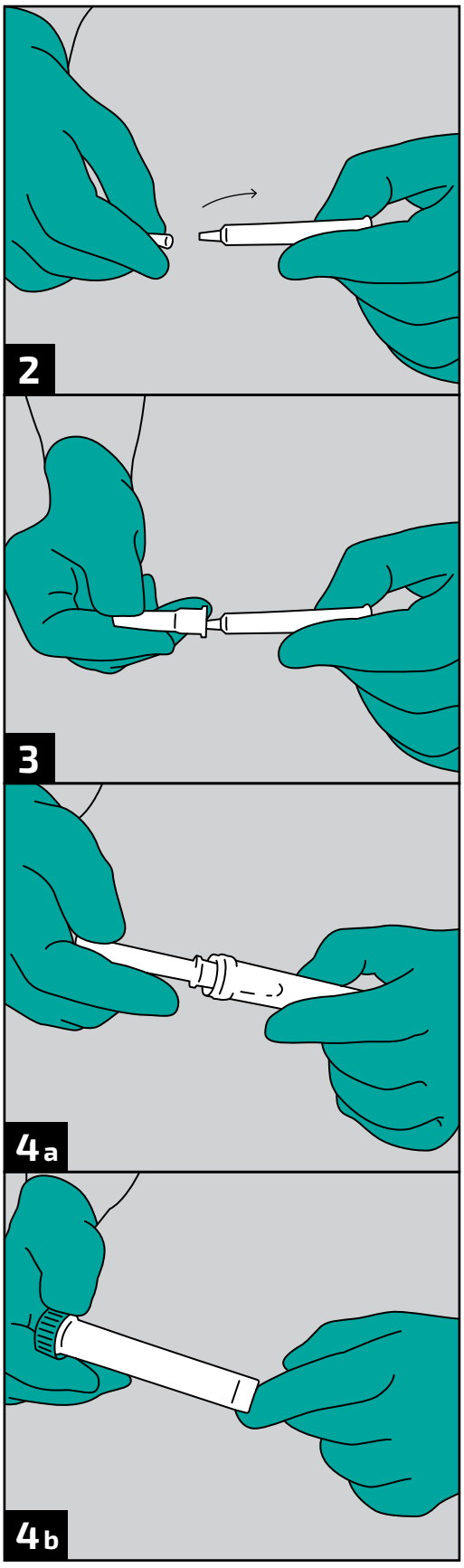
Each lot of MAXI SELECT-D TE, G-25 have been tested for recoveries and retention. IBI Spin Columns have been found to meet or exceed the above specifications. Each lot is also tested for sterility, and the absence of detectable DNASE and RNASE.

Storage

Columns should be stored at 2–8°C and are stable for a period of at least one year.
NOTE: DO NOT FREEZE COLUMNS.

Protocol

NOTE: Please read the entire protocol before using spin columns



Steps 2, 3 and 4 are illustrated

1) Invert the column several times to suspend the gel.

2) Remove the small white cap from the output end of the column.

3) Place column inside one of the included collection tubes.

4) Insert assembled column and collection tube into 10ml or 15ml centrifuge tube and fasten cap.

5) Place assembled tube with column into a swinging bucket or horizontal rotor centrifuge and centrifuge for 1 minute at 1100 x g.

6) Remove carrier tube containing spin column from centrifuge and dis-assemble. Remove the collection tube, containing buffer, from the column and discard. (If needed, discard buffer in collection tube and re-assemble column and centrifuge tube and spin again for 1 minute at 1100 x g)

7) Take a new collection tube and place on column.

8) Remove the large cap on input end of column and pipet your sample into the center of the shrunken gel bed inside the column.

9) Re-assemble column, collection tube with centrifuge tube and place in centrifuge.

10) Centrifuge at 1100 x g for 4 minutes.

11) The labeled nucleic acid will be recovered inside the collection tube in approximately 50µl of TE buffer.

12) Greater than 90% of the unincorporated dNTPs will be retained in the column gel, discard the used column in an appropriate fashion.

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