

NIH Public Access

Author Manuscript

Recent Pat Antiinfect Drug Discov. Author manuscript; available in PMC 2011 June 1

Published in final edited form as: *Recent Pat Antiinfect Drug Discov.* 2010 June 1; 5(2): 124–151.

Topical Antimicrobials for Burn Wound Infections

Tianhong Dai^{1,2}, Ying-Ying Huang^{1,2,3}, Sulbha K. Sharma¹, Javad T. Hashmi^{1,2}, Divya B. Kurup¹, and Michael R. Hamblin^{1,2,4,*}

¹ Wellman Center for Photomedicine, Massachusetts General Hospital, Boston, MA

² Department of Dermatology, Harvard Medical School, Boston, MA

³ Aesthetic and Plastic Center of Guangxi Medical University, Nanning, P.R China

⁴ Harvard-MIT Division of Health Sciences and Technology, Cambridge, MA

Abstract

Throughout most of history, serious burns occupying a large percentage of body surface area were an almost certain death sentence because of subsequent infection. A number of factors such as disruption of the skin barrier, ready availability of bacterial nutrients in the burn milieu, destruction of the vascular supply to the burned skin, and systemic disturbances lead to immunosuppression combined together to make burns particularly susceptible to infection. In the 20th century the introduction of antibiotic and antifungal drugs, the use of topical antimicrobials that could be applied to burns, and widespread adoption of early excision and grafting all helped to dramatically increase survival. However the relentless increase in microbial resistance to antibiotics and other antimicrobials has led to a renewed search for alternative approaches to prevent and combat burn infections. This review will cover patented strategies that have been issued or filed with regard to new topical agents, preparations, and methods of combating burn infections. Animal models that are used in preclinical studies are discussed. Various silver preparations (nanocrystalline and slow release) are the mainstay of many approaches but antimicrobial peptides, topical photodynamic therapy, chitosan preparations, new iodine delivery formulations, phage therapy and natural products such as honey and essential oils have all been tested. This active area of research will continue to provide new topical antimicrobials for burns that will battle against growing multi-drug resistance.

Keywords

Burn infection; multi-drug resistance; animal models; bioluminescence imaging; nanocrystalline silver; antimicrobial peptide; skin substitute; chitosan; photodynamic therapy

1. INTRODUCTION

1.1. History

Despite various current topical treatment regimens designed to eradicate the bacterial load within the burn wound, sepsis remains the leading cause of death in burn units around the world [1]. Advances in resuscitation, surgical management, control of infection, control of the hypermetabolic response and rehabilitation have resulted in dramatic improvements in burn mortality and morbidity in the last 60 years [2]. Infection is a major complication of burns. Infection is

Address correspondence to this author at the 40 Blossom Streetb BAR414, Wellman Center for Photomedicine, Massachusetts General Hospital, Boston, MA 02114-2696 USA; Tel: (617) 726-6182; Fax: (617) 726-6182; hamblin@helix.mgh.harvard.edu. **CONFLICT OF INTEREST** None

linked to impaired resistance from disruption of the skin's mechanical integrity and generalized immune suppression. The skin barrier is replaced by eschar. This moist, protein rich avascular environment encourages microbial growth. Migration of immune cells is hampered, and there is a release of intermediaries that impede the immune response. Eschar also restricts distribution of systemically administered antibiotics because of its avascularity.

1.2. Physiology of Burn

Burns are classified by their depth and severity as 1st, 2nd, 3rd and 4th degree Fig (1). Firstdegree burns are usually limited to redness (erythema), a white plaque and minor pain at the site of injury. These burns involve only the epidermis. Second-degree burns manifest as erythema with superficial blistering of the skin, involving the superficial (papillary) dermis and may also involve the deep (reticular) dermis layer. Third-degree burns occur when the epidermis is lost with damage to the subcutaneous tissue. Burn victims will exhibit charring and extreme damage of the epidermis, and sometimes hard eschar will be present. Fourthdegree burns damage muscle, tendon, and ligament tissue, thus result in charring and catastrophic damage of the subcutaneous tissue.

Burns can also be assessed in terms of total body surface area (%TBSA), which is the percentage affected by partial thickness or full thickness burns. The rule of nines is used as a quick and useful way to estimate the affected TBSA. More accurate estimation can be made using Lund & Browder charts which take into account the different proportions of body parts in adults and children [3]Fig. (2). Burns greater than 10% in children or 15% in adults are potentially life-threatening injuries (because of the risk of hypovolemic shock) and should have formal fluid resuscitation and monitoring in a burns unit.

Severely burned skin ceases to perform its natural protective and barrier role and allows a dramatic increase in water loss and can become a portal for bacterial invasion. The activation of a pro-inflammatory cascade after a burn appears to be important in the development of subsequent immune dysfunction, bacterial translocation from the gut, susceptibility to sepsis and multiple organ failure. A local response to burning involves not only direct tissue coagulation but also burn tissue conversion, a process where the damaged cells, rather than recover, progress to cell death, extending the depth and severity of the original injury [4]. The systemic response to burning is driven by the loss of the skin barrier and release of vasoactive mediators from the wound and from subsequent infection. When the burn size exceeds about 25% of the body surface, interstitial edema develops in distant organs and soft tissues, mainly secondary to a combination of wound-released mediators and hypoproteinemia [5]. Thermal injury causes a massive release of proinflammatory cytokines, chemical mediators including histamine, complement, arachidonic acid, products of the coagulation cascade, and oxygen free radicals, which produce an increase in vascular permeability leading to hypovolemia and acute renal failure [5]. This may be complicated by systemic inflammatory response syndrome and marked immune suppression. Subsequent wound infection and bacterial translocation from the gastrointestinal tract promotes sepsis [6]. This adds to the final common pathway to multiorgan failure and death Fig. (3).

1.3. Microbiology of Burn Infection

Table 1 [7–15] gives a listing of the microbial species responsible for invasive burn wound infection [16]. The incidence of antibiotic resistance amongst these species is also given.

A consensus conference of the American Burn Association laid down guidelines on the distinction between colonization (virtually all burns) vs. infection (likely to produce disease) in burn wounds [17]. The most serious microbial infections are classified as "invasive". This property is a function of the seriousness of the burn and the virulence of the microbial species

or strain. Virulence is a combination of a variety of factors including the ability to colonize the burn (including adhesion to host cells and tissues), the expression of proteolytic enzymes, the ability to obtain nutrition from the host, and the ability to evade the host's immune response.

There is a fairly predictable time course of the microbial species that colonize or infect burns [18]. Gram-positive bacteria that survive the thermal insult, such as staphylococci located deep within sweat glands and hair follicles, heavily colonize the wound surface within the first 48 hours. Eventually (after an average of 5–7 days), burns are colonized with other microbes derived from the host's normal gastrointestinal and upper respiratory flora or from the hospital environment. These pathogens include Gram-negative bacteria, while yeasts and fungi tend to be the latest colonizers. The cell wall structures of these three classes of pathogenic microorganisms are shown in Fig. (4). The architectural differences largely govern the relative effectiveness of many antimicrobial agents that induce disruption of the integrity of the cell wall and subsequently the cell membrane.

1.4. Physiology of Burn Infection

Many physiological properties of the burn environment predispose to infection and inhibit traditional treatment by systemic antibiotics. The destruction of the microvasculature by the thermal damage means that the burn is considered to be non-vascularized tissue. Proteolysis and lipolysis is up-regulated in burns [19]. A marked increase in matrix metalloproteinases (and reduction in inhibitors) leads to breakdown of proteins thus providing more nutrients for microbes and making microbial penetration into the tissue easier.

Immunosuppression plays a big role in predisposing to infection in burns. Both innate and adaptive immune systems are involved in immunosuppression. Innate immunity is triggered immediately after microbial invasion in response to highly conserved structures present in large groups of microorganisms, while adaptive immunity is a function of T and B cells, a small number of which are activated selectively by particular antigens to expand clonally, a process that takes at least three to five days to become evident. Alterations in the innate immune system following thermal injury include neutrophils, macrophages, monocytes, basophils, NK-cells, and complement. In contrast, Thermal injury is associated with many mediators originating from the cytokine network, the endocrine system, and the arachidonic acid cascade which mediates immunosuppression. Initially, the immunologic response to severe burn injury is proinflammatory but later becomes predominately anti-inflammatory in an effort to maintain homeostasis and restore normal physiology. The anti-inflammatory response and the subsequent immunosuppression following burn injury are characterized by a set of opposing cell types and cytokines. Many of the changes in cytokine levels represent alterations of the adaptive immune system following burn injury, more specifically within the T-lymphocyte population.

Bacterial translocation is another physiological response caused by a combination of burns with a microbial insult and predisposes to development of sepsis [18]. The term 'bacterial translocation' is used to describe the passage of viable resident bacteria from the gastrointestinal tract to normally sterile tissues such as the mesenteric lymph nodes and other internal organs. Many systemic stresses can destroy the mucosal barrier of the intestines such as elective abdominal surgery, organ donors and those with intestinal obstruction, colorectal cancer, ischemia-reperfusion injury shock and pancreatitis. It has been shown [20] in a rat model that a two-hit injury consisting of burn and *Enterococcus faecalis* infection increased intestinal permeability and Gram-negative sepsis.

1.5. Antibiotic Resistance

The world-wide problem of growing antibiotic resistance shows no sign of ameliorating and the present time has been termed the "end of the antibiotic era" [21]. Commentators have asserted that if nothing is done, hitherto trivial infections that were easily treated with a simple course of antibiotics, could in future become essentially untreatable as in the days before antibiotics were discovered [22]. Resistance has been attributed to the widespread availability of antibiotics over the counter, wrong prescription practices (for instance giving antibiotics for respiratory viral infections), poor patient compliance leading to stopping treatment too early and overuse of antibiotics in livestock feedstuffs [23]. As shown in Table 1 resistance is also becoming common in burn pathogens.

1.6. Topical Antimicrobials

Because of the factors discussed above, topical antimicrobial preparations have been particularly applied to prevent and treat burn infections compared to other traumatic, surgical and medical indications which could be susceptible to infection. Non-healing chronic wounds such as diabetic, vascular and pressure ulcers are another similar indication [24]. Many of the agents are designed to be used prophylactically to prevent infection developing, while others are designed to kill the actually microbial cells that are proliferating within the burn when an infection has developed. Table 2 [25-80] gives a listing of topical antimicrobial therapy agents used for burns that forms the bulk of this review. Since many of these topical agents are applied onto the surface of the burn, if they are designed to be actively microbicidal, consideration must be given to the degree which the agents will penetrate into the burned infected tissue to reach the microbial cells that have invaded. Stephanides et al. [81] measured the extent to which several agents (gentamicin sulfate, mafenide acetate, nitrofurazone, povidone-iodine, silver nitrate, and silver sulfadiazine) penetrated through burn eschar. Studies [82,83] have been carried out investigating the ability of "penetration enhancing agents" such as (glycerin, saline, sodium dodecyl sulfate, ethanol, hexane: ethanol, ethyl acetate:ethanol dimethyl sulfoxide, glycine and terpenes) to increase the penetration of diverse antimicrobial agents into the eschar of 3rd degree burns. Another consideration that is important in the field of topical antimicrobials is the selectivity towards microbial cells versus cytotoxicity to host cells and tissue. Many antimicrobial agents such as topical antibiotics [84], antiseptics [85], silver preparations [86,87], antimicrobial peptides [88], antimicrobial photodynamic therapy [4,89] have been investigated for possibility toxicity towards skin and other human cells.

2. ANIMAL MODELS OF BURN INFECTIONS

The use of animals as models for burn infections has been a fundamental part of burn infection research for many decades. By establishing a burn infection similar to that observed in humans, the reasons for the development of infection can be researched, and ultimately, the goal is to seek approaches by which the infection can be thwarted. Animal species that have been used for burn models include mouse, rat, rabbit, as well as pig with the means of inflicting the burn injuries covering diverse techniques such as boiling water, ethanol bath, heated brass blocks, etc.

The severity of burn infections in animals and whether the animals develop sepsis and die is determined by the following factors (among others): the number of bacteria applied to the wound, the virulence of the particular bacterial strain, the size of the wound expressed as % of total body-surface area (TBSA), whether the bacteria are applied to the surface or injected into or beneath the burn, and the length of time the heated object or liquid is in contact with the skin (which determines whether a full thickness or partial thickness burn is delivered).

2.1. Mason-Walker Burn Model

About 40 years ago in 1968, Mason and Walker developed a burn model of rat using boiling water to inflict the thermal injury [90]. It was the first animal model burn for research and was used as a standard scald burn model by the U.S. Army Surgical Research Center in San Antonio TX for various studies [90]. The burning device was made of a thin metal half-cylinder with an aperture of calculated size cut from the central portion of the half-cylinder. The dimensions of the aperture were selected to provide the size of burn desired in animal of known weight (the upper limit of burn size permitting by this approach was approximately 30% of TBSA). The anesthetized animal was shaved on the back and then placed supine in the burning device. The exposed area was immersed in boiling water. It was reported that 10 seconds exposure produced a full-thickness burn, and 3 seconds a partial-thickness burn. The burns *per se* did not interfere greatly with mobility and the animals ate and drank easily. This model has been used widely in the studies on burn infections, including bacterial translocation in burn infections [91], development of antimicrobial peptides for eliminating multi-drug resistant bacteria in burns [92], development of wound dressing for preventing burn infections [93], etc.

2.2. Stieritz-Holder Burn Model

Stieritz and Holder reported an ethanol bath burn model in mouse [94]. In this model, female CF1 mice weighing 22–24 g were used. Mice were anesthetized and shaved on the back. An asbestos board with a window accounting for approximately 30% of TBSA was pressed firmly against the back of mice. Ethanol was evenly spread over the area outline by the window, ignited, and allowed to burn for 10 seconds. The infections were initiated by an immediate subcutaneous injection of 100 CFU of *Pseudomonas aeruginosa* in the burned area. The infections turned out to be rapidly fatal and the animals appeared moribund 20 hours after infection.

2.3. Other Burn Models

Stevens *et al.* [95] developed a mouse model of full-thickness burn by applying two pre-heated brass blocks (92–95 °C) to the opposing sides of an elevated skin folder on the backs of shaved mice for 5 seconds. This procedure is illustrated in Fig. (5). Six-week old male CD1 mice weighing 25–30 g were used. The combined brass block area was 1.8 cm × 1.8 cm, corresponding to a 5% of TBSA. After the infliction of burns, the eschars were immediately injected intradermally with $10-10^6$ CFU of *P. aeruginosa*. Survival at 10 days was 100% with burn injury alone and 60% with infected burns. *P. aeruginosa* (consistently 10^8 CFU/g-tissue) were recovered from the unburned muscle by 24 hours for all the different inocula of bacteria.

A model of a partial thickness burn was developed by Bahar *et al.* [96] using male Wistar rats weighing 250–300 g. After the animal was anesthetized, a 2.5 cm \times 2.5 cm, 5 mm-thick piece of absorbent lint cloth that had been immersed in boiling water (100 °C) was placed over the dorsum. The hot cloth was applied for different time intervals. This method produced reliable superficial and deep dermal burns, as confirmed by the histological features.

Suzuki *et al.* [97] described a burn model of rat based on skin contact with a glass chamber through which water circulates at a predetermined temperature. This allows application at a constant pressure of 10 g/cm^2 . The great advantage of this model is the possibility of varying temperature and exposure time, as required by the researcher, and also of applying higher or lower contact pressure.

2.4. Bioluminescence Imaging of Infection

Contag *et al.* [98] developed a method to monitor bacterial numbers and viability in real-time in infected wounds of living animals using genetically engineered bacteria that emit

luminescence together with a sensitive low-light imaging camera. The rate of luciferase enzyme turnover in the presence of substrate allows for real-time measurements, and the enzyme is active at the body temperature of mammals. This method is a significant improvement on the traditional use of survival or body fluid sampling and subsequent plating and colony counting. The first method suffers from the disadvantage of being wasteful of animals, and does not really address the question of where the bacteria are in the animal, while the second method suffers from the disadvantage that tissue sampling introduces another source of experimental error, is laborious and does not give real-time results.

