Ver. 04.19.19

For Research Use Only

IBI SCIENTIFIC

INSTRUCTION MANUAL

Midi Fast Ion Plasmid Kit &
Midi Fast Ion Plasmid Kit (Endotoxin Free)

IB47110, IB47112 (2 Preparation Sample Kit)

IB47111, IB47113 (25 Preparation Kit)

Advantages

Sample: cultured bacterial cells (high-copy = 50-200 ml, low-copy = 100-300 ml)

Yield: 350 µg of transfection grade plasmid DNA from 100 ml of cultured bacterial cells

Format: anion-exhange resin column, gravity flow

Endotoxin Removal: <0.1 EU/µg DNA verified by LAL when using PER Buffer

Operation Time: within 80 minutes (only 50 minutes when combined with the Plasmid Concentrator)

Elution Volume: 500 µl-2 ml

Kit Storage: dry at room temperature (15-25°C)

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Introduction

The Midi Fast Ion Plasmid Kit uses pre-packed anion-exchange resin columns to purify plasmid DNA from 50-300 ml of cultured bacterial cells. I-Blue Lysis Buffer (an optional color indicator) is included with the kit in order to prevent common handling errors, ensuring efficient cell lysis and neutralization. A modified alkaline lysis method and RNase treatment are used to obtain clear cell lysate with minimal genomic DNA/RNA contaminants. Using an efficient gravity-flow procedure, plasmid DNA is bound to the column and contaminants are efficiently removed. The purified plasmid DNA is eluted then precipitated with isopropanol for desalting. The entire procedure can be completed without ultracentrifuges, HPLC or other toxic reagents and the purified plasmid DNA is suitable for transfection, sequencing reactions, ligation, PCR, in-vitro transcription, microinjection, restriction enzyme digestion and gene gun.

Quality Control

The quality of the Midi Fast Ion Plasmid Kit is tested on a lot-to-lot basis by isolating plasmid DNA from a 50 ml overnight E. coli (DH5α) culture, containing plasmid pBluescript (A600 > 2 U/ml). More than 300 µg of plasmid DNA is quantified with a spectrophotometer. The purified plasmid (1 µg) is used in EcoRI digestion and analyzed by electrophoresis.

Kit Components

Component	IB47110/112	IB47111/113
PM1 Buffer[1]	10 ml	110 ml
PM2 Buffer[2]	10 ml	110 ml
PM3 Buffer	10 ml	110 ml
PER Buffer*	4 ml	40 ml
PEQ Buffer	12 ml	130 ml
PMC Buffer	35 ml	125 ml x 1 260 ml x 1
PEL Buffer	25 ml	220 ml
RNase A (50 mg/ml)	Added	200 µl
Plasmid Midi Columns	2	25
I-Blue Lysis Buffer	150 µl	1.5 ml

[1] For IB47111, IB47113 add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle. PM1 and RNase A mixture should be stored at 2-8°C for up to 6 months. For IB47110, IB47112 samples, RNase A was already added to PM1 Buffer.

[2] If precipitates have formed in PM2 Buffer, warm the buffer in a 37°C water bath, followed by gentle shaking to dissolve.

* PER Buffer is used for endotoxin removal and is included in IB47112 and IB47113 only.

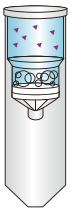
During the procedure, always wear a lab coat, disposable gloves, and protective goggles

Quick Protocol Diagram

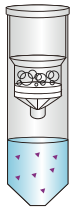
1. Harvest cultured bacterial cells by centrifuge to form a cell pellet, followed by resuspension

2. Lyse bacterial cells (optional color indicator will turn blue when lysis is successful)

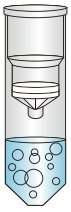
3. Neutralize suspension (optional color indicator will become clear when neutralization is successful). When using IB47112/113, neutralization is followed by PER Buffer treatment to remove endotoxin.



4. DNA binding to silica resin while contaminants remain suspended



5. Wash (removal of contaminants while DNA remains bound to silica resin)



6. Elution and precipitation of pure plasmid DNA which is ready for subsequent reactions

Recommended Culture Volume

Plasmid Type	Pellet Wet Weight	OD600 = 2	OD600 = 4	OD600 = 6
High-copy number	0.75 g	200 ml	100 ml	66 ml
Low-copy number	1.12 g	300 ml	150 ml	100 ml

Midi Fast Ion Plasmid Kit Protocol

Please read the entire instruction manual prior to starting the Protocol Procedure.