The imagining system Fig. (6a) used in our laboratory [99] consists of an ICCD photoncounting camera (Hamamatsu Photonics, Bridgewater, NJ) mounted in a light-tight specimen chamber, fitted with a light-emitting diode, a setup that allowed for a background gray-scale image of the entire mouse to be captured. By accumulating many images containing binary photon information (an integration time of 2 minutes was used), a pseudo-color luminescence image was generated. Superimposition of this image onto the gray-scale background image yielded information on the location and intensity in terms of photon number Fig. (6b). The camera was also connected to a computer system through an image processor (Argus-50, Hamamatsu Photonics). Argus-50 control program (Hamamatsu Photonics) was used to acquire images and to process the image data collected. This entire wound photon count was quantified as relative luminescence units (RLU) and was displayed in a false color scale ranging from pink (most intense) to blue (least intense). Another popular bioluminescence camera is the *In Vivo* Imaging System (IVIS) made by Caliper Life Sciences (Hopkinton, MA) [100].

Figure (6C) displays the successive bioluminescence images of representative mouse burns infected with bioluminescent strains of *P. aeruginosa, Acinetobacter baumannii, Staphylococcus aureus,* and *Candida albicans,* respectively. For *P. aeruginosa* infection, which was acute and lethal, the mouse died on day 3 post-infection. For *A. baumannii, S. aureus,* and *C. albicans* infections, which were chronic, the infections sustained in the mice for 20, 17, and 14 days, respectively.

3. SKIN SUBSTITUTES

In the 1980's the critical need to address the problem of early coverage of extensive burn injuries in F73patients with inadequate sources of autologous skin for grafting prompted the emergence of skin substitutes primarily as an experimental therapy [101]. The preponderance of studies and trials demonstrates that the use of skin substitutes facilitates better healing in burns, and minimizes the risk of infections. It should be emphasized that these products are not active antimicrobial agents but rather serve as biocompatible barriers to prevent infection [102]. Nevertheless they are frequently compared with the standard antimicrobial agent AgSD. Their use results in a measured improvement in elasticity and flexibility of the healed burn, lessened scar contracture a reduction in the number of scar contracture procedures and plastic surgical reconstructions required long term

Most pediatric burns are small-sized burns of intermediate depth and warrant only local treatment. The use of skin substitutes eliminates the need of changing dressings and repeated visits to the hospital clinics for that purpose. This helps in avoiding the pain, emotional trauma and promotes a cost effective method of management whereby any further treatment is unnecessary and the child returns to its daily activities unrestricted.

Synthetic skin substitutes include Duoderm (polyurethane and hydrocolloids) Opsite (polyurethane film), Omiderm (acrylamide film and hydroxyethylmethyl-metha-crylate with polyurethane). Biosynthetic skin substitutes (biological materials like collagen in combination with synthetic materials) include Biobrane (silicone film, nylon, collagen -derived peptides)

and Transcyte (polymer with fibroblasts). Biological skin substitutes include homologous skin, porcine skin, collagen derivatives, human amniotic membrane and cultured allografts.

3.1. Duoderm

Duoderm is a hydrocolloid preparation that provides a moist environment that enhances the wound healing process and prevents infection at the site patented in 1985 [25]. Martin *et al.* [103] studied the effect of Duoderm on partial-thickness scalds in the pediatric age group Two hundred forty-eight pediatric burns were analyzed and the Duoderm-managed patients demonstrated a significantly lower graft rate.

3.2. Omniderm

Omniderm (ITG Laboratories, Redwood City, CA) consists of a water-vapor permeable polyurethane film [27]. Eldad *et al.* [104] examined the use of Omniderm as an interface in burn wounds requiring skin grafts, particularly on problematical areas and wounds that were heavily contaminated with bacteria. They concluded that about 75 per cent improvement was achieved in comparison with other dressings under similar conditions.

3.3. Suprathel

Suprathel[®] (PolyMedics Innovations Gmbh, Stuttgart, Germany) is an absorbable, synthetic wound dressing with properties of natural epithelium [28]. It consists of a copolymer of polylactide, trimethylene carbonate and epsilon-caprolactone. It becomes transparent upon application which allows close wound monitoring. Schwarze and others [105] evaluated the impact on wound healing of Suprathel in partial-thickness burn injuries. Thirty patients suffering from second-degree burn injuries were included in the study, with a mean of age 40.4 years. Burn injuries were randomly selected, partly treated with Omiderm and partly treated with Suprathel. They concluded that Suprathel showed a good impact on wound healing and pain reduction in partial-thickness burn injuries and also provided a significant increase of patient comfort.

3.4. Epigard and SYSpur-Derm

Epigard is a two-layered, non-medicated wound dressing which approximates the function of human skin [29]. The synthetic skin substitute SYSpur-derm is a bilaminar poly-urethane foam substance [30]. Mahnke and others [106] evaluated the efficacy of SYSpur-derm and Epigard by examining 85 skin biopsies taken from 23 pigs with experimental 3rd-degree burns. It was concluded that both the materials demonstrated an exceptional wound healing, showed good adhesion to the wound surface and had no toxic or antigenic effect.

3.5. Biobrane

Biobrane (Smith and Nephew and UDL Laboratories Inc.) was patented in 1989 [31] and consists of a silicone film, bonded to a cross-linked nylon fabric. Both layers are covered with a layer of hydrophilic collagen peptides which makes the dressing hydrophilic and tissue-compatible. Blood/sera clot in the nylon matrix, thereby enabling firm adherence of the dressing to the wound until epithelialization occurs. Barret *et al.* [107] hypothesized that the treatment of second-degree burns with Biobrane is superior to topical treatment. They concluded that the treatment of partial-thickness burns with Biobrane is superior to topical therapy with 1% silver sulfadiazine. It also markedly reduced pain, pain medication requirements, wound healing time, and length of hospital stay. Hassan and Shaw reported [108] punctate scarring from use of porous Biobrane which makes the use of this particular product questionable.

3.6. Transcyte and Dermagraft

TransCyte (Smith and Nephew and Advanced Tissue Sciences) is a human fibroblast-derived temporary skin substitute composed of a polymer membrane and neonatal human fibroblast cells cultured under aseptic conditions *in vitro* on a nylon mesh. The nylon mesh is coated with porcine dermal collagen and bonded to a polymer membrane (silicone), which provides a transparent synthetic epidermis upon application. As fibroblasts proliferate within the nylon mesh, they secrete human dermal collagen, matrix proteins and growth factors. Following freezing, no cellular metabolic activity remains; however, the tissue matrix and bound growth factors are left intact. It provides a temporary protective barrier and is transparent which allows direct visual monitoring of the wound bed. Amani *et al.* [26] undertook a study to determine if Transcyte reduces the length of hospital stay for partial thickness burns of any size or etiology. Significant difference was found between patients who were treated with dermabrasion and Transcyte compared with a population receiving standard therapy. The authors suggest that this modality is more efficacious and markedly reduces length of stay compared to traditional management.

Dermagraft (Advanced BioHealing) is a cryopreserved human fibroblast-derived dermal substitute; it is composed of fibroblasts, extracellular matrix, and a bioabsorbable scaffold patented in 1990 [109]. Dermagraft is manufactured from human fibroblast cells derived from newborn foreskin tissue seeded onto a bioabsorbable polyglactin mesh scaffold. The fibroblasts proliferate to fill the interstices of this scaffold and secrete human dermal collagen, matrix proteins, growth factors, and cytokines to create a three-dimensional human dermal substitute containing metabolically active, living cells.

3.7. Xenoderm and Alloderm

Xenoderm and Alloderm (LifeCell, Inc) are acellular matrices derived from porcine (Xenoderm) or human (Alloderm) skin. The tissue goes through a cell removal process while retaining the important biochemical and structural components [32]. Hosseini *et al.* [110] studied the outcomes of Xenoderm dressing compared with 1% silver sulfadiazine (SSD) in partial-thickness burns taking into account the wound infection, length of hospital stay, number of dressings and doses of analgesics used. 78 patients were considered for the study of which 37 were treated with daily washing along with application of topical SSD dressing and 39 with Xenoderm. The study concluded that Xenoderm was significantly more effective.

3.8. Other Skin Substitutes

Apligraf (Organogenesis Inc) is supplied as a living, bi-layered skin substitute: the epidermal layer is formed by human keratinocytes and has a well-differentiated stratum corneum; the dermal layer is composed of human fibroblasts in a bovine Type I collagen lattice [111]. Epicel (Genzyme Biosurgery) is a sheet of skin cells ranging from 2 to 8 cell layers thick. The grafts are grown or cultured from a postage stamp sized sample of patient's own healthy skin, which is sent to GenzymeBiosurgery for processing. The cells within the epidermis of the skin sample are separated and grown by tissue culture. During this process, irradiated mouse 3T3 cells, are used to promote cell growth and to ensure that there will be a sufficient number of grafts available as soon as possible for treatment [112]. OrCel (Ortec International Inc) is a bilayered cellular matrix in which normal human allogeneic skin cells (epidermal keratinocytes and dermal fibroblasts) are cultured in two separate layers into a Type I bovine collagen sponge. Donor dermal fibroblasts are cultured on and within the porous sponge side of the collagen matrix while keratinocytes, from the same donor, are cultured on the coated, non-porous side of the collagen matrix [113].

4. TOPICAL ANTIBIOTICS

Widespread application of effective topical antimicrobial agents reduced the microbial load on the burn wound surface and reduced the risk of infection [33,114]. Selection of topical antimicrobial therapy should be based on the agent's ability to inhibit the microorganisms recovered from burn wound surveillance cultures. The chemical structures of the antibiotics that are particularly used for topical application in burns are shown in Fig. (7). Current patents describe the potential applications of these agents for wound dressings, bio-membrane, and wound coating material, postoperative adherent and cosmetic material applications. Burn units may rotate the use of various topical antimicrobial preparations to decrease the development of antibiotic resistance [115,116].

4.1. Mafenide Acetate

Mafenide acetate (Sulfamylon) cream has a broad spectrum of activity against gram-negative bacteria, particularly *Pseudomonas aeruginosa*, but has little activity against gram-positive aerobic bacteria such as *Staphylococcus aureus* [33]. Mafenide acetate 0.5% cream (mafenide) is a methylated topical sulfonamide compound. Since topical mafenide acetate cream can be used without dressings, so it allows open burn wound therapy and regular examination of the burn wound surface [117]. Mafenide was introduced prior to silver sulfadiazine and was widely used for the treatment of burns. A 5% solution must be applied to saturate gauze dressings. The solution appears to be as effective as the cream preparation when used in this way [118]. This agent should be used in combination with nystatin to prevent this complication due to its limited antifungal activity [119]. In burn patients with concomitant respiratory acidosis, the use of mafenide acetate over a large surface area of this compound can be fatal [120], because it's converted to *p*-sulfamylvanzoic acid, causing metabolic acidosis in the burn patient [33].

4.2. Bacitracin

Bacitracin is a mixture of related cyclic polypeptides produced by organisms of the licheniformis group of Bacillus subtilis var Tracy, isolation of which was first reported in 1945 [34]. It is a good alternative to silver sulfadiazine (Silvadene) for burn patients with a sulfa allergy. Bacitracin zinc is effective to promote regenerative healing when topically applied to wounds in concentrations from about 5% to 8%. The bacitracin zinc can be applied in a hydrophilic or hydrophobic carrier, and is advantageously impregnated in an absorbent pad [121].

4.3. Mupirocin

Mupirocin is a fermentation product of *Pseudomonas fluorescens* [35], which was a potent inhibitor against gram-positive skin flora such as coagulase-negative staphylococci and *Staphylococcus aureus* [122,123]. Pharmaceutical compositions comprising mupirocin and chlorhexidine are of use in treating topical bacterial infections, in particular infected burn injuries [124]. Mupirocin has also increasingly been used as a burn wound topical agent in North America, where MRSA has become a problem [125,126]. Various topical antibiotic preparations, including 1% silver sulfadiazine, 2% mupirocin, and 2% fusidic acid, were recently compared for their antibacterial effect in an MRSA-infected rat burn model [125]. Therefore, hospitals where MRSA is a problem may rotate the use of topical mupirocin in combination with these other agents to decrease the development of resistance.

4.4. Neosporin

Neosporin is a broad spectrum antibiotic ointment containing three different antibiotics: bacitracin, neomycin, and polymyxin B, in a petroleum jelly base. A study in total of 1053 patients with superficial burn injury, in which qualitative analysis showed Staphylococcus

aureus and Pseudomonas spp. to be the most common infecting organisms. Quantitatively, fewer patients showed infection on the 7th and 18th day post-treatment in the povidone iodine plus neosporin group (PVP + N) than in silver sulphadiazine group (SSD). Similarly, healing times were also better with PVP + N, with a maximum number of patients having healed within 15 days. However, the mortality rates were not much different between the two groups [36].

4.5. Polymyxin B

Polymyxin B is a simple, basic peptide antibiotic that is generally available in an ointment base (white petrolatum). Interaction of polymyxin B with phospholipids allows penetration and disruption of bacterial cell walls and therefore affects membrane permeability [37]. It was reported that Polymyxin B inhibited the ability of PMNs to destroy ingested microorganisms, which may suppress the ability of the wound to restrain bacterial proliferation [127]. Considering the side effects, the absorption of topically applied polymyxin B is negligible, and systemic toxicity is rare and is often related to use over a large area for an extended period of time. If with bacitracin, polymyxin B encourages healing by containing surface bacteria.

4.6. Nitrofurazone

Nitrofurazone is a topical anti-infective agent effective against gram-negative and grampositive bacteria. Treatment of nitrofurazone cream for patients with invasive Nitro-bacteria cloacae burn wound sepsis resulted in an overall 66% survival compared with an 86% mortality reported in the literature [38]. A nitrofurazone-impregnated catheter was introduced exclusively for use in burn patients [128]. The nitrofurazone-impregnated catheter was effective in reducing the incidence of catheter-associated urinary tract infection in a regional Burn Center. Use of these catheters may play an important role in minimizing the risk of urinary tract infection in burn patients. In an era when concern about bacterial resistance to many antiinfective agents is growing, the nitrofurans have continued to be active against organisms that have developed resistance to antibacterials [129].

4.7. Nystatin

Nystatin is an effective fungicide against Candida. Nystatin is produced by *Streptomyces noursei* and has its potent antifungal effect by binding to the sterols in the fungal cell membrane [35]. A lower concentration of this agent inhibits *Candida albicans*, but a higher concentration is needed to inhibit other fungi [130]. The nystatin can be encapsulated within a liposome, preferably a stable multilamellar vesicle, which broadly comprises one or more lipids one or more of phosphomonoglyceride, phosphatidic acid and sphingolipid [131]. However, since nystatin has no activity against bacteria, it should be used in combination with a topical agent that has activity against the broad spectrum of pathogenic bacteria that cause burn wound colonization and infection [33]. Furthermore, a Nystatin formulation was described as having reduced toxicity in a new patent [132].

5. TOPICAL SILVER PREPARATIONS

The medical uses of silver date back to at least the time of Hippocrates (the "father of modern medicine" [133]), who in his writings discussed silver's role in wound care. By the late nineteenth and early twentieth century, surgeons routinely used silver sutures to reduce the risk of postoperative inflammation and infection [134]. During World War I, soldiers used silver leaf to limit and treat battle wound infections [135]. Silver fell into disuse around World War II due to the advent of antibiotics. Interest in the antimicrobial properties of silver was only reinvigorated in the year 1965 by Moyer who published a paper on silver nitrate's antibacterial effects [39]. Today, silver has reemerged as a clinical treatment for the prevention and treatment of infections encountered in wounds, ulcers, and burns [136].

Many different silver containing antibacterial agents exist in the market, including topical creams, ointments, and solutions: silver nitrate, silver sulfadiazine (SSD), SSD with cerium nitrate, and newer sustained release silver products [136]. Silver requires ionization to fully exhibit its antimicrobial effect; positively charged silver ions readily bind to negatively charged proteins within the wound fluid, inhibiting delivery of silver ions to the wound bed. Indeed, a major problem with topical silver agents lies in their inability to penetrate deeply into the tissue and the low levels of silver release into the target area. In the last few years, several silver products have been introduced to the market in order to address these issues [137]. Fig. (7) graphically illustrates the use of silver as a topical antimicrobial agent in burns.

5.1. Silver Nitrate

Colloidal silver solutions consist of submicroscopic positively charged silver particles suspended in liquid base. These were commonly used up until Moyer's article was published in the 1960's [39], following which silver salts replaced colloidal silver solutions in clinical use. Silver salts consist of positively charged silver ions coupled to negatively charged ions (AgNO₃), an arrangement that is much more stable than that of colloidal solutions. Silver nitrate is the most commonly used silver salt, exerting an antibacterial action (*in vitro* and *in vivo*) at a concentration of 0.5% and a significant toxic effect at a concentration greater than 1% [138]. Silver nitrate is most commonly administered topically by soaking gauzes, which are then applied to severe burns. Patents have been issued for silver nitrate gels, which are claimed to be easier to administer [139–141].

The use of silver nitrate in burn patients remains controversial [40]. Nitrate is toxic to tissues and wounds, and decreases wound healing. Reduced nitrate impairs wound re-epithelialization, thereby partially offsetting the benefits of silver. (Interestingly, a patent was issued to exploit silver nitrate's toxicity as an intrauterine cauterizing agent [142]).

Other clinical uses of silver nitrate, aside from burns, exist. One percent silver nitrate drops were traditionally administered to newborns to prevent ophthalmia neonatorum (neonatal conjunctivitis). Many hospitals, however, have opted to use antibiotics (such as erythromycin drops) instead, due to fears of chemical irritation (and associated chemical conjunctivitis) following silver nitrate administration. Despite this well-founded concern, Zanoni *et al.* [143] found silver nitrate drops to be more effective in the prevention of ophthalmia neonatorum.

5.2. Silver Sulfadiazine

As discussed above, the use of silver fell into disuse with the advent of antibiotics. The renewed interest in silver comes as a consequence of the rapid development of bacterial resistance. In the 1970s, Fox [144] decided that it was unnecessary to choose between silver and antibiotics; he combined silver nitrate with the antibiotic sodium sulphadiazine. This formulation, known as silver sulfadiazine (Flammazine^R Silvadene^R), benefits from the inhibitory effect of silver and the antibacterial action of sulphadiazine [136]. Silver sulfadiazine (SSD) was initially formulated as an ointment, but was later incorporated into a hydrophilic cream [40]. Although SSD remains the gold standard in topical burn treatment, recent studies indicate that (similar to silver nitrate) SSD may delay wound healing. Cho Lee *et al.* [145] found that epidermal growth factor (EGF) could significantly counter the deleterious effects of SSD on wound healing, and concluded that future preparations of SSD may come coupled with EGF. Another problem associated with SSD remains bacterial resistance to the sulphadiazine component [146]. SSD-resistant Pseudomonal species have been reported; a patent was issued for a formulation to combat these species, a combination of SSD and sodium piperacillin (the latter of which is very active against pseudomonas) [147]. Other patents have been issued for various

SSD delivery systems, including animal tissue dressings [148], water-dispersible hydrophilic carriers [149], and topical spray preparations [150].

5.3. Cerium Nitrate-SSD

In 1976, Monafo *et al.* added cerium nitrate to SSD in the treatment of burn wounds, and concluded that the two ingredients had a synergistic effect [42]. Since that time, cerium nitrate-SSD (Flammacerium) has been routinely used in wound care. However, the efficacy of this combined formulation is disputed [40]. In 2005, Garner *et al.* conducted a review of pertinent literature and concluded that cerium nitrate-SSD reduces mortality and morbidity in severe burns when immediate excision and closure is not possible [151]. However, contrary to the hypothesis furthered by Monafo, Garner *et al.* concluded that cerium nitrate possesses minimal antimicrobial action, and gains its benefit from its action on burn eschar. Burned skin produces a lipid protein complex that causes immunosuppression in the burn patient, which leads to increased susceptibility to infection (and subsequent morbidity and mortality). Cerium nitrate binds to and then denatures this lipid protein complex, thereby preventing immunosuppression [151]. Patents have been issued for various delivery systems for cerium nitrate-SSD, including a creamlike water-dispersible hydrophilic composition [152].

5.4. Sustained Silver Releasing Systems

Silver nitrate, SSD, and cerium nitrate-SSD are considered the old guard in silver products. These are solutions, salts, or compounds that must be applied to gauze. Silver-based dressings are newer products that incorporate silver in the dressing itself [43]. In the last few years, there has been a proliferation of such products, accompanied by aggressive marketing campaigns by manufacturers. Patents have been issued for various silver dressings, including such products as ActicoatTM, Aquacel-AgTM, SilvercelTM, and ContreetTM.

Acticoat-7 (Smith & Nephew, Hull, United Kingdom) dressing utilizes nanotechnology: nanocrystalline (<20 nm diameter) silver crystals are released from the dressing to the burn wound [43]. The silver crystals are released as an initial large-bolus, followed by a sustained release. Fong *et al.* [43] cautiously concluded that nanocrystalline (NC) silver not only reduces wound infection and promotes wound healing, but decreases the frequency of dressing changes, reduces pain levels, and cuts cost. A meta-analysis by Gravante *et al.* [153] confirmed these findings, and found NC silver treated patients to have a significantly lower incidence of infections than those treated with the older silver products (silver nitrate and SSD). It was concluded on this basis that NC silver possessed a significantly stronger antimicrobial action than older silver formulations. Numerous patents [154–157] have been issued for various NC silver dressings and topical preparations that were developed by Nucryst Pharmaceuticals Corp. which was subsequently taken over by Smith and Nephew. Despite the encouraging results of NC silver products, concern remains over toxicity of nanocrystalline products in general which has been demonstrated *in vitro* (but not *in vivo*) [43].

Several other sustained silver releasing products exist, such as Aquacel Ag[®] (ConvaTec, Princeton, NJ, USA). Ionic silver is incorporated into the hydrofiber wound dressing, which turns into a gel upon contact with wound tissue. Barnea *et al.* [44] presented a review of literature, and determined that Aquacel Ag is both effective and safe in acute and chronic wounds, exhibiting a broad antimicrobial profile with no delay in wound healing.

Silvercel[®] (Johnson and Johnson) is another sustained release product, which combines the antimicrobial effects of silver with alginate's benefits [45]. Alginate is a polysaccharide found in the cell walls of algae that is able to absorb 200–300 times its weight in water, a capability that makes it particularly useful in exudate management of wounds. Bell *et al.* (2007) [158] compared Silvercel with Aquacel Ag, and found the former superior in its ability to manage

exudate, hardly a surprising finding considering alginate's powerful absorptive capabilities. Silvercel was able to maintain its shape and strength even in profusely wet wounds, unlike Aquacel Ag. On the other hand, Aquacel Ag-treated wounds were less likely to suffer from the adverse effects of excessive dressing adherence to wound tissues.

A patent was issued in 2003 for silver alginate foam [159]. Foams have emerged as another delivery system for sustained silver release, providing a simple and more convenient method of application. Jorgensen *et al.* [42] found Contreet Foam (sustained silver release foam, Coloplast) to be superior to Allevyn (hydrocellular foam dressing without added silver, Smith and Nephew), resulting in 45% greater reduction in ulcer area as compared to 25% in Allevyn.

Patents have been issued for numerous sustained silver release delivery preparations [160–163], and such formulations remain an area of active research and development.

5.5. Silver-Impregnated Biological Material

During the 1970's, human amniotic membrane emerged as a useable dressing in burn patients. Sawhney [46] concluded that this biological dressing was superior to conventional dressing, as it promoted rapid re-epithelialization and healing of wounds. Since 1978, silver has been incorporated into amniotic membrane, producing significantly better results than amniotic membrane alone [164,165]. A patent was issued for a wound dressing consisting of animal tissue (including amniotic membrane) incorporated with silver sulfadiazine; it was claimed that more silver was imparted onto the tissue than would be with other silver containing dressing [166]. The incorporation of silver into amniotic based membrane adds another dimension to the proliferation of silver containing products. EZ Derm (Brennen Medical, Inc) is modified pigskin impregnated with a soluble silver compound intended for treatment of burns. Originally developed by Genetic Laboratories. A patent describes the silver that EZ-Derm uses [167].

6. IODINE PREPARATIONS

6.1. Povidone-lodine

Iodine and its antibacterial properties have been used for the prevention or management of wound infections for over 150 years [168]. However, the use of solutions (tincture) of iodine has been replaced by the widespread use of povidone-iodine, a water-soluble compound, which is a combination of molecular iodine and polyvinylpyrrolidone [169]. Povidone- iodine's antiseptic action is due to the available iodine present in the complex. This solution is active against a wide variety of bacteria, fungi, protozoa, and viruses [170]. A patent on wound-healing compositions containing povidone iodine describes admixtures of an antifungal/ antibacterial agent such as povidone iodine, sugar and a suitable carrier as substantially non allergenic, having excellent healing properties when applied to burns or open wounds and serves as an effective barrier to growth of healing tissues in gauze or similar dressing [47]. A device has been described in a recent patent [171] for the in-situ preparation of a pharmaceutical composition comprising povidone-iodine and a steroid or non-steroidal anti- inflammatory.

6.2. Liposomal Iodine (Repithel)

The combination of povidone-iodine (PVP-I) and liposomes unites the microbicidal activity of the antiseptic substance with the tolerability and lack of immunogenicity of liposomes; in addition, liposomes provide a moist molecular film for the wound environment. An animal study on the efficacy and tolerability of different formulations of a hydrogel with PVP-I liposomes in deep dermal burn wounds has indicated a good quality of wound healing with smooth granulation tissue, less inflammation, less wound contraction and no hyperkeratotic reactivity, especially with the 3% PVP-I liposome formulation [49]. Repithel (Mundipharma Research GmbH & Co. KG, Limburg/Lahn, Germany) is a new liposomal hydrogel

formulation of PVP-I developed to provide a moist wound-healing environment with antiinfective and debriding properties. The pronounced positive effect of Repithel in burns in smokers (since smoking and excessive inflammation have a similar impeding effect on wound healing) [172]. There is a patent [173] describing a dry liposomal PVP-I composition which was directed towards a storage stable package.

6.3. Cadexomer lodine (lodosorb)

Cadexomer iodine (Iodosorb, Smith and Nephew) is a hydrophilic starch powder containing iodine, which is a suitable dressing for granulating wounds such as venous ulcers. Formulations of iodophors or "iodine carriers," however, release low levels of iodine over a longer period. Cadexomer iodine is a slow-release antimicrobial capable of absorbing excess wound exudates while offering a sustained level of iodine in the wound bed. Cadexomer iodine also has established effectiveness *in vivo* against *S. aureus* and MRSA. Iodine needs moisture to be activated, similar to silver ion-containing products. A total of 61 outpatients with chronic venous ulcers participated in a randomized optional crossover trial using cadexomer iodine or a standard dressing for their ulcers. Both treatments were highly effective, but the epithelium of ulcers dressed with cadexomer iodine grew again significantly faster [174]. A recent patent describes an improved method for the preparation of cadexomer iodine [175].

6.4. Other lodine Preparations

A patent describes iodine-containing derivatives, including, free iodine and iodoform for otic and nasal infections [176]. lodoform is a potent germicidal agent and the composition also contains one or more anti-inflammatory agents and one or more natural or synthetic compounds which provide analgesic benefits. Another patent (3M Innovative Properties Company, Saint Paul, MN) [177] describes liquid antiseptic compositions containing iodine and a sugar and/ or sugar alcohol intended primarily for tissue antisepsis. Iocide[™] (Biomedical Development Corporation, San Antonio, TX) is composed of a monobasic iodide salt, an organic acid having up to eight carbon atoms (such as citric acid, ascorbic acid or oxalic acid), an oxidizing agent (such as sodium percarbonate or sodium perborate) and an aqueous buffer [50]. Mixing these ingredients together generates iodine in a biologically compatible antimicrobial formulation. Another patent [178] described a stable, aqueous, iodine-based germicidal composition including iodine, a non-ionic surfactant (e.g. polyoxyethylene, polyoxypropylene block copolymers) and an iodine-solubilizing halide ion (chloride or bromide).

7. PHOTODYNAMIC THERAPY

Photodynamic therapy (PDT) [179] was discovered over 100 years ago by observing the killing of microorganisms when harmless photosensitizers (PS) and visible light were combined in vitro [180]. Light of the correct wavelength excites the PS to its excited singlet state that can then undergo intersystem crossing to the long-lived excited triplet state. In the presence of the oxygen, the triplet state PS transfers energy to ground state molecular oxygen (a triplet) producing reactive oxygen species (such as singlet oxygen) that are able to kill microbial cells [181]. Fig. (9). graphically illustrates the mechanism of action of antimicrobial PDT and emphasizes its broad spectrum activity due to the ability to effectively kill or inactivate all known classes of microbial pathogen. Since its discovery PDT has primarily been developed as a treatment for cancer, ophthalmologic disorders and in dermatology [179]. However, in recent years interest in the antimicrobial effects of PDT has revived due to the inexorable increase in drug resistance among many classes of pathogen [182]. Numerous investigations have been performed [183] to develop and refine the PS with particular molecular properties that are able to selectively inactivate target microbial cells with minimal damage to host cells. The molecular structures of several of these PS that are discussed in the present section are shown in Fig. (10). Most of the powerful antimicrobial PS have an overall positive or cationic

charge that may be provided by quaternized nitrogen atoms (with a constitutive cationic charge) or by basic amino groups with a pKa that allows them to be positively charged at the pH values that prevail in bacteria or in infected tissue.

7.1. Patents on Antimicrobial PDT

Nifantiev *et al.* [184] patented molecular conjugates with photosensitizers (PS) which can bind selectively to the surface of a microorganism bringing about photo-destruction upon irradiation. The key to these conjugates is a special spacer connecting at least one PS to a microorganism receptor (vector). Spacers having hydrophilic structure, such as ethylene glycol units and amino carboxyl end capped ethylene glycol units, are used for linking the vector to the PS. Bacterial species *S. aureus*, *P. aeruginosa*, and *E. coli* were used to test the effectiveness of the conjugates. The conjugates were found to be very effective in inactivating these bacterial species in the real patient-related environments where blood, serum and other body fluids are always present.

Hasan *et al.* patented [185] another group of PS conjugates for antimicrobial PDT. These involved covalently attaching a PS to a "non-pair member" (NPM) moiety. This NPM can be a linear, branched, or cyclic polypeptide including a small anti-microbial peptide (AMP) (see later section of this review). Histatins, defensins, cecropins, magainins, Gram positive bacteriocins, and peptide antibiotics can be attached to PS. Moreover the polypeptide can also be a poly-L-lysine, polyornithine or polyarginine molecule whose length and substitution ratio can vary [186,187]. A further variation on the NPM can be polymers that are basic but not composed of amino-acids such as polyethylenimine (PEI, see Fig. 10). These conjugates were originally proposed for oral anti-bacterial applications (PDT for periodontitis [188]), but subsequent studies showed that PEI-ce6 was highly effective for treating burn infections in a mouse model [52].

Hamblin *et al.* [54] patented PS compounds based on functionalized fullerenes in antimicrobial PDT. In this invention, fullerene molecules, higher fullerenes, and their functionalized derivatives are modified to include a variety of properties needed for application of PDT to microbial cells. This is achieved by controlling hydrophobicity, molecular charge, and water solubility of the carbon nanomaterial specifically to target microbial species preferentially over other types of cells for PDT. A positive charge allows the fullerenes to bind to cells and overcome microbial permeability barriers. Cationic fullerenes in particular performed better as antimicrobial PS than the widely employed antimicrobial PS toluidine blue O. Accordingly, cationic fullerenes may find significant application for PDT of a wide variety of conditions, such as for example, localized infections in wounds, burns, and mucus membranes.