IMPORTANT BEFORE USE!

1. For IB47111 add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle. PM1 and RNase A mixture should be stored at 2-8°C for up to 6 months. For IB47110 samples, RNase A was already added to PM1 Buffer.
2. If precipitates have formed in PM2 Buffer, warm the buffer in a 37°C water bath followed by gentle shaking to dissolve.

Additional Requirements: 50 ml centrifuge tubes, isopropanol, 75% ethanol, TE or ddH2O

PW BUFFER HAS BEEN REPLACED WITH PMC BUFFER. DO NOT USE LEFTOVER PW BUFFER. ONLY USE PMC BUFFER WITH THIS KIT.

Protocol Procedure With Color Indicator

1. Harvesting

Transfer **cultured bacterial cells** to a 50 ml centrifuge tube then centrifuge at ≥3,000 x g for 15 minutes at room temperature to form a cell pellet. Discard the supernatant completely. Use a narrow pipette tip to ensure the supernatant is completely removed. Repeat the Harvesting step as required for up to 200 ml of high-copy or 100-300 ml of low-copy cultured bacterial cells using the same 50 ml centrifuge tube.

NOTE! Using 2 OD600 - 6 OD600 units of bacterial culture is recommended. Do not use overgrown bacterial cultures (≤16 hours incubated in a flask at 37°C with 150-180 rpm shaking). Use fresh bacterial cultures only. Solid medium and liquid medium (i.e. LB medium) should contain an antibiotic such as ampicillin.

2. Equilibration

During centrifugation, place a **Plasmid Midi Column** in a new 50 ml centrifuge tube. Equilibrate the **Plasmid Midi Column** by adding **5 ml of PEQ Buffer**. Allow the column to empty completely by gravity flow. Discard the flow-through and place the **Plasmid Midi Column** back in the 50 ml centrifuge tube then set it aside for Step 6.

3. Resuspension

Add **4 ml of PM1 Buffer (make sure RNase A was added) and 40 µl of I-Blue Lysis Buffer** to a new 50 ml centrifuge tube. Mix by shaking gently.

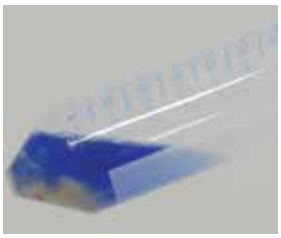
NOTE! It is normal for precipitates to form after mixing I-Blue Lysis Buffer with PM1 Buffer.

Transfer the mixture to the 50 ml centrifuge tube containing the cell pellet. Resuspend the cell pellet by vortex, pipette or scraping the tube across the top of a 1.5 ml microcentrifuge tube rack until all traces of the cell pellet have been completely dissolved.

4. Cell Lysis

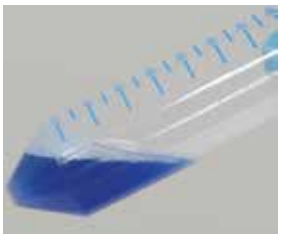
Add **4 ml of PM2 Buffer** to the resuspended sample then mix gently by inverting the tube 10 times. Close PM2 Buffer bottle immediately after use to avoid CO2 acidification. Do not vortex to avoid shearing the genomic DNA. Let stand at room temperature for at least 2 minutes to ensure the lysate is homogeneous. Do not exceed 5 minutes.

NOTE! After adding PM2 Buffer, any precipitates will be completely dissolved and the color of the suspension will become blue. If the suspension contains colorless regions or brownish cell clumps, continue mixing until the suspension is completely blue.



Insufficient Mixing

If colorless regions or brownish cell clumps are present, continue mixing until the suspension is completely blue.



Correct Mixing

5. Neutralization

Add **4 ml of PM3 Buffer** and mix immediately by inverting the tube 10 times. Do not vortex to avoid shearing the genomic DNA. Centrifuge at ≥3,000 x g for 20 minutes at room temperature.

NOTE! After adding PM3 Buffer, the suspension will become colorless. If blue regions remain in the suspension, continue mixing until it becomes colorless.

Insufficient Mixing

If blue regions are present, continue mixing until the suspension is completely colorless.