In another patent, Hamblin and Tegos [189] described the use of phenothiaziniums and microbial multi-drug resistance inhibitors to inactivate microbial cells. It has now been shown that phenothiaziniums are substrates for microbial multidrug resistance pumps ("MDR Pumps"). Inactivation of the microbial MDR pumps by inhibitors ("MDR inhibitors") increases the amount of phenothiazinium that can be retained by the microorganism, thereby increasing efficacy upon photoactivation of the phenothiazinium by irradiation.

Biel [190] filed a patent utilizing a combined solution of a PS (e.g., methylene blue or toluidine blue) and a surfactant compound (e.g. polymyxin B or SDS) for PDT against bacterial or fungal infections, or for cancer cell eradication. Surfactants are used to affect the permeability of the membranes of the target cells. The treatable pathogens include *S. aureus, C. albicans, E. coli, Enterococcus, Streptococcus, P. aeruginosa, Hemophilus influenza*, and *Clostridia*.

Hamblin and Foley [58] patented a group of PS composed of chalcogen analogs of benzo(A) phenoxazinium dyes that are effective antimicrobial PS. In particular when the chalcogen is selenium (EtNBSe) the activity is maximized (Fig 10).

Albrecht *et al.* [191] disclosed a method of destroying bacteria in a treatment area of a patient by introducing a composition comprising Erythrosin B as a PS in a gel to a treatment area on a biological surface, allowing a predetermined time period to elapse to allow Erythrosin B to couple with bacteria in the treatment area, and applying radiation of a preselected wavelength to the treatment area to activate the PS and thus stimulating a photodynamic reaction to destroy bacteria.

In another patent, Albrecht *et al.* [192] described the use of liposomal formulations for improved efficacy of antimicrobial PDT. The formulations comprise a hydrophobic porphyrin PS, a monosaccharide, and one or more synthetic phospholipids, which are stable in storage especially through freeze-drying process and reconstitution.

Van Der Haas *et al.* [193] patented a method of preparing quaternized porphyrin derivatives suitable for antimicrobial PDT. A further patent [194] described the use of these porphyrins typified by Sylsens B (Fig 10) for carrying out PDT of fungal skin infections including burns.

Roncucci *et al.* patented [57] a series of phthalocyanine derivatives Fig. (10) having photosensitizing characteristics and high solubility in water, useful for photodynamic treatment of bacterial infections, in particular infections generated by Gram-negative bacteria.

In another patent, Bommer and Jori [195] proposed the use of porphyrins that are characterized by the presence of up to four positive charges in the peripheral substituent, and at least one hydrophobic tail comprising between 6 and 22 carbons inclusive, originating at one or more of the charged sites. *In vitro* test was carried out to evaluate the photosensitizing efficacies of the compounds. It was observed that a $4-\log_{10}$ -unit inactivation of *E. coli* was achieved upon 15 minutes irradiation in the presence of 1 μ M porphyrin.

Love and his colleagues [53] patented a method of using porphyrin derivatives containing cationic groups for selective photodynamic inactivation of target microbial cells with minimal damage to host cells. It was claimed that the porphyrin derivatives typified by XF73 Fig. (10). can be used as selective PDT agents, i.e., for selectively killing microorganisms, and can also be used in combination with conventional antibiotics.

7.2. PDT for Burn Infections

The first paper describing PDT for burn infection was by Orenstein *et al.* [56]. These workers used deuteroporphyrin-hemin complex as an agent for PDT of burn wounds infected with a multiple-drug resistant strain of *Staphylococcus aureus*. The *in vivo* experiments by application of the above porphyrins in combination to infected burn wounds in guinea pigs was an effective way to reduce dramatically the contaminating *S. aureus*. Reduction of more than 99% of the viable bacteria was noted after the porphyrin mixture was dropped on the eschar or injected into the eschar, an effect that lasted for up to 24 hours.

Lambrechts *et al.* [55] used PDT mediated by meso-mono-phenyl-tri(*N*-methyl-4-pyridyl)porphyrin (Sylsens B, Fig. 10) to treat burn wounds in mice with established bioluminescent *S. aureus* infections. PDT was applied after one day of bacterial growth by adding a 25% DMSO/500 microM PS solution to the wound followed by illumination with red light and periodic imaging of the mice using a sensitive camera to detect the bioluminescence. More than 98% of the bacteria were eradicated after a light dose of 210 J cm² in the presence of PTMPP. However, bacterial re-growth was observed. Dai *et al.* [52] evaluated the effectiveness of PDT for *A. baumannii* burn infections using mouse models. Burns were created on the backs of shaved mice by using two heated brass blocks (95 °C) with contact time between the blocks and mouse skin 10 seconds. Bioluminescent *A. baumannii* were inoculated to the burns, followed by addition of poly-ethylenimine-chlorin e6 conjugate and subsequent illumination with non-coherent red light at 660±15 nm.

Figure 11 shows the PDT dose-response of the bacterial luminescence of a representative mouse burn infected with *A. baumannii* and treated with PDT at 30 minutes after infection, a mouse burn treated with PS only (dark control), and a mouse burn treated with light only (light control). After 240 J/cm² red light had been delivered (40 minutes irradiation time at an irradiance of 100 mW/cm²), PDT induced an approximately 3-log-unit reduction in bacterial luminescence from the mouse burn, while during the same period of time, only modest reduction of bacterial luminescence was observed in the dark control and the light control. When PDT was performed at 30 minutes after infection, over $3-\log_{10}$ inactivation of *A. baumannii* was achieved, as quantified by luminescent imaging analysis. When PDT was performed 24 hours after infection (the time when the infections were fully established), over $1 \log_{10}$ inactivation was achieved.

PDT did not delay wound healing in A. baumannii infected burns.

8. CHITOSAN PREPARATIONS

Chitosan is a polycationic polymer with a specific structure and properties. Chitosan molecules contain more than 5000 glucosamine units and the substance is obtained commercially from shrimp and crabshell chitin (a N-acetyl-glucosamine polymer) by alkaline deacetylation (NaOH, 40–50%) as shown in Fig. (12). Chitosan preparations of various molecular weights, degrees of deacetylation (DD), and with further molecular derivatization patterns have attracted much attention because of their potentially beneficial biological properties. Chitosan's properties allow it to rapidly clot blood, and has recently gained approval in the United States for use in bandages and other hemostatic agents. Chitosan is a polymer with a number of basic amino groups and hence possesses an overall cationic charge especially at acidic pH. In common with many polycationic polymers it has pronounced antimicrobial effects due to destabilization of the outer membrane and pemeabilization of the plasma membrane [196].

8.1. Patents for Antimicrobial Chitosan Preparations

A large number of patents have been filed or issued concerned with the preparation and use of various chitosan preparations as antimicrobial burn treatments or dressings. In this review we can only cite a few of them.

One of the first patents was by Sparkes *et al.* [197] who described a wound dressing comprising a blend of gelatin and chitosan. Yura and colleagues patented [198] a functional chitosan derivative comprising chitosan, incorporating a photo-reactive functional group, an amphipathic group, e.g. a polyoxyethylene alkyl ether, and a glycosaminoglycan and that has solubility in a neutral medium, self-crosslinking ability, and able to act as a wound dressing. Baker and Wiesmann [199] filed a patent for chitosan or chitosan derivatives such as chitosan-arginine and chitosan-acid amines, that exhibit bactericidal activity against bacterial pathogens, e.g., drug resistant bacteria such as Methicillin-resistant *S. aureus* (MRSA). Scherr patented [200] a foamed gel containing chitosan that may be layered onto a suitable backing for use as a wound dressing, or the gel may be directly applied to wounds to effect hemostatic activity as a result of the action of the chitosan. The composition of the foamed chitosan gel can reduce the risk of microbial infections in wounds. Muzarelli patented [201] a methyl pyrrolidinone chitosan derivative that could be used as a wound dressing material in the form of a freeze-dried material, powder, film, non-woven fabric, bandage, adhesive tape etc.

8.2. Chitosan for Burn Infections

Alsarra [202] investigated the formulation of chitosan gel for improving wound healing of burns in rats. High molecular weight chitosan with a 2,000,000 MWt and 92% DD, medium molecular weight chitosan with a 750,000 MWt and 75% DD, and low molecular weight chitosan with a 70,000 MWt and 63% DD were tested. It was shown that chitosan samples with high molecular weight and high degree of deacetylation had the best effect for the healing of burns.

Aoyagi *et al.* [203] described a chitosan dressing composed of chitosan film and minocycline hydrochloride. Chitosan with DD of 67%, 83% and 96% (mol/mol) were used, respectively. Various formulations were applied to burns in rats. Chitosan with DD of 83% containing 2 mg minocycline hydrochloride showed an excellent effect.

Boucard *et al.* [204] developed a bi-layer physical hydrogel, which consisted of chitosan and water. The first layer was a rigid protective gel that ensured good mechanical properties and gas exchanges, and the second soft and flexible layer allowed the material to follow the geometry of the wound and ensured a good superficial contact. A chitosan with 2.6% DD and molecular weight of 540,000 g/mol was used. This material was tested for skin regeneration after third-degree skin burns on the pig back. The results showed that chitosan materials were well-tolerated and promoted good tissue regeneration.

Cardenas [205] made several types of chitosan composite films by adding several additives to acetic acid chitosan solutions, such as: glycerol, oleic acid and linoleic acid in different proportions. They used chitosan acetate films, which were prepared using shrimp chitosan of low and high molecular weight (MWt = 68,000 g/mol and MWt = 232,000 g/mol). The films exhibited different physical properties depending upon the additives and/or mixture of them. The addition of glycerol to composite improves the elasticity of the films. In-vitro tests were carried out using the chitosan composite solution against *S. aureus*, *P. aeruginosa*, and *A. baumannii*, showing that the molecular weight has an influence on the bactericidal properties of the chitosan composite films and on its effect against Gram positive and Gram negative bacteria. Medical applications of the composite films were studied in patients with burns, ulcers and injuries, the films containing glycerol showed good adhesion in comparison with those without it. Excellent results in skin recovery were obtained after 7–10 days.

Ribeiro *et al.* [59] proposed a chitosan hydrogel for wound healing of burns. By preparing this hydrogel, chitosan solution 4% (w/w) was dispersed in lactic acid 2% (v/v). *In vitro* studies showed that chitosan-hydrogel was able to promote cell adhesion and proliferation, and are non-cyto-toxic to cells. *In vivo* studies using a rat burn model revealed that chitosan-hydrogel stimulated the wound healing, and was locally and systemically biocompatible based on the evidence that there was no granulomatous inflammatory reaction in burns treated with chitosan-hydrogel and no pathological abnormalities in the organs.

Sezer *et al.* [206] developed a chitosan film containing fucoidan and investigated its effectiveness for treating burns in rabbits. Fucoidan is a sulfated polysaccharide commonly obtained from seaweeds and shows significant gel contraction-promoting, integrin expression-enhancing, and heparin activities. It was found that chitosan film containing fucoidan induced significant wound contraction, and accelerated the wound closure and healing process. The authors also prepared a fucoidan-chitosan hydrogel and investigated its treatment efficiency on dermal burns in rabbits [60]. Similar results were obtained to those with chitosan film containing fucoidan.

Damour et al. [207] reported a clinical use of a dermal substrate made of collagen--GAG-chitosan grafted immediately after early excision of severe burns, then epidermalized either

with autologous meshed autograft or with autologous cultured epidermis. The dermal substrate replaces the excised dermis by adhering to the underlying tissue, promoting fibrovascular ingrowth. In comparison to homograft, this dermal substrate is always available, can be stored, and is exempt from micro-organism transmission.

Peh *et al.* [208] investigated the mechanical, bioadhesive strength, and water vapor permeability of chitosan films prepared using two different solvents, acetic acid (chitosan-AA) and lactic acid (chitosan-LA). The properties of the two films were compared with those of a commercial preparation, Omiderm®. In addition, biological studies were conducted to investigate the skin irritation and systemic toxicity of the films. It was found that the three preparations differed significantly in terms of mechanical and bioadhesive strength properties. Chitosan-LA was more soft, flexible, pliable, and bioadhesive than chitosan-AA. Furthermore, chitosan-LA was non-allergic and non-toxic. As a result, the author concluded that chitosan-LA is more suitable to be used in the management of wound healing and skin burn.

Dai *et al.* [61] investigated the use of a freeze-dried chitosan acetate bandage (HemConTM) that was originally developed as a hemostatic dressing. They applied the dressing to 3rd degree burns in mice that had been infected with bioluminescent *P. aeruginosa and Proteus mirabilis*,

Figure (13). shows the successive bioluminescence images from day 0 to day 3 after *P. aeruginosa* infection of an untreated burn, a silver dressing-treated burn, and a chitosan acetate-treated burn, respectively. In the untreated burn, the bacteria multiplied approximately 1000 fold from day 0 to day 3, while in both the silver dressing and chitosan acetate bandage-treated burns, a decrease in bioluminescence signal was seen at day 1 as the dressing immediately quenched the light. There was a detectable increase in signal from the silver-dressing burn at day 2 and 3 (compared to day 1) that was not seen in the chitosan-acetate burn. The inset showing a silver-treated burn at day 3 was taken after the silver dressing was removed from a dead mouse and shows a vigorous bacterial proliferation underneath the dressing.

In the case of *P. aeruginosa* infections, the survival rate of mice treated with the chitosan acetate bandage was 73.3%, whereas the survival rate of mice treated with a nano-crystalline silver dressing was 27.3% (p = 0.0055) and that of untreated mice was 13.3% (p < 0.0002). For *P. mirabilis* infections, the comparable survival rates were 66.7%, 62.5%, and 23.1% respectively. Quantitative bioluminescent signals showed that the chitosan acetate bandage effectively controlled the growth of bacteria in the burn and prevented the development of systemic sepsis, as shown by blood culture.

9. ANTIMICROBIAL PEPTIDES

The term 'antimicrobial peptides' refers to a large number of peptides found among all classes of life possessing both antibacterial and antifungal activities. They have been shown to kill Gram-negative and Gram-positive bacteria (including strains that are resistant to conventional antibiotics), mycobacteria (including *Mycobacterium tuberculosis*), enveloped viruses, and fungi. Besides acting as endogenous antibiotics AMPs also participate in multiple aspects of immunity (inflammation, wound repair, and regulation of the adaptive immune system). So unlike the majority of conventional antibiotics the antimicrobial peptides also have the ability to enhance immunity by functioning as immunomodulators. AMPs are divided into heterogeneous groupings on the basis of their primary and secondary structures, their antimicrobial potential, their effects on host cells, and the regulation of their expression. Most AMPs are small (12–50 aminoacids), have a positive charge provided by Arg and Lys residues, and an amphipathic structure that enables them to interact with bacterial membranes [209]. Fig. (14). illustrates the selectivity obtained by AMPs that have a higher affinity for microbial membranes compared to the membranes of host cells and therefore preferentially lyse the pathogenic, microorganisms.

These natural antimicrobials are a part of the body's natural defense mechanisms, and any loss of these compounds will obviously lead to a reduced ability to combat infection and predispose to sepsis. There are many reports of the association of AMPs with different skin diseases. Overexpression of AMPs can lead to increased protection against skin infections as seen in patients with psoriasis and rosacea; inflammatory skin-diseases which rarely result in superinfection. In other skin diseases, e.g. in patients with acne vulgaris, increased levels of AMPs are often found in inflamed or infected skin areas indicating a role of these peptides in the protection from infection. In addition, decreased levels of AMPs are associated with burns and resulting sepsis is often fatal [210]. This observation suggests a possible therapeutic role for AMPs in the management of burn wounds.

9.1. Defensins

Defensins are a family of cationic antimicrobial peptides exhibiting bactericidal, fungicidal and antiviral activity [211–213]. They are part of a non-specific chemical defense system secreted by non-lymphoid cells [62]. When released into the extracellular milieu, they exert their antimicrobial activity directly by attacking the microbial membrane, and contribute to the oxygen-independent killing of phagocytosed microorganisms. Furthermore, defensins are mediators in the crosstalk between the innate and adaptive immune systems [214]. Two human beta-defensins have been identified. Human B-defensin-1 has been detected in urogenital, gastrointestinal and pulmonary epithelia and most recently in skin [215–217]. Studies have shown decreased levels of AMPs mainly defensin associated with burns [210]. which facilitates infection and subsequent sepsis. This reduction in defensins may also alter functions of T- and B-lymphocytes, neutrophils, macrophages, and complement, thereby contributing to the pathophysiology of burn-related systemic inflammatory responses [218]. The broad antimicrobial properties of defensins, their chemical resistance and their lack of antigenicity suggest that they may also be useful as topical or systemic agents for the prophylaxis and treatment of infections [62]. They may also protective role in preventing infection within burn wounds as altered expression of these peptides could promote local overgrowth of microorganisms.

9.2. Demegel

Some of the synthetic AMPs have increased potency, enhanced specificity, and reduced toxicity relative to natural defensins. These peptides are quite resistance to the effects of high solute levels, and demonstrate greater antibiotic activity than natural peptides. One such synthetic peptide is Demegel D2A21 (Demegen, Inc. Pittsburgh, PA), which has had patents issued [219,220]. This peptide has shown significant promise with *in vitro* studies against a large number of pathogens [221]. Chalekson *et al.* [63] demonstrated use of D2A21 in an infected burn model in improving survival and recovery. Later they compared the effect of this peptide on survival in an infected wound model with that of traditionally used wound therapies, silver sulfadiazine and sulfamylon and reported D2A21 demonstrated significant superior activity as compared with controls and standard therapy [222].

9.3. Histone H1.2

Histones are the most abundant proteins bound to DNA in eukaryotic cells and among the most evolutionary conserved proteins known. They are small basic proteins with a molecular weight between 11 and 20 kDa and they contain a high percentage of positively charged amino acids (circa 20%) such as lysine and arginine. Eukaryotic cells contain mainly five types of histones: histone linker H1 and core histones H2A, H2B, H3, and H4. Histone H1.2 is a member of the histone H1 family [223] which has a potential role in innate antimicrobial defense [224]. It has been demonstrated that histone H1.2 has good *in vitro* and *in vivo* (infected rat burn model

with *P. aeruginosa*) activity against a number of burn wound infection pathogens [64]. A patent was issued for the recombinant production of H1.2 [225].

9.4. Cecropin B

Cecropin B and related peptides have been patented as antibacterial agents [226]. The antibacterial effect of cecropin B on *P. aeruginosa* infection of wounds in mice was investigated and it was observed application of cecropin B could reduce the mortality of the mice [65]. HB-107, a peptwide fragment derived from cecropin B, lacks antimicrobial activity yet showed up to 64% improvement in wound closure at day13 when compared to either scrambled peptide or vehicle controls in murine model of aseptic full-thickness excisional injury. This effect was comparable to that seen after treatment with currently accepted therapy. Histological evaluation of wounds treated with HB-107 displayed keratinocyte hyperplasia and increased leukocyte infiltration compared to controls. In two different rat models used by Ghiselli *et al.* [227], beta-lactams even increased plasma endotoxin and TNF-alpha levels, whereas the combination with cecropin B was shown to be the most effective treatment, with the highest survival rates (more than 90%), the highest antimicrobial activities, and the strongest.

9.5. Recombinant Bactericidal/Permeability-Increasing Protein (rBPI21)

This recombinant peptide derived from a human neutrophil sequence was patented as an antimicrobial agent [228]. Early treatment with rBPI21may be effective in attenuating multiple organ damage resulting from gut-origin endotoxin translocation in case of thermal injury. This might be associated with the down-regulation effects of tissue lipopolysaccharide-binding protein and CD14 gene expression by the use of rBPI21. Currently, there is an ongoing clinical trial investigating therapeutic effects of rBPI21 in burn patients [66].

9.6. Ceragenins

Ceragenins, or cationic steroid antibiotics (CSAs), are synthetically produced small molecule chemical compounds consisting of a sterol backbone with amino acids and other chemical groups attached to them. They were described in a series of patents [229–231]. While CSAs have a mechanism of action that is also seen in antimicrobial peptides, which form part of the body's innate immune system, they avoid many of the difficulties associated with their use as medicines. Several of these derivatives exhibit antimicrobial activity against a broad range of bacteria. Although some forms of ceragenins are effective against both Gram-negative and Gram-positive bacteria, they are generally more potent against Gram-positive bacteria. Ceragenins have the unusual property of forming complexes with phospholipids. Thus ceragenins are a class of agents with many properties to make them favorable for application as antiinfective agents [67].

9.7. Other AMPs

A patent [232] described the house-fly (*Musca domestic*) pupa natural AMP as a product with broad-spectrum antibiotic activity and no hemolytic activity to blood erythrocytes of mice and was characterized by high temperature resistance, acid and alkali resistance and good stability.

Another patent related to multimeric forms of antimicrobial peptides, for example, defensin peptides that possess antimicrobial activity and may be formulated into antimicrobial compositions for coating medical devices and the like. The invention also relates to the use of these multimeric forms of peptides for inhibiting and/or reducing the growth of microorganisms in infections including burns [233].

Antimicrobial bolisin peptides were originally extracted from bovine thymus glands and were described in a patent [234]. Other patents have been issued based on antimicrobial peptides from histatin [235], silkworm hemolymph [236], heparin-binding protein or human neutrophil elastase [237–239].

10. MISCELLANEOUS TOPICAL ANTIMICROBIALS

Topical chlorhexidine has been studied in several formulations to combat burn infection. Macmanus *et al.* [68] studied chlorhexidine diphosphanilate 2% cream (CHP) in two rat models of fatal burn wound infection (20% scalds infected with *Pseudomonas aeruginosa* or *Proteus mirabilis*). It performed as well as silver sulfadiazine. Miller *et al.* [69] compared different concentrations of CHP in a clinical trial in 29 burn patients, each with two similar burns which could be separately treated. One burn site was treated with each of four different CHP concentrations, from 0.25 per cent to 2 per cent while the other site was treated with AgSD cream. There was a direct relationship between CHP concentration and patients' ratings of pain on an analogue scale. The 0.25 per cent CHP cream was closest to AgSD in pain tolerance. A number of patents have been issued covering the use of chlorhexidine as a topical antimicrobial agent for use in burns [124,240–242]

Superoxidized water is based on hypochlorous acid obtained by the electrochemical treatment of a saline solution, and was disclosed in a US patent [70]. Preferably, the pH of the super-oxidized water is in a range of 4 to 7, and the water has a redox potential of > 950 mV. Medicaments based on the super-oxidized water may be in liquid or gel form and can be applied to control the microbial population within the wound or burns and at the same time permit cell proliferation.

An antimicrobial nanoemulsion was disclosed in another US patent [243]. It is composed of an oil-in-water nanoemulsion consisting of about 5% by volume of a surfactant such as polyoxyethylenesorbitan monolaurate or polyoxyethyl-enesorbitan monooleate with 8% ethanol, 64% oil, 1% cetylpyridinium chloride and distilled water. Burn infections were among other applications proposed.

FPQC (1-ethyl-6-fluoro-1,4-dihydro-4-oxo-7-[1-pipera-zinyl]-quinoline-carboxylic acid) [72] is a topical agent that has been tested in burned mice infected with resistant *Pseudomonas* strains; mortality in groups receiving topical therapy with FPQC was 0%, but 80% to 100% with sulfadiazine silver and 100% without treatment. Similar results were obtained in burned rats.

Acidfied nitrite has been shown to be an effective antimicrobial treatment for various skin pathogens including *C. albicans* and *S. aureus* [73]. It works by releasing nitric oxide and other reactive nitrogen intermediates locally in a phenomenon known as "nitrosative stress" [244].

Phage therapy was discovered as long ago as the 1920s [245], and has been considered to be a possible approach to burn infections. The disadvantage has been considered to be the narrow specificity of phages to particular species and strains of bacteria. McVay *et al.* [74] found that a cocktail of 3 phages could significantly decrease the mortality of thermally injured, *P. aeruginosa*-infected mice. Malik and Chhibber [246] demonstrated the protective effect of *K. pneumoniae*-specific bacteriophage KO1 isolated from the environment in a mouse model of burn wound infection induced by *K. pneumoniae*. A substantial decrease in the bacterial load of blood, peritoneal lavage, and lung tissue was noted following treatment with the bacteriophage preparation. A patent was issued for a cocktail of anti-*P. aeruginosa* phages for topical application [247].

Ipaktchi *et al.* [75] used topical application of an inhibitor of p38 mitogen activated proein kinase (MAPK) in a rat burn model infected with *P. aeruginosa.* They found that bacterial wound growth was significantly attenuated in animals treated with topical p38 MAPK inhibitor. They concluded that attenuation of excessive burn wound inflammatory signaling might prevent secondary damage of the dermal barrier and reduce the growth of opportunistic pathogens.

Probiotic therapy uses "good" bacteria to out-compete and in some cases actively kill harmful pathogenic bacteria. *Lactobacillus* sp. produce "lantibiotics", which are antimicrobial proteins belonging to the general class of bacteriocins. A patent was issued for a lantibiotoiic in combination with mupirocin as a topical antimicrobial [248]. A preparation consisting of *Lactobacillus plantarum* was equivalent to silver sulfadiazine in a clinical trial of patients with infected second- and third-degree burns [76].

Honey has long been known to be a naturally occurring antimicrobial preparation [249]. It is even considered to be a "folk therapy" for burns [250]. Its antibacterial properties are due to a combination of its high osmolarity, and the production of hydrogen peroxide that is formed in a slow-release manner by the enzyme glucose oxidase present in honey. It becomes active only when honey is diluted by wound fluid. The non-peroxide antimicrobial activity is due to methylglyoxal and an unidentified synergistic component. In a clinical trial [77] honey dressing was compared with boiled potato peel dressings as a cover for fresh partial-thickness burns in two groups of 50 randomly allocated patients. In the 50 patients treated with honey, 90 per cent of wounds were rendered sterile within 7 days, while the 50 patients treated with boiled potato peel dressings had persistent infection for 7 days. Of the wounds treated with honey, 100 per cent healed within 15 days as against 50 per cent in the wounds treated with boiled potato peel dressings. A number of patents have been issued related to topical application of honey as an antimicrobial dressing [251–255].

Another alternative burn therapy has been proposed to be plant essential oils [78]. Citricidal is a grapefruit seed extract that was patented as an antimicrobial perfume ingredient [256]. Patchouli, tea tree, geranium, lavender essential oils and Citricidal were compared against *S. aureus* in a burn dressing *in vitro*. Citricidal, geranium oil and tea tree oil were the most effective in combination.

Moist exposed burn ointment (MEBO) was developed from traditional Chinese medicine [257]. It contains sesame oil, beta-sitosterol, berberine, and other small quantities of plant ingredients. Berberine has long been known to be an antimicrobial compound [258]. MEBO has been tested in clinical trials for partial-thickness thermal burns covering less than 40% of body surface area and shown to be equivalent to conventional therapy [257].

Due to the limited resources for the management of burns in most regions of Africa there is a significant role for many aspects of traditional African medicine. Carica papaya [80] is currently used in The Gambia as the major component of burns dressings, where the pulp of the papaya fruit is mashed and applied daily to full-thickness and infected burns. It appears to be effective in desloughing necrotic tissue, preventing burn wound infection, and providing a granulating wound suitable for the application of a split thickness skin graft. Possible mechanisms of action include the activity of proteolytic enzymes chymopapain and papain, as well as an antimicrobial activity.

11. CURRENT & FUTURE DEVELOPMENTS

There are a large number of diverse topical antimicrobial agents that have been approved, tested or proposed for the prevention and/or treatment of burn infections. The particular pathophysiology of burns does predispose them to infection with a variety of pathogens, and

experience has shown that topical therapies produce better outcomes than traditional systemic antibiotics. Moreover the ever-increasing spread of multidrug resistance only serves to underline the importance of effective and safe topical therapies. The development of these topical antimicrobial approaches has been a fertile field for both small and large biotechnology companies as may be judged by the number of patents covered in this review. We believe that research in the area of topical antimicrobial agents will only continue to produce ever more effective hi-tech products for advanced societies as well as affordable and traditional preparations for the third world.

Acknowledgments

Research in the Hamblin laboratory is supported by the NIH (grant RO1AI050875), US Air Force MFEL program (contract FA9550-04-1-0079), Center for Integration of Medicine and Innovative Technology (DAMD17-02-2-0006), Congressionally Directed Medical Research Program (W81XWH-09-1-0514). T. Dai was supported by the Bullock-Wellman Postdoctoral Fellowship Award.

References

- Sharma BR, Harish D, Singh VP, Bangar S. Septicemia as a cause of death in burns: an autopsy study. Burns 2006 Aug;32(5):545–9. [PubMed: 16797127]
- 2. Monafo WW. Then and now: 50 years of burn treatment. Burns 1992;18(Suppl 2):S7–10. [PubMed: 1418521]
- Malic CC, Karoo RO, Austin O, Phipps A. Resuscitation burn card--a useful tool for burn injury assessment. Burns 2007;33(2):195–9. [PubMed: 17222978]
- Tegos GP, Demidova TN, Arcila-Lopez D, Lee H, Wharton T, Gali H, et al. Cationic fullerenes are effective and selective antimicrobial photosensitizers. Chem Biol 2005;12(10):1127–35. [PubMed: 16242655]
- 5. Robins EV. Burn shock. Crit Care Nurs Clin North Am 1990;2(2):299-307. [PubMed: 2357324]
- Macintire DK, Bellhorn TL. Bacterial translocation: Clinical implications and prevention. Vet Clin North Am Small Anim Pract 2002;32(5):1165–78. [PubMed: 12380171]
- Ben Saida N, Ferjeni A, Benhadjtaher N, Monastiri K, Boukadida J. Clonality of clinical methicillinresistant *Staphylococcus epidermidis* isolates in a neonatal intensive care unit. Pathol Biol (Paris) 2006;54(6):337–42. [PubMed: 16631317]
- Daoud Z, Moubareck C, Hakime N, Doucet-Populaire F. Extended spectrum beta-lactamase producing *Enterobacteriaceae* in Lebanese ICU patients: Epidemiology and patterns of resistance. J Gen Appl Microbiol 2006;52(3):169–78. [PubMed: 16960333]
- 9. Migliavacca R, Migliavacca A, Nucleo E, Ciaponi A, Spalla M, De Luca C, et al. Molecular epidemiology of ESbetaL producing *P. mirabilis* strains from a long-term care and rehabilitation facility in Italy. New Microbiol 2007;30(3):362–6. [PubMed: 17802927]
- Joseph-Horne T, Hollomon DW. Molecular mechanisms of azole resistance in fungi. FEMS Microbiol Lett 1997;149(2):141–9. [PubMed: 9141655]
- Perlin DS. Antifungal drug resistance: do molecular methods provide a way forward? Curr Opin Infect Dis 2009;22(6):568–73. [PubMed: 19741524]
- Zhang YJ, Yu JJ, Zhang YN, Zhang X, Cheng CJ, Wang JX, et al. Effect of carbendazim resistance on trichothecene production and aggressiveness of Fusarium graminearum. Mol Plant Microbe Interact 2009;22(9):1143–50. [PubMed: 19656048]
- Avenot HF, Sellam A, Karaoglanidis G, Michailides TJ. Characterization of mutations in the ironsulphur subunit of succinate dehydrogenase correlating with Boscalid resistance in Alternaria alternata from California pistachio. Phytopathology 2008;98(6):736–42. [PubMed: 18944299]
- Morera-Lopez Y, Torres-Rodriguez JM, Jimenez-Cabello T. *In vitro* susceptibility of fungal and yeast clinical isolates to itraconazole and voriconazole. Rev Iberoam Micol 2005;22(2):105–9. [PubMed: 16107169]

Dai et al.