Correct Mixing

6. DNA Binding

Transfer the supernatant to the equilibrated **Plasmid Midi Column**. Allow the column to empty completely by gravity flow. Discard the flow-through then place the **Plasmid Midi Column** back in the 50 ml centrifuge tube.

7. Wash

Wash the **Plasmid Midi Column** by adding **15 ml of PMC Buffer** and allow the column to empty completely by gravity flow then discard the flow-through.

8. Elution

Place the **Plasmid Midi Column** in a clean 50 ml centrifuge tube then add **8 ml of PEL Buffer** to elute the DNA by gravity flow. Discard the **Plasmid Midi Column** once it has emptied completely.

9. DNA Precipitation

Add **6 ml (0.75 volumes) of isopropanol** to the eluted DNA from Step 8. Mix the tube completely by inverting then centrifuge at ≥3,000 x g for 20 minutes (preferably at 15,000 x g for 30 minutes) at 4°C. Carefully remove the supernatant then wash the DNA pellet with **5 ml of 75% ethanol**. Centrifuge at ≥3,000 x g for 5 minutes (preferably at 15,000 x g for 10 minutes) at 4°C. Carefully remove the supernatant then air-dry the DNA pellet for 10 minutes. Once the DNA pellet is dry, add **500 µl-2 ml (or a suitable volume) of TE[1] or water[2]** then place the tube in a 60°C water bath for 5-10 minutes to dissolve the DNA pellet.

NOTE! Following both centrifugation steps, extra caution is needed when removing the supernatant to avoid contacting the DNA pellet.

[1] Using TE (10 mM Tris-HCl, 1 mM EDTA, pH8.0) is beneficial as EDTA preserves DNA for long term storage. However, EDTA will affect PCR and other sensitive downstream applications.

[2] If using water, ensure the water pH is ≥8.0. ddH2O should be fresh as ambient CO2 can quickly cause acidification.

IMPORTANT BEFORE USE!

1. For IB47111 add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle. PM1 and RNase A mixture should be stored at 2-8ºC for up to 6 months. For IB47110 samples, RNase A was already added to PM1 Buffer.
2. If precipitates have formed in PM2 Buffer, warm the buffer in a 37ºC water bath followed by gentle shaking to dissolve.

Additional Requirements: 50 ml centrifuge tubes, isopropanol, 75% ethanol, TE or ddH2O

PW BUFFER HAS BEEN REPLACED WITH PMC BUFFER. DO NOT USE LEFTOVER PW BUFFER. ONLY USE PMC BUFFER WITH THIS KIT.

Protocol Procedure Without Color Indicator

1. Harvesting

Transfer **cultured bacterial cells** to a 50 ml centrifuge tube then centrifuge at ≥3,000 x g for 15 minutes at room temperature to form a cell pellet. Discard the supernatant completely. Use a narrow pipette tip to ensure the supernatant is completely removed. Repeat the Harvesting step as required for up to 200 ml of high-copy or 100-300 ml of low-copy cultured bacterial cells using the same 50 ml centrifuge tube.

NOTE! Using 2 OD600 - 6 OD600 units of bacterial culture is recommended. Do not use overgrown bacterial cultures (≤16 hours incubated in a flask at 37ºC with 150-180 rpm shaking). Use fresh bacterial cultures only. Solid medium and liquid medium (i.e. LB medium) should contain an antibiotic such as ampicillin.

2. Equilibration

During centrifugation, place a **Plasmid Midi Column** in a new 50 ml centrifuge tube. Equilibrate the **Plasmid Midi Column** by adding **5 ml of PEQ Buffer**. Allow the column to empty completely by gravity flow. Discard the flow-through and place the **Plasmid Midi Column** back in the 50 ml centrifuge tube then set it aside for Step 6.

3. Resuspension

Add **4 ml of PM1 Buffer (make sure RNase A was added)**. Resuspend the cell pellet by vortex, pipette or scraping the tube across the top of a 1.5 ml microcentrifuge tube rack until all traces of the cell pellet have been completely dissolved.

4. Cell Lysis

Add **4 ml of PM2 Buffer** to the resuspended sample then mix gently by inverting the tube 10 times. Close PM2 Buffer bottle immediately after use to avoid CO2 acidification. Do not vortex to avoid shearing the genomic DNA. Let stand at room temperature for at least 2 minutes to ensure the lysate is homogeneous. Do not exceed 5 minutes.