- Khan ZU, Ahmad S, Brazda A, Chandy R. Mucor circinelloides as a cause of invasive maxillofacial zygomycosis: an emerging dimorphic pathogen with reduced susceptibility to posaconazole. J Clin Microbiol 2009 Apr;47(4):1244–8. [PubMed: 19171681]
- Church D, Elsayed S, Reid O, Winston B, Lindsay R. Burn wound infections. Clin Microbiol Rev 2006 Apr;19(2):403–34. [PubMed: 16614255]
- Greenhalgh DG, Saffle JR, Holmes JHt, Gamelli RL, Palmieri TL, Horton JW, et al. American Burn Association consensus conference to define sepsis and infection in burns. J Burn Care Res 2007;28 (6):776–90. [PubMed: 17925660]
- Sharma BR. Infection in patients with severe burns: causes and prevention thereof. Infect Dis Clin North Am 2007;21(3):745–59. ix. [PubMed: 17826621]
- Neely AN, Brown RL, Clendening CE, Orloff MM, Gardner J, Greenhalgh DG. Proteolytic activity in human burn wounds. Wound Repair Regen 1997;5(4):302–9. [PubMed: 16984440]
- Samonte VA, Goto M, Ravindranath TM, Fazal N, Holloway VM, Goyal A, et al. Exacerbation of intestinal permeability in rats after a two-hit injury: Burn and *Enterococcus faecalis* infection. Crit Care Med 2004;32(11):2267–73. [PubMed: 15640640]
- 21. Poole MD. Are we facing the end of the antibiotic era? Ear Nose Throat J 1993;72(6):433. [PubMed: 8344186]
- Davies J. Where have All the Antibiotics Gone? Can J Infect Dis Med Microbiol 2006;17(5):287– 90. [PubMed: 18382641]
- 23. Dai T, Huang YY, Hamblin MR. Photodynamic therapy for localized infections-State of the art. Photodiagnosis Photodyn Ther 2009;6(3–4):170–88. [PubMed: 19932449]
- 24. Lipsky BA, Hoey C. Topical antimicrobial therapy for treating chronic wounds. Clin Infect Dis 2009;49(10):1541–9. [PubMed: 19842981]
- 25. Pawelchak, JM.; Freeman, FM. Dressings, granules, and their use in treating wounds. US4538603. 1985.
- Amani H, Dougherty WR, Blome-Eberwein S. Use of Transcyte and dermabrasion to treat burns reduces length of stay in burns of all size and etiology. Burns 2006;32(7):828–32. [PubMed: 16997480]
- 27. Weiss J, Herman O, Rosenberg L, Shafir R. Use of meshed omiderm. Burns Incl Therm Inj 1988;14 (2):161. [PubMed: 3390737]
- Hierlemann, H.; Planck, H. Absorbable medical coating material and method for its manufacture. EP1181941. 2006.
- 29. Murray, DG. Burn wound dressing material. US4659572. 1987.
- Kiene S. Differential therapeutic indications for the useof Xenoderm and SYSpur-derm in burns and other skin defects. Zentralbl Chir 1981;106(18):1201–3. [PubMed: 7304001]
- 31. Woodruff, EA. Bio compatible and blood compatible materials and methods. US4820302. 1989.
- 32. Griffey, ES.; Livesey, SA.; Schiff, CM.; Boerboom, LE. Particulate acellular tissue matrix. US7358284. 2008.
- Heggers, JP.; Hawkins, H.; Edgar, P.; Villarreal, C.; Herndon, DN. Treatment of infections in burns. In: Herndon, DN., editor. Total burn care. 3. London, England: Elsevier Health Sci; 2002. p. 120-69.
- Johnson BA, Anker H, Meleney FL. Bacitracin: A New Antibiotic Produced by a Member of the *B. subtilis* Group. Science 1945;102(2650):376–7. [PubMed: 17770204]
- 35. Palmieri TL, Greenhalgh DG. Topical treatment of pediatric patients with burns: a practical guide. Am J Clin Dermatol 2002;3(8):529–34. [PubMed: 12358554]
- Sinha R, Agarwal RK, Agarwal M. Povidone iodine plus neosporin in superficial burns--a continuing study. Burns 1997;23(7–8):626–8. [PubMed: 9568337]
- Brown MR, Wood SM. Relation between cation and lipid content of cell walls of *Pseudomonas* aeruginosa, Proteus vulgaris and Klebsiella aerogenes and their sensitivity to polymyxin B and other antibacterial agents. J Pharm Pharmacol 1972;24(3):215–8. [PubMed: 4402781]
- Munster AM. Treatment of invasive *Enterobacter cloacae* burn wound sepsis with topical nitrofurazone. J Trauma 1984;24(6):524. [PubMed: 6737529]
- 39. Moyer CA, Brentano L, Gravens DL, Margraf HW, Monafo WW Jr. Treatment of large human burns with 0.5 per cent silver nitrate solution. Arch Surg 1965;90:812–67. [PubMed: 14333527]

Dai et al.

- Klasen HJ. A historical review of the use of silver in the treatment of burns. II. Renewed interest for silver. Burns 2000;26(2):131–8. [PubMed: 10716355]
- 41. Jorgensen B, Price P, Andersen KE, Gottrup F, Bech-Thomsen N, Scanlon E, et al. The silverreleasing foam dressing, Contreet Foam, promotes faster healing of critically colonised venous leg ulcers: A randomised, controlled trial. Int Wound J 2005;2(1):64–73. [PubMed: 16722854]
- 42. Monafo WW, Tandon SN, Ayvazian VH, Tuchschmidt J, Skinner AM, Deitz F. Cerium nitrate: A new topical antiseptic for extensive burns. Surgery 1976;80(4):465–73. [PubMed: 135364]
- Fong J, Wood F. Nanocrystalline silver dressings in wound management: a review. Int J Nanomedicine 2006;1(4):441–9. [PubMed: 17722278]
- 44. Barnea Y, Weiss J, Gur E. A review of the applications of the hydrofiber dressing with silver (Aquacel Ag) in wound care. Ther Clin Risk Manag 2010;6:21–7. [PubMed: 20169033]
- 45. Meaume S, Vallet D, Morere MN, Teot L. Evaluation of a silver-releasing hydroalginate dressing in chronic wounds with signs of local infection. J Wound Care 2005;14(9):411–9. [PubMed: 16240620]
- 46. Sawhney CP. Amniotic membrane as a biological dressing in the management of burns. Burns 1989;15 (5):339–42. [PubMed: 2590408]
- 47. Knutson, RA. Wound-healing compositions containing povidone-iodine. US4401651. 1983.
- Johnson A. A combative healer with no ill-effect. Iodosorb in the treatment of infected wounds. Prof Nurse 1991;7(1):60, 2, 4. [PubMed: 1946486]
- Reimer K, Fleischer W, Brogmann B, Schreier H, Burkhard P, Lanzendorfer A, et al. Povidone-iodine liposomes-an overview. Dermatology 1997;195 (Suppl 2):93–9. [PubMed: 9403264]
- 50. Siegel, P.; Bhatt, B. Process and composition for oral hygiene. US20070292360. 2007.
- Wong TW, Wang YY, Sheu HM, Chuang YC. Bactericidal effects of toluidine blue-mediated photodynamic action on *Vibrio vulnificus*. Antimicrob Agents Chemother 2005;49(3):895–902. [PubMed: 15728881]
- Dai T, Tegos GP, Lu Z, Huang L, Zhiyentayev T, Franklin MJ, et al. Photodynamic therapy for *Acinetobacter baumannii* burn infections in mice. Antimicrob Agents Chemother 2009;53(9):3929– 34. [PubMed: 19564369]
- 53. Love, W.; Brundish, D.; Rhys-Williams, W.; Feng, XD.; Pugin, B. Porphyrin derivatives and their use in photodynamic therapy. US7244841. 2007.
- 54. Hamblin, MR.; Wharton, T.; Gali, H. Photosensitizers for targeted photodynamic therapy. WO2008136906. 2008.
- Lambrechts SA, Demidova TN, Aalders MC, Hasan T, Hamblin MR. Photodynamic therapy for *Staphylococcus aureus* infected burn wounds in mice. Photochem Photobiol Sci 2005;4(7):503–9. [PubMed: 15986057]
- Orenstein A, Klein D, Kopolovic J, Winkler E, Malik Z, Keller N, et al. The use of porphyrins for eradication of *Staphylococcus aureus* in burn wound infections. FEMS Immunol Med Microbiol 1997;19(4):307–14. [PubMed: 9537756]
- Roncucci, G.; Chiti, G.; Dei, D.; Cocchi, A.; Fantetti, L.; Mascheroni, S. Phthalocyanine derivatives, process for their preparation, pharmaceutical compositions containing them and their use. US20090131392. 2009.
- Hamblin, MR.; Foley, JW. Antimicrobial photoinactivation using chalcogen analogs of benzo(A) phenoxazinium dyes. US20080015189. 2008.
- Ribeiro MP, Espiga A, Silva D, Baptista P, Henriques J, Ferreira C, et al. Development of a new chitosan hydrogel for wound dressing. Wound Repair Regen 2009;17(6):817–24. [PubMed: 19903303]
- 60. Sezer AD, Cevher E, Hatipoglu F, Ogurtan Z, Bas AL, Akbuga J. Preparation of fucoidan-chitosan hydrogel and its application as burn healing accelerator on rabbits. Biol Pharm Bull 2008;31(12): 2326–33. [PubMed: 19043221]
- 61. Dai T, Tegos GP, Burkatovskaya M, Castano AP, Hamblin MR. Chitosan acetate bandage as a topical antimicrobial dressing for infected burns. Antimicrob Agents Chemother 2009;53(2):393–400. [PubMed: 19015341]
- 62. Ganz T, Lehrer RI. Defensins. Pharmacol Ther 1995;66(2):191-205. [PubMed: 7667395]

- Chalekson CP, Neumeister MW, Jaynes J. Improvement in burn wound infection and survival with antimicrobial peptide D2A21 (Demegel). Plast Reconstr Surg 2002;109(4):1338–43. [PubMed: 11964988]
- 64. Jacobsen F, Mittler D, Hirsch T, Gerhards A, Lehnhardt M, Voss B, et al. Transient cutaneous adenoviral gene therapy with human host defense peptide hCAP-18/LL-37 is effective for the treatment of burn wound infections. Gene Ther 2005;12(20):1494–502. [PubMed: 15973442]
- 65. Ren HT, Han CM, Zhang R, Xu ZJ, Meng ZQ, Weng HB, et al. The antibacterial effect of cecropin B on *Pseudomonas aeruginosa* infection of wounds in mice. Zhonghua Shao Shang Za Zhi 2006;22 (6):445–7. [PubMed: 17438692]
- 66. Steinstraesser L, Koehler T, Jacobsen F, Daigeler A, Goertz O, Langer S, et al. Host defense peptides in wound healing. Mol Med 2008;14(7–8):528–37. [PubMed: 18385817]
- 67. Epand RM, Epand RF, Savage PB. Ceragenins (cationic steroid compounds), a novel class of antimicrobial agents. Drug News Perspect 2008;21(6):307–11. [PubMed: 18836587]
- McManus AT, Denton CL, Mason AD Jr. Topical chlorhexidine diphosphanilate (WP-973) in burn wound sepsis. Arch Surg 1984;119(2):206–11. [PubMed: 6421265]
- Miller LM, Loder JS, Hansbrough JF, Peterson HD, Monafo WW, Jordan MH. Patient tolerance study of topical chlorhexidine diphosphanilate: a new topical agent for burns. Burns 1990;16(3):217–20. [PubMed: 2383364]
- 70. Selkon, JB. Wound and ulcer treatment with super-oxidized water. US7276255. 2007.
- Hamouda T, Hayes MM, Cao Z, Tonda R, Johnson K, Wright DC, et al. A novel surfactant nanoemulsion with broad-spectrum sporicidal activity against *Bacillus* species. J Infect Dis 1999;180 (6):1939–49. [PubMed: 10558951]
- 72. Modak SM, Fox CL Jr. Sulfadiazine silver-resistant *Pseudomonas* in burns. New topical agents. Arch Surg 1981;116(7):854–7. [PubMed: 6455105]
- Weller R, Price RJ, Ormerod AD, Benjamin N, Leifert C. Antimicrobial effect of acidified nitrite on dermatophyte fungi, Candida and bacterial skin pathogens. J Appl Microbiol 2001;90(4):648–52. [PubMed: 11309079]
- 74. McVay CS, Velasquez M, Fralick JA. Phage therapy of *Pseudomonas aeruginosa* infection in a mouse burn wound model. Antimicrob Agents Chemother 2007;51(6):1934–8. [PubMed: 17387151]
- 75. Ipaktchi K, Mattar A, Niederbichler AD, Hoesel LM, Vollmannshauser S, Hemmila MR, et al. Topical p38 MAPK inhibition reduces bacterial growth in an *in vivo* burn wound model. Surgery 2007;142 (1):86–93. [PubMed: 17630004]
- 76. Peral MC, Martinez MA, Valdez JC. Bacteriotherapy with *Lactobacillus plantarum* in burns. Int Wound J 2009;6(1):73–81. [PubMed: 19291120]
- 77. Subrahmanyam M. Honey dressing versus boiled potato peel in the treatment of burns: A prospective randomized study. Burns 1996;22(6):491–3. [PubMed: 8884013]
- Edwards-Jones V, Buck R, Shawcross SG, Dawson MM, Dunn K. The effect of essential oils on methicillin-resistant *Staphylococcus aureus* using a dressing model. Burns 2004;30(8):772–7. [PubMed: 15555788]
- 79. Zhang HQ, Yip TP, Hui I, Lai V, Wong A. Efficacy of moist exposed burn ointment on burns. J Burn Care Rehabil 2005;26(3):247–51. [PubMed: 15879746]
- Starley IF, Mohammed P, Schneider G, Bickler SW. The treatment of paediatric burns using topical papaya. Burns 1999;25(7):636–9. [PubMed: 10563690]
- 81. Stefanides MM Sr, Copeland CE, Kominos SD, Yee RB. *In vitro* penetration of topical antiseptics through eschar of burn patients. Ann Surg 1976;183(4):358–64. [PubMed: 1267492]
- Moghimi HR, Makhmalzadeh BS, Manafi A. Enhancement effect of terpenes on silver sulphadiazine permeation through third-degree burn eschar. Burns 2009;35(8):1165–70. [PubMed: 19447550]
- Manafi A, Hashemlou A, Momeni P, Moghimi HR. Enhancing drugs absorption through third-degree burn wound eschar. Burns 2008;34(5):698–702. [PubMed: 18226460]
- Lineaweaver W, Howard R, Soucy D, McMorris S, Freeman J, Crain C, et al. Topical antimicrobial toxicity. Arch Surg 1985;120(3):267–70. [PubMed: 3970664]

- Damour O, Hua SZ, Lasne F, Villain M, Rousselle P, Collombel C. Cytotoxicity evaluation of antiseptics and antibiotics on cultured human fibroblasts and keratinocytes. Burns 1992;18(6):479– 85. [PubMed: 1489497]
- Samberg ME, Oldenburg SJ, Monteiro-Riviere NA. Evaluation of silver nanoparticle toxicity in skin in vivo and keratinocytes in vitro. Environ Health Perspect 2010;118(3):407–13. [PubMed: 20064793]
- 87. Supp AP, Neely AN, Supp DM, Warden GD, Boyce ST. Evaluation of cytotoxicity and antimicrobial activity of Acticoat Burn Dressing for management of microbial contamination in cultured skin substitutes grafted to athymic mice. J Burn Care Rehabil 2005;26(3):238–46. [PubMed: 15879745]
- 88. Jacobsen F, Mohammadi-Tabrisi A, Hirsch T, Mittler D, Mygind PH, Sonksen CP, et al. Antimicrobial activity of the recombinant designer host defence peptide P-novispirin G10 in infected full-thickness wounds of porcine skin. J Antimicrob Chemother 2007;59(3):493–8. [PubMed: 17289767]
- 89. Soukos NS, Ximenez-Fyvie LA, Hamblin MR, Socransky SS, Hasan T. Targeted antimicrobial photochemotherapy. Antimicrob Agents Chemother 1998;42(10):2595–601. [PubMed: 9756761]
- 90. Walker HL, Mason AD Jr. A standard animal burn. J Trauma 1968;8(6):1049-51. [PubMed: 5722120]
- Jones WG 2nd, Minei JP, Barber AE, Rayburn JL, Fahey TJ 3rd, Shires GT, et al. Bacterial translocation and intestinal atrophy after thermal injury and burn wound sepsis. Ann Surg 1990;211 (4):399–405. [PubMed: 2108621]
- 92. Steinstraesser L, Klein RD, Aminlari A, Fan MH, Khilanani V, Remick DG, et al. Protegrin-1 enhances bacterial killing in thermally injured skin. Crit Care Med 2001;29(7):1431–7. [PubMed: 11445704]
- 93. Ulkur E, Oncul O, Karagoz H, Celikoz B, Cavuslu S. Comparison of silver-coated dressing (Acticoat), chlorhexidine acetate 0.5% (Bactigrass), and silver sulfadiazine 1% (Silverdin) for topical antibacterial effect in Pseudomonas aeruginosa-contaminated, full-skin thickness burn wounds in rats. J Burn Care Rehabil 2005;26(5):430–3. [PubMed: 16151289]
- 94. Stieritz DD, Holder IA. Experimental studies of the pathogenesis of infections due to *Pseudomonas aeruginosa*: Description of a burned mouse model. J Infect Dis 1975;131(6):688–91. [PubMed: 805812]
- 95. Stevens EJ, Ryan CM, Friedberg JS, Barnhill RL, Yarmush ML, Tompkins RG. A quantitative model of invasive *Pseudomonas* infection in burn injury. J Burn Care Rehabil 1994;15(3):232–5. [PubMed: 8056812]
- 96. Bahar T, Bilezikci B, Maral T, Borman H. A modified partial-thickness burn model in rats. Burns 2007;33:S52–S3.
- Suzuki T, Hirayama T, Aihara K, Hirohata Y. Experimental studies of moderate temperature burns. Burns 1991;17:443–51. [PubMed: 1793491]
- Contag CH, Contag PR, Mullins JI, Spilman SD, Stevenson DK, Benaron DA. Photonic detection of bacterial pathogens in living hosts. Mol Microbiol 1995;18(4):593–603. [PubMed: 8817482]
- Demidova TN, Gad F, Zahra T, Francis KP, Hamblin MR. Monitoring photodynamic therapy of localized infections by bioluminescence imaging of genetically engineered bacteria. JPhotochemPhotobiol B 2005;81:15–25.
- 100. Xiong YQ, Willard J, Kadurugamuwa JL, Yu J, Francis KP, Bayer AS. Real-time *in vivo* bioluminescent imaging for evaluating the efficacy of antibiotics in a rat Staphylococcus aureus endocarditis model. Antimicrob Agents Chemother 2005;49(1):380–7. [PubMed: 15616318]
- 101. Pham C, Greenwood J, Cleland H, Woodruff P, Maddern G. Bioengineered skin substitutes for the management of burns: a systematic review. Burns 2007;33(8):946–57. [PubMed: 17825993]
- 102. Queen D, Evans JH, Gaylor JD, Courtney JM, Reid WH. Burn wound dressings--a review. Burns Incl Therm Inj 1987;13(3):218–28. [PubMed: 3607565]
- 103. Martin FT, O'Sullivan JB, Regan PJ, McCann J, Kelly JL. Hydrocolloid dressing in pediatric burns may decrease operative intervention rates. J Pediatr Surg 2010;45(3):600–5. [PubMed: 20223327]
- Eldad A, Tuchman I. The use of Omiderm as an interface for skin grafting. Burns 1991;17(2):155– 8. [PubMed: 2054075]
- 105. Schwarze H, Kuntscher M, Uhlig C, Hierlemann H, Prantl L, Ottomann C, et al. Suprathel, a new skin substitute, in the management of partial-thickness burn wounds: Results of a clinical study. Ann Plast Surg 2008;60(2):181–5. [PubMed: 18216512]

- 106. Mahnke PF, Rose E, Riedeberger J. A temporary synthetic skin subsitute: SYSpur-derm. Comparative histopathological examination. Zentralbl Chir 1980;105(3):145–53. [PubMed: 7415620]
- 107. Barret JP, Dziewulski P, Ramzy PI, Wolf SE, Desai MH, Herndon DN. Biobrane versus 1% silver sulfadiazine in second-degree pediatric burns. Plast Reconstr Surg 2000;105(1):62–5. [PubMed: 10626971]
- 108. Hassan Z, Shah M. Punctate scarring from use of porous Biobrane. Burns 2006;32(2):258–60. [PubMed: 16448772]
- 109. Naughton, GK.; Naughton, BA. Three-dimensional cell and tissue culture system. US4963489. 1990.
- Hosseini SN, Karimian A, Mousavinasab SN, Rahmanpour H, Yamini M, Zahmatkesh SH. Xenoderm Versus 1% Silver Sulfadiazine in Partial-thickness Burns. Asian J Surg 2009;32(4):234– 9. [PubMed: 19892627]
- 111. Bell, E. Tissue-equivalent and method for preparation thereof. US4485096. 1984.
- 112. Allen-Hoffmann, BL. Method and composition for skin grafts. US6964869. 2005.
- 113. Ceres, RA.; Brown, DJ.; Lesnoy, DC. Bilayered collagen construct. US6500464. 2002.
- 114. Murphy KD, Lee JO, Herndon DN. Current pharmacotherapy for the treatment of severe burns. Expert Opin Pharmacother 2003;4(3):369–84. [PubMed: 12614189]
- 115. Altoparlak U, Erol S, Akcay MN, Celebi F, Kadanali A. The time-related changes of antimicrobial resistance patterns and predominant bacterial profiles of burn wounds and body flora of burned patients. Burns 2004;30(7):660–4. [PubMed: 15475138]
- 116. Elliott TS, Lambert PA. Antibacterial resistance in the intensive care unit: mechanisms and management. Br Med Bull 1999;55(1):259–76. [PubMed: 10695090]
- 117. Mcmanus, A. Anti microbial mafenide-phosphanilate compound, pharmaceutical compositions and method of use therefor. US200402. 1993.
- 118. Falcone PA, Harrison HN, Sowemimo GO, Reading GP. Mafenide acetate concentrations and bacteriostasis in experimental burn wounds treated with a three-layered laminated mafenide-saline dressing. Ann Plast Surg 1980;5(4):266–9. [PubMed: 6985504]
- 119. Heggers JP, Robson MC, Herndon DN, Desai MH. The efficacy of nystatin combined with topical microbial agents in the treatment of burn wound sepsis. J Burn Care Rehabil 1989;10(6):508–11. [PubMed: 2600098]
- 120. Monafo WW, West MA. Current treatment recommendations for topical burn therapy. Drugs 1990p; 40(3):364–73. [PubMed: 2226220]
- 121. Alvarez, OM. Zinc bacitracin containing wound dressing patent. US5061689. 1991.
- Rode H, de Wet PM, Millar AJ, Cywes S. Bactericidal efficacy of mupirocin in multi-antibiotic resistant *Staphylococcus aureus* burn wound infection. J Antimicrob Chemother 1988;21(5):589– 95. [PubMed: 3134320]
- 123. Strock LL, Lee MM, Rutan RL, Desai MH, Robson MC, Herndon DN, et al. Topical bactroban (mupirocin): Efficacy in treating burn wounds infected with methicillin-resistant *Staphylococci.* J Burn Care Rehabil 1990;11(5):454–9. [PubMed: 2123203]
- 124. Drogemoller, ML.; Linley, SL. Composition comprising mupirocin and chlorhexidine. US6214866. 2001.
- 125. Embil JM, McLeod JA, Al-Barrak AM, Thompson GM, Aoki FY, Witwicki EJ, et al. An outbreak of methicillin resistant *Staphylococcus aureus* on a burn unit: Potential role of contaminated hydrotherapy equipment. Burns 2001;27(7):681–8. [PubMed: 11600247]
- 126. Meier PA, Carter CD, Wallace SE, Hollis RJ, Pfaller MA, Herwaldt LA. A prolonged outbreak of methicillin-resistant *Staphylococcus aureus* in the burn unit of a tertiary medical center. Infect Control Hosp Epidemiol 1996;17(12):798–802. [PubMed: 8985766]
- 127. Hansbrough JF, Zapata-Sirvent RL, Cooper ML. Effects of topical antimicrobial agents on the human neutrophil respiratory burst. Arch Surg 1991;126(5):603–8. [PubMed: 1850590]
- 128. Darouiche, RO.; Raad, I. Antimicrobial impregnated catheters and other medical implants. US5624704. 1999.

NIH-PA Author Manuscript

Dai et al.

- 129. Guay DR. An update on the role of nitrofurans in the management of urinary tract infections. Drugs 2001;61(3):353–64. [PubMed: 11293646]
- Mousa HA. Fungal infection of burn wounds in patients with open and occlusive treatment methods. East Mediterr Health J 1999;5(2):333–6. [PubMed: 10793810]
- 131. Lopez-Berestein, G.; Mehta, R.; Hopfer, RL.; Juliano, RL. Liposome-incorporated nystatin. US4812312. 1989.
- 132. Kwon, GS.; Yoo, BK. Nystatin formulation having reduced toxicity. US6413537. 2002.
- 133. Yapijakis C. Hippocrates of Kos, the father of clinical medicine, and Asclepiades of Bithynia, the father of molecular medicine. Review In vivo 2009;23(4):507–14.
- Alexander JW. History of the medical use of silver. Surg Infect (Larchmt) 2009;10(3):289–92. [PubMed: 19566416]
- Borsuk DE, Gallant M, Richard D, Williams HB. Silver-coated nylon dressings for pediatric burn victims. Can J Plast Surg 2007;15(1):29–31. [PubMed: 19554127]
- 136. Atiyeh BS, Costagliola M, Hayek SN, Dibo SA. Effect of silver on burn wound infection control and healing: Review of the literature. Burns 2007;33(2):139–48. [PubMed: 17137719]
- 137. Warriner R, Burrell R. Infection and the chronic wound: A focus on silver. Adv Skin Wound Care 2005;18 (Suppl 1):2–12. [PubMed: 16220035]
- Poon VK, Burd A. *In vitro* cytotoxity of silver: Implication for clinical wound care. Burns 2004;30 (2):140–7. [PubMed: 15019121]
- 139. Kershaw, D.; DeBoorder, B.; Law, SJ. Antibacterial wound dressing. US20100015208. 2010.
- 140. Mason, ADJ.; Johnson, AAJ.; Ritchey, CR. Protective gel composition for treating white phosphorus burn wounds. US4391799. 1983.
- 141. Schmolka, IR. Silver ion gel compositions. US4376764. 1983.
- 142. Neuwirth, RS. Intrauterine chemical cauterizing method and composition. US2008895424. 2008.
- 143. Zanoni D, Isenberg SJ, Apt L. A comparison of silver nitrate with erythromycin for prophylaxis against ophthalmia neonatorum. Clin Pediatr (Phila) 1992;31(5):295–8. [PubMed: 1582096]
- 144. Fox CL Jr, Modak SM. Mechanism of silver sulfadiazine action on burn wound infections. Antimicrob Agents Chemother 1974;5(6):582–8. [PubMed: 15825409]
- 145. Cho Lee AR, Leem H, Lee J, Park KC. Reversal of silver sulfadiazine-impaired wound healing by epidermal growth factor. Biomaterials 2005;26(22):4670–6. [PubMed: 15722137]
- 146. Rosenkranz HS, Coward JE, Wlodkowski TJ, Carr HS. Properties of silver sulfadiazine-resistant Enterobacter cloacae. Antimicrob Agents Chemother 1974;5(2):199–201. [PubMed: 4840432]
- 147. Fox, CLJ.; Modak, SM. Antibacterial composition comprising silver sulfadiazine and sodium piperacillin. US4535078. 1985.
- 148. Modak, SM.; Fox, CLJ.; Fox, P. Wound dressing comprising silver sulfadiazine incorporated in animal tissue. CA1219209. 1987.
- 149. Fox, CLJ. Silver sulfadiazine used in the treatment of burns. US3761590. 1973.
- 150. Chen, JR. Topical spray for burn treatment and anti-infection. US6551577. 2003.
- 151. Garner JP, Heppell PS. Cerium nitrate in the management of burns. Burns 2005;31(5):539–47. [PubMed: 15955636]
- 152. Monafo, WW. Water-soluble cerium (cerous) salts in burn therapy. US4088754. 1978.
- 153. Gravante G, Caruso R, Sorge R, Nicoli F, Gentile P, Cervelli V. Nanocrystalline silver: A systematic review of randomized trials conducted on burned patients and an evidence-based assessment of potential advantages over older silver formulations. Ann Plast Surg 2009;63(2):201–5. [PubMed: 19571738]
- 154. Ma, RH.; Yu, YH. Nano-silver wound dressing. US7462753. 2008.
- 155. Holladay, RJ.; Christensen, H.; Moeller, WD. Treatment of humans with colloidal silver composition. US7135195. 2006.
- 156. Gillis, SH. Therapeutic treatments using the direct application of antimicrobial metal compositions. US20060083792. 2006.
- 157. Gillis, SH. Therapeutic treatments using the direct application of antimicrobial metal compositions. US6989156. 2006.

Dai et al.

- 158. Bell A, Hart J. Evaluation of two absorbent silver dressings in a porcine partial-thickness excisional wound model. J Wound Care 2007;16(10):445–8. 50–3. [PubMed: 18065021]
- 159. Scherr, GH. Silver alginate foam compositions. US20030021832. 2003.
- 160. Kim, YS.; Kim, JY. Methods for producing silver-bonded antimicrobial moist wound dressings and moist wound dressings produced by the methods. EP1993620. 2008.
- 161. Ma, RH.; Yu, YH. Nano-silver wound dressing. US7462753. 2008.
- 162. Mcknight, JJ.; Guldalian, JJ. Laminated collagen film dressing. US3800792. 1974.
- 163. Lerat, YJ.; Poncelet, OJ. Dressing and antiseptic agent containing silver. US7323614. 2008.
- 164. Haberal M, Oner Z, Bayraktar U, Bilgin N. The use of silver nitrate-incorporated amniotic membrane as a temporary dressing. Burns Incl Therm Inj 1987;13(2):159–63. [PubMed: 3580940]
- 165. Singh R, Kumar D, Kumar P, Chacharkar MP. Development and evaluation of silver-impregnated amniotic membrane as an antimicrobial burn dressing. J Burn Care Res 2008;29(1):64–72. [PubMed: 18182899]
- 166. Fox, CLJ.; Modak, SM.; Fox, P. Wound dressing comprising silver sulfadiazine incorporated in animal tissue. US4446124. 1984.
- 167. Capelli, C. Stabilized silver-ion amine complex compositions and methods. US6923990. 2005.
- 168. Leaper DJ, Durani P. Topical antimicrobial therapy of chronic wounds healing by secondary intention using iodine products. Int Wound J 2008;5(2):361–8. [PubMed: 18494641]
- 169. Durani P, Leaper D. Povidone-iodine: use in hand disinfection, skin preparation and antiseptic irrigation. Int Wound J 2008;5(3):376–87. [PubMed: 18593388]
- 170. Zamora JL. Iodine toxicity. Ann Thorac Surg 1986;41(4):462–3. [PubMed: 3963929]
- 171. Capriotti, JA.; Liang, B.; Samson, CM.; Stein, J.; Weise, M. Device for in situ generation of povidone-iodine compositions. WO2009097123. 2009.
- 172. Beukelman CJ, van den Berg AJ, Hoekstra MJ, Uhl R, Reimer K, Mueller S. Anti-inflammatory properties of a liposomal hydrogel with povidone-iodine (Repithel) for wound healing *in vitro*. Burns 2008;34(6):845–55. [PubMed: 18378399]
- 173. Muhlau, S.; Fleischer, W. Dry liposomal PVP-iodine compositions. US2004234589. 2004.
- 174. Ormiston MC, Seymour MT, Venn GE, Cohen RI, Fox JA. Controlled trial of Iodosorb in chronic venous ulcers. Br Med J (Clin Res Ed) 1985;291(6491):308–10.
- 175. Kota, S.; Adibhatla, KS.; Venkaiah, CN. Improved process for the preparation of cadexomer iodine. WO2008117300. 2008.
- 176. Larsen, N. Anti-infective iodine based compositions for otic and nasal use. US2006280809. 2006.
- 177. Scholtz, MT. Liquid antiseptic compositions containing iodine and A sugar and/or sugar alcohol. WO2009088826. 2009.
- 178. Foret, C.; Hemling, TC. Iodine antimicrobial compositions containing nonionic surfactants and halogen anions. US5916581. 1999.
- 179. Hamblin, MR.; Mroz, P. Advances in photodynamic therapy: basic, translational and clinical. Norwood, MA: Artech House; 2008.
- 180. Moan J, Peng Q. An outline of the hundred-year history of PDT. Anticancer Res 2003;23(5A):3591– 600. [PubMed: 14666654]
- 181. Castano AP, Demidova TN, Hamblin MR. Mechanisms in photodynamic therapy: part onephotosensitizers, photochemistry and cellular localization. Photodiagn Photodyn Ther 2004;1(4): 279–93.
- Hamblin MR, Hasan T. Photodynamic therapy: A new antimicrobial approach to infectious disease? 2004;3(5):436–50.
- 183. Phoenix DA, Harris F. Light activated compounds as antimicrobial agents patently obvious? Recent Pat Antiinfect Drug Discov 2006;1(2):181–99. [PubMed: 18221144]
- 184. Nifantiev, NE.; Gitter, B.; Yashunsky, DV. Anti-microbial photodynamic therapy. US2007016951. 2007.
- 185. Hasan, T.; Hamblin, MR.; Soukos, NS. Photosensitizer conjugates for pathogen targeting. US7268155. 2007.