5. Neutralization

Add **4 ml of PM3 Buffer** and mix immediately by inverting the tube 10 times. Do not vortex to avoid shearing the genomic DNA. Centrifuge at ≥3,000 x g for 20 minutes at room temperature.

6. DNA Binding

Transfer the supernatant to the equilibrated **Plasmid Midi Column**. Allow the column to empty completely by gravity flow. Discard the flow-through then place the **Plasmid Midi Column** back in the 50 ml centrifuge tube

7. Wash

Wash the **Plasmid Midi Column** by adding **15 ml of PMC Buffer** and allow the column to empty completely by gravity flow then discard the flow-through.

8. Elution

Place the **Plasmid Midi Column** in a clean 50 ml centrifuge tube then add **8 ml of PEL Buffer** to elute the DNA by gravity flow. Discard the **Plasmid Midi Column** once it has emptied completely.

9. DNA Precipitation

Add **6 ml (0.75 volumes) of isopropanol** to the eluted DNA from Step 8. Mix the tube completely by inverting then centrifuge at ≥3,000 x g for 20 minutes (preferably at 15,000 x g for 30 minutes) at 4°C. Carefully remove the supernatant then wash the DNA pellet with **5 ml of 75% ethanol**. Centrifuge at ≥3,000 x g for 5 minutes (preferably at 15,000 x g for 10 minutes) at 4°C. Carefully remove the supernatant then air-dry the DNA pellet for 10 minutes. Once the DNA pellet is dry, add **500 µl-2 ml (or a suitable volume) of TE[1] or water[2]** then place the tube in a 60°C water bath for 5-10 minutes to dissolve the DNA pellet.

NOTE! Following both centrifugation steps, extra caution is needed when removing the supernatant to avoid contacting the DNA pellet.

[1] Using TE (10 mM Tris-HCl, 1 mM EDTA, pH8.0) is beneficial as EDTA preserves DNA for long term storage. However, EDTA will affect PCR and other sensitive downstream applications.
[2] If using water, ensure the water pH is ≥8.0. ddH2O should be fresh as ambient CO2 can quickly cause acidification.

Midi Fast Ion Plasmid Kit (Endotoxin Free) Protocol

Please read the entire instruction manual prior to starting the Protocol Procedure.

IMPORTANT BEFORE USE!

1. For IB47113 add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle. PM1 and RNase A mixture should be stored at 2-8°C for up to 6 months. For IB47112 samples, RNase A was already added to PM1 Buffer.
2. If precipitates have formed in PM2 Buffer, warm the buffer in a 37°C water bath followed by gentle shaking to dissolve.

Additional Requirements: 50 ml centrifuge tubes, isopropanol, 75% ethanol, TE or ddH2O

PW BUFFER HAS BEEN REPLACED WITH PMC BUFFER. DO NOT USE LEFTOVER PW BUFFER. ONLY USE PMC BUFFER WITH THIS KIT.

Protocol Procedure With Color Indicator

1. Harvesting

Transfer **cultured bacterial cells** to a 50 ml centrifuge tube then centrifuge at ≥3,000 x g for 15 minutes at room temperature to form a cell pellet. Discard the supernatant completely. Use a narrow pipette tip to ensure the supernatant is completely removed. Repeat the Harvesting step as required for up to 200 ml of high-copy or 100-300 ml of low-copy cultured bacterial cells using the same 50 ml centrifuge tube.

NOTE! Using 2 OD600 - 6 OD600 units of bacterial culture is recommended. Do not use overgrown bacterial cultures (≤16 hours incubated in a flask at 37°C with 150-180 rpm shaking). Use fresh bacterial cultures only. Solid medium and liquid medium (i.e. LB medium) should contain an antibiotic such as ampicillin.

2. Equilibration

During centrifugation, place a **Plasmid Midi Column** in a new 50 ml centrifuge tube. Equilibrate the **Plasmid Midi Column** by adding **5 ml of PEQ Buffer**. Allow the column to empty completely by gravity flow. Discard the flow-through and place the **Plasmid Midi Column** back in the 50 ml centrifuge tube then set it aside for Step 7.

3. Resuspension

Add **4 ml of PM1 Buffer (make sure RNase A was added) and 40 µl of I-Blue Lysis Buffer** to a new 50 ml centrifuge tube. Mix by shaking gently.

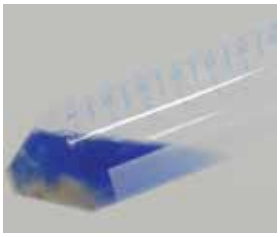
NOTE! It is normal for precipitates to form after mixing I-Blue Lysis Buffer with PM1 Buffer.

Transfer the mixture to the 50 ml centrifuge tube containing the cell pellet. Resuspend the cell pellet by vortex, pipette or scraping the tube across the top of a 1.5 ml microcentrifuge tube rack until all traces of the cell pellet have been completely dissolved.

4. Cell Lysis

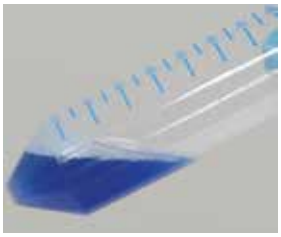
Add **4 ml of PM2 Buffer** to the resuspended sample then mix gently by inverting the tube 10 times. Close PM2 Buffer bottle immediately after use to avoid CO2 acidification. Do not vortex to avoid shearing the genomic DNA. Let stand at room temperature for at least 2 minutes to ensure the lysate is homogeneous. Do not exceed 5 minutes.

NOTE! After adding PM2 Buffer, any precipitates will be completely dissolved and the color of the suspension will become blue. If the suspension contains colorless regions or brownish cell clumps, continue mixing until the suspension is completely blue.



Insufficient Mixing

If colorless regions or brownish cell clumps are present, continue mixing until the suspension is completely blue.



Correct Mixing

5. Neutralization

Add **4 ml of PM3 Buffer** and mix immediately by inverting the tube 10 times. Do not vortex to avoid shearing the genomic DNA. Centrifuge at ≥3,000 x g for 20 minutes at room temperature.

NOTE! After adding PM3 Buffer, the suspension will become colorless. If blue regions remain in the suspension, continue mixing until it becomes colorless.

Insufficient Mixing

If blue regions are present, continue mixing until the suspension is completely colorless.

Correct Mixing

6. Endotoxin Removal

NOTE! Invert PER Buffer bottle 3-5 times immediately prior to use.

Transfer the supernatant to a clean 50 ml centrifuge tube. Add 1.2 ml of PER Buffer then mix by inverting 5-10 times. Incubate on ice for 30 minutes.

NOTE! Following PER Buffer addition, the mixture will become cloudy.

7. DNA Binding

Following ice incubation, **transfer the mixture** to the equilibrated **Plasmid Midi Column**. Allow the column to empty completely by gravity flow. Discard the flow-through then place the **Plasmid Midi Column** back in the 50 ml centrifuge tube.

8. Wash

Wash the **Plasmid Midi Column** by adding **15 ml of PMC Buffer** and allow the column to empty completely by gravity flow then discard the flow-through.

9. Elution

Place the **Plasmid Midi Column** in a clean 50 ml centrifuge tube then add **8 ml of PEL Buffer** to elute the DNA by gravity flow. Discard the **Plasmid Midi Column** once it has emptied completely.

10. DNA Precipitation

Add **6 ml (0.75 volumes) of isopropanol** to the eluted DNA from Step 9. Mix the tube completely by inverting then centrifuge at ≥3,000 x g for 20 minutes (preferably at 15,000 x g for 30 minutes) at 4ºC. Carefully remove the supernatant then wash the DNA pellet with **5 ml of 75% ethanol**. Centrifuge at ≥3,000 x g for 5 minutes (preferably at 15,000 x g for 10 minutes) at 4ºC. Carefully remove the supernatant then air-dry the DNA pellet for 10 minutes. Once the DNA pellet is dry, add **500 µl-2 ml (or a suitable volume) of TE[1] or water[2]** then place the tube in a 60ºC water bath for 5-10 minutes to dissolve the DNA pellet.

NOTE! Following both centrifugation steps, extra caution is needed when removing the supernatant to avoid contacting the DNA pellet.

[1] Using TE (10 mM Tris-HCl, 1 mM EDTA, pH8.0) is beneficial as EDTA preserves DNA for long term storage. However, EDTA will affect PCR and other sensitive downstream applications.
[2] If using water, ensure the water pH is ≥8.0. ddH2O should be fresh as ambient CO2 can quickly cause acidification.

IMPORTANT BEFORE USE!

1. For IB47113 add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle. PM1 and RNase A mixture should be stored at 2-8ºC for up to 6 months. For IB47112 samples, RNase A was already added to PM1 Buffer.
2. If precipitates have formed in PM2 Buffer, warm the buffer in a 37ºC water bath followed by gentle shaking to dissolve.

Additional Requirements:
50 ml centrifuge tubes, isopropanol, 75% ethanol, TE or ddH2O

PW BUFFER HAS BEEN REPLACED WITH PMC BUFFER. DO NOT USE LEFTOVER PW BUFFER. ONLY USE PMC BUFFER WITH THIS KIT.

Protocol Procedure Without Color Indicator

1. Harvesting

Transfer **cultured bacterial cells** to a 50 ml centrifuge tube then centrifuge at ≥3,000 x g for 15 minutes at room temperature to form a cell pellet. Discard the supernatant completely. Use a narrow pipette tip to ensure the supernatant is completely removed. Repeat the Harvesting step as required for up to 200 ml of high-copy or 100-300 ml of low-copy cultured bacterial cells using the same 50 ml centrifuge tube.

NOTE! Using 2 OD600 - 6 OD600 units of bacterial culture is recommended. Do not use overgrown bacterial cultures (≤16 hours incubated in a flask at 37ºC with 150-180 rpm shaking). Use fresh bacterial cultures only. Solid medium and liquid medium (i.e. LB medium) should contain an antibiotic such as ampicillin.

2. Equilibration

During centrifugation, place a **Plasmid Midi Column** in a new 50 ml centrifuge tube. Equilibrate the **Plasmid Midi Column** by adding **5 ml of PEQ Buffer**. Allow the column to empty completely by gravity flow. Discard the flow-through and place the **Plasmid Midi Column** back in the 50 ml centrifuge tube then set it aside for Step 7.

3. Resuspension

Add **4 ml of PM1 Buffer (make sure RNase A was added)**. Resuspend the cell pellet by vortex, pipette or scraping the tube across the top of a 1.5 ml microcentrifuge tube rack until all traces of the cell pellet have been completely dissolved.

4. Cell Lysis

Add **4 ml of PM2 Buffer** to the resuspended sample then mix gently by inverting the tube 10 times. Close PM2 Buffer bottle immediately after use to avoid CO2 acidification. Do not vortex to avoid shearing the genomic DNA. Let stand at room temperature for at least 2 minutes to ensure the lysate is homogeneous. Do not exceed 5 minutes.

5. Neutralization

Add **4 ml of PM3 Buffer** and mix immediately by inverting the tube 10 times. Do not vortex to avoid shearing the genomic DNA. Centrifuge at ≥3,000 x g for 20 minutes at room temperature.

6. Endotoxin Removal

NOTE! Invert PER Buffer bottle 3-5 times immediately prior to use.

Transfer the supernatant to a clean 50 ml centrifuge tube. Add **1.2 ml of PER Buffer** then mix by inverting 5-10 times. Incubate on ice for 30 minutes.

NOTE! Following PER Buffer addition, the mixture will become cloudy.

7. DNA Binding

Following ice incubation, **transfer the mixture** to the equilibrated **Plasmid Midi Column**. Allow the column to empty completely by gravity flow. Discard the flow-through then place the **Plasmid Midi Column** back in the 50 ml centrifuge tube.

8. Wash

Wash the **Plasmid Midi Column** by adding **15 ml of PMC Buffer** and allow the column to empty completely by gravity flow then discard the flow-through.

9. Elution

Place the **Plasmid Midi Column** in a clean 50 ml centrifuge tube then add **8 ml of PEL Buffer** to elute the DNA by gravity flow. Discard the **Plasmid Midi Column** once it has emptied completely.

10. DNA Precipitation

Add **6 ml (0.75 volumes) of isopropanol** to the eluted DNA from Step 9. Mix the tube completely by inverting then centrifuge at ≥3,000 x g for 20 minutes (preferably at 15,000 x g for 30 minutes) at 4°C. Carefully remove the supernatant then wash the DNA pellet with **5 ml of 75% ethanol**. Centrifuge at ≥3,000 x g for 5 minutes (preferably at 15,000 x g for 10 minutes) at 4°C. Carefully remove the supernatant then air-dry the DNA pellet for 10 minutes. Once the DNA pellet is dry, add **500 µl-2 ml (or a suitable volume) of TE[1] or water[2]** then place the tube in a 60°C water bath for 5-10 minutes to dissolve the DNA pellet.

NOTE! Following both centrifugation steps, extra caution is needed when removing the supernatant to avoid contacting the DNA pellet.

[1] Using TE (10 mM Tris-HCl, 1 mM EDTA, pH8.0) is beneficial as EDTA preserves DNA for long term storage. However, EDTA will affect PCR and other sensitive downstream applications.
[2] If using water, ensure the water pH is ≥8.0. ddH2O should be fresh as ambient CO2 can quickly cause acidification.

Troubleshooting

Low Yield

Incomplete buffer preparation.

For IB47111/113 add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle then store at 2-8°C for up to 6 months. For IB47110/112 samples, RNase A was already added to PM1. If precipitates have formed in PM2 Buffer, warm in a 37°C water bath followed by gentle shaking to dissolve.

Incomplete cell culture preparation.
We recommend using a single freshly isolated E. coli colony to inoculate into 50-100 ml of LB medium. Solid and liquid medium should contain antibiotics. Do not use overgrown bacterial cultures (≤16 hours incubated in a flask at 37ºC with 150-180 rpm shaking).

Culture growth medium was not removed completely.
Following centrifugation in the Harvesting step, use a narrow pipette tip to ensure the supernatant is completely removed.

Cell pellet was not resuspended completely.
Resuspend the cell pellet completely by vortex or pipette.

Bacterial cells were not lysed completely.
Using 2 OD600 - 6 OD600 units of bacterial culture is recommended.
When using I-Blue Lysis Buffer: Following PM2 Buffer addition, the color of the suspension will become blue. If the suspension contains colorless regions or brownish cell clumps, continue mixing until the suspension is completely blue. Do not vortex to avoid shearing the genomic DNA.

Bacterial cells were not neutralized completely.
When using I-Blue Lysis Buffer: Following PM3 Buffer addition, the suspension will become colorless. If blue regions remain in the suspension, continue mixing until it becomes colorless. Do not vortex to avoid shearing the genomic DNA.

Incorrect DNA Rehydration.
If using water to dissolve the DNA pellet, ensure the water pH is ≥8.0. ddH2O should be fresh as ambient CO2 can quickly cause acidification.

No yield of plasmid DNA.
Increase volume of low-copy number plasmid to 300 ml. We recommend using a single freshly isolated E. coli colony to inoculate into 50-100 ml of LB medium. Solid and liquid medium should contain antibiotics. Do not use overgrown bacterial cultures.

Eluted DNA Does Not Perform Well In Downstream Applications

RNA contamination.
Add provided RNase A to PM1 Buffer then mix by shaking for a few seconds. Check the box on the bottle then store at 2-8ºC for up to 6 months.

Genomic DNA contamination.
Do not use overgrown bacterial cultures. Use only fresh cultures as they will contain less genomic DNA than old cultures. During PM2 and PM3 Buffer addition, mix gently to prevent genomic DNA shearing. After adding PM3 Buffer, mix thoroughly then incubate the lysate on ice for 10-15 minutes to enhance precipitation and reduce genomic DNA contamination.

Midi Fast Ion Plasmid Kit Functional Test Data

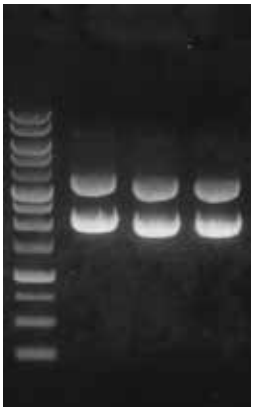


Figure 1. Plasmid DNA from a 100 ml overnight E. coli (DH5α) culture, containing plasmid pBluescript (OD600 = 3.65) was purified using the IBI Midi Plasmid Kit. The purified supercoiled plasmid DNA was analyzed by electrophoresis on a 1% agarose gel. M = IBI 1 Kb DNA Ladder

Test	DNA Conc.	260/280	260/230	Yield
1	332.1 µg/ml	1.87	2.22	332.1 µg
2	345.5 µg/ml	1.87	2.25	345.5 µg
3	339.2 µg/ml	1.87	2.24	339.2 µg

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