- 186. Hamblin MR, O'Donnell DA, Murthy N, Rajagopalan K, Michaud N, Sherwood ME, et al. Polycationic photosensitizer conjugates: Effects of chain length and Gram classification on the photodynamic inactivation of bacteria. J Antimicrob Chemother 2002;49(6):941–51. [PubMed: 12039886]
- 187. Tegos GP, Anbe M, Yang C, Demidova TN, Satti M, Mroz P, et al. Protease-stable polycationic photosensitizer conjugates between polyethyleneimine and chlorin(e6) for broad-spectrum antimicrobial photoinactivation. Antimicrob Agents Chemother 2006;50(4):1402–10. [PubMed: 16569858]
- 188. Raghavendra M, Koregol A, Bhola S. Photodynamic therapy: a targeted therapy in periodontics. Aust Dent J 2009;54 (Suppl 1):S102–9. [PubMed: 19737261]
- Hamblin, MR.; Tegos, GP. Inactivation of Microorganisms with multidrug resistance inhibitors and phenothiaziniums. US20080166304. 2008.
- 190. Biel, MA. Photodynamic therapy utilizing a solution of photosensitizing compound and surfactant. US7229447. 2007.
- Albrecht, V.; Gitter, B. Antimicrobial photodynamic therapy compound and method of use. US20050049228. 2005.
- 192. Albrecht, V.; Fahr, A.; Scheglmann, D.; Gräfe, S.; Neuberger, W. Liposomal formulations of hydrophobic photosensitizer for photodynamic therapy. US7354599. 2008.
- 193. Van Der Haas, HN.; Lugtenburg, J.; Schuitmaker, J.; De Jong, RL. Method of preparing a porphyrin derivative, a porphyrin derivative, use of said porphyrin derivative and a pharmaceutical composition containing said porphyrin derivative. US20090215851. 2009.
- 194. Smijs, GM.; Schuitmaker, JJ. Use of a porphyrin compound for the treatment of skin fungi. US20060258635. 2006.
- 195. Love, W.; Brundish, D.; Rhys-Williams, W.; Feng, XD.; Pugin, B. Porphyrin derivatives and their use in photodynamic therapy. US7244841. 2007.
- 196. Rabea EI, Badawy ME, Stevens CV, Smagghe G, Steurbaut W. Chitosan as antimicrobial agent: Applications and mode of action. Biomacromolecules 2003;4(6):1457–65. [PubMed: 14606868]
- 197. Sparkes, BG.; Murray, DG. Chitosan based wound dressing materials. US4572906. 1986.
- 198. Yura, H.; Saito, Y.; Ishihara, M.; Ono, K.; Saeki, S. Functional chitosan derivative. US7125968. 2006.
- 199. Baker, S.; Wiesmann, WP. Methods and compositions for the prevention of and treatment of infections utilizing chitosan-derivative compounds. US20100056474. 2010.
- 200. Scherr, GH. Chitosan wound dressing. US20070237811. 2007.
- 201. Muzzarelli, R. Methyl pyrrolidinone chitosan, production process and uses thereof. US5378472. 1995.
- 202. Alsarra IA. Chitosan topical gel formulation in the management of burn wounds. Int J Biol Macromol 2009;45(1):16–21. [PubMed: 19447254]
- 203. Aoyagi S, Onishi H, Machida Y. Novel chitosan wound dressing loaded with minocycline for the treatment of severe burn wounds. Int J Pharm 2007;330(1–2):138–45. [PubMed: 17049772]
- 204. Boucard N, Viton C, Agay D, Mari E, Roger T, Chancerelle Y, et al. The use of physical hydrogels of chitosan for skin regeneration following third-degree burns. Biomaterials 2007;28(24):3478–88. [PubMed: 17482258]
- 205. Cardenas G, Anaya P, von Plessing C, Rojas C, Sepulveda J. Chitosan composite films. Biomedical applications. J Mater Sci Mater Med 2008;19(6):2397–405. [PubMed: 18165888]
- 206. Sezer AD, Hatipoglu F, Cevher E, Ogurtan Z, Bas AL, Akbuga J. Chitosan film containing fucoidan as a wound dressing for dermal burn healing: preparation and *in vitro/in vivo* evaluation. AAPS Pharm Sci Tech 2007;8(2) Article 39.
- 207. Damour O, Gueugniaud PY, Berthin-Maghit M, Rousselle P, Berthod F, Sahuc F, et al. A dermal substrate made of collagen-GAG-chitosan for deep burn coverage: first clinical uses. Clin Mater 1994;15(4):273–6. [PubMed: 10147171]
- 208. Peh K, Khan T, Ch'ng H. Mechanical, bioadhesive strength and biological evaluations of chitosan films for wound dressing. J Pharm Pharm Sci 2000;3(3):303–11. [PubMed: 11177648]

- Mor A. Multifunctional host defense peptides: Antiparasitic activities. FEBS J 2009;276(22):6474– 82. [PubMed: 19817857]
- Schittek B, Paulmann M, Senyurek I, Steffen H. The role of antimicrobial peptides in human skin and in skin infectious diseases. Infect Disord Drug Targets 2008;8(3):135–43. [PubMed: 18782030]
- 211. Selsted ME, Szklarek D, Lehrer RI. Purification and antibacterial activity of antimicrobial peptides of rabbit granulocytes. Infect Immun 1984;45(1):150–4. [PubMed: 6735465]
- 212. Lehrer RI, Szklarek D, Ganz T, Selsted ME. Correlation of binding of rabbit granulocyte peptides to *Candida albicans* with candidacidal activity. Infect Immun 1985;49(1):207–11. [PubMed: 3891625]
- 213. Lehrer RI, Daher K, Ganz T, Selsted ME. Direct inactivation of viruses by MCP-1 and MCP-2, natural peptide antibiotics from rabbit leukocytes. J Virol 1985;54(2):467–72. [PubMed: 2985808]
- 214. Pazgier M, Prahl A, Hoover DM, Lubkowski J. Studies of the biological properties of human betadefensin 1. J Biol Chem 2007;282(3):1819–29. [PubMed: 17071614]
- Fulton C, Anderson GM, Zasloff M, Bull R, Quinn AG. Expression of natural peptide antibiotics in human skin. Lancet 1997;350(9093):1750–1. [PubMed: 9413472]
- 216. Bensch KW, Raida M, Magert HJ, Schulz-Knappe P, Forssmann WG. hBD-1: A novel beta-defensin from human plasma. FEBS Lett 1995;368(2):331–5. [PubMed: 7628632]
- 217. Zhao C, Wang I, Lehrer RI. Widespread expression of beta-defensin hBD-1 in human secretory glands and epithelial cells. FEBS Lett 1996;396(2–3):319–22. [PubMed: 8915011]
- 218. Bhat S, Milner S. Antimicrobial peptides in burns and wounds. Curr Protein Pept Sci 2007;8(5): 506–20. [PubMed: 17979765]
- 219. Strom, RM.; Brondsema, PJ. Periodic antimicrobial peptides. US7091185. 2006.
- 220. Strom, RM.; Brondsema, PJ. Periodic antimicrobial peptides. US20050187151. 2005.
- 221. Schwab U, Gilligan P, Jaynes J, Henke D. *In vitro* activities of designed antimicrobial peptides against multidrug-resistant cystic fibrosis pathogens. Antimicrob Agents Chemother 1999;43(6): 1435–40. [PubMed: 10348766]
- 222. Chalekson CP, Neumeister MW, Jaynes J. Treatment of infected wounds with the antimicrobial peptide D2A21. J Trauma 2003 Apr;54(4):770–4. [PubMed: 12707542]
- 223. Hadnagy A, Beaulieu R, Balicki D. Histone tail modifications and noncanonical functions of histones: perspectives in cancer epigenetics. Mol Cancer Ther 2008;7(4):740–8. [PubMed: 18413789]
- 224. Rose FR, Bailey K, Keyte JW, Chan WC, Greenwood D, Mahida YR. Potential role of epithelial cell-derived histone H1 proteins in innate antimicrobial defense in the human gastrointestinal tract. Infect Immun 1998;66(7):3255–63. [PubMed: 9632593]
- 225. Pohlmeyer, K.; Behnke, B.; Wick, RZ.; Mayer, G. Recombinant production of human histone 1 subtypes and their use for therapeutic purposes. US20030078204. 2003.
- 226. Lai, JS.; Lee, JH.; Callaway, JE. Cecropin polypetides with activity against Gram-positive and Gramnegative bacteria. US5166321. 1992.
- 227. Ghiselli R, Giacometti A, Cirioni O, Mocchegiani F, Orlando F, D'Amato G, et al. Cecropin B enhances betalactams activities in experimental rat models of gram-negative septic shock. Ann Surg 2004;239(2):251–6. [PubMed: 14745334]
- 228. Horwitz, A.; Lambert, LHJ.; Little, RG. Anti-gram-positive bacterial methods and materials. US5578572. 1996.
- 229. Savage, PB.; Li, C. Steroid derived antibiotics. US6486148. 2002.
- 230. Savage, PB.; Li, C. Steroid derived antibiotics. US6767904. 2004.
- 231. Savage, PB.; Li, C. Steroid derived antibiotics. US7598234. 2009.
- 232. Dang, X.; Zhang, W.; Wang, Y.; Zheng, X.; Wang, JX.; Wang, L. Musca domestic pupae natural antimicrobial peptide products and preparation method and use thereof. CN101392020. 2009.
- Beuerman, RW.; Liu, S.; Li, J.; Zhou, L.; Verma, CS.; Tan, D. Multimeric forms of antimicrobial peptides. WO2009131548. 2009.
- 234. Liepke, C.; Baxmann, S.; Adermann, K. Novel antimicrobial bolisin peptides. US2009149391. 2009.
- Oppenheim, FG.; Xu, T.; Roberts, FD.; Spacciapoli, P.; Friden, PM. Anti-fungal and anti-bacterial histatin-based peptides. US5912230. 1999.

Dai et al.

- 236. Hara, S. Peptide, antibacterial agent, peptide gene, recombinant DNA and method for preparing the peptide. US5646014. 1997.
- 237. Djurup, R.; Flodgaard, HJ.; Norris, K. Bactericidal, anti-apoptotic, pro-inflammatory and antiinflammatory peptides of heparin-binding protein (Hbp) Or human neutrophil elastase. WO2004016653. 2004.
- 238. Schmidtchen, A.; Malmsten, M. Improved antimicrobial peptides. KR0027213. 2009.
- Krieger, TJ.; Taylor, R.; Erfle, D.; Fraser, JR.; West, MH.; Mcnichol, PJ. Compositions and methods for treating infections using cationic peptides alone or in combination with antibiotics. US6503881. 2009.
- 240. Drake, D.; Marek, CL. Chlorhexidine compositions. US20050191247. 2005.
- 241. Warner, PLJ.; Kornegay, GB.; Redl, G. Chlorhexidine salts and compositions of same. US4666896. 1987.
- 242. Friedman, J. Compositions for the sustained-release of chlorhexidine. US5002769. 1991.
- 243. Baker, JRJ.; Hamouda, T.; Shih, A.; Myc, A. Antimicrobial nanoemulsion compositions and methods. US7655252. 2010.
- 244. Schlag S, Nerz C, Birkenstock TA, Altenberend F, Gotz F. Inhibition of *Staphylococcal* biofilm formation by nitrite. J Bacteriol 2007;189(21):7911–9. [PubMed: 17720780]
- 245. Summers WC. Bacteriophage therapy. Annu Rev Microbiol 2001;55:437-51. [PubMed: 11544363]
- 246. Malik R, Chhibber S. Protection with bacteriophage KO1 against fatal *Klebsiella pneumoniae*induced burn wound infection in mice. J Microbiol Immunol Infect 2009;42(2):134–40. [PubMed: 19597645]
- 247. Soothill, J.; Hawkins, C.; Harper, D. Bacteriophage-containing therapeutic agents. US20070190033. 2007.
- 248. Walsh, SM.; Pittaway, MC. Compositions and methods for treating bacteria. US7517852. 2009.
- Wahdan HA. Causes of the antimicrobial activity of honey. Infection 1998;26(1):26–31. [PubMed: 9505176]
- Subrahmanyam M. Topical application of honey in treatment of burns. Br J Surg 1991;78(4):497– 8. [PubMed: 2032114]
- 251. Mousa, MA. Honey preparations. US5980875. 1999.
- 252. Mousa, MA. Honey preparations. US6171604. 2001.
- 253. Van Den Berg, AJ.; Hoekstra, M. Wound dressings incorporating honey. US20090304780. 2009.
- 254. Caskey, PR. Honey in wound dressings. US20040127826. 2004.
- 255. Molan, P. Honey based wound dressing. US20040054313. 2004.
- 256. Holzner, G. Antimicrobial perfume compositions. US6479456. 2002.
- 257. Ang ES, Lee ST, Gan CS, See P, Chan YH, Ng LH, et al. The role of alternative therapy in the management of partial thickness burns of the face-experience with the use of moist exposed burn ointment (MEBO) compared with silver sulphadiazine. Ann Acad Med Singapore 2000;29(1):7– 10. [PubMed: 10748957]
- 258. Amin AH, Subbaiah TV, Abbasi KM. Berberine sulfate: Antimicrobial activity, bioassay, and mode of action. Can Microbiol 1969;15(9):1067–76.

NIH-PA Author Manuscript

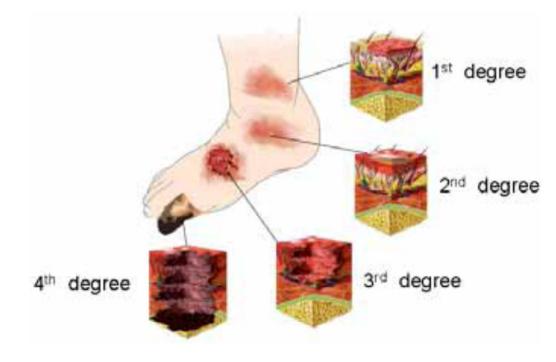


Fig. 1. Degrees of burn Relative depths of 1st, 2nd, 3rd, and 4th degree burns illustrated on a foot.

Dai et al.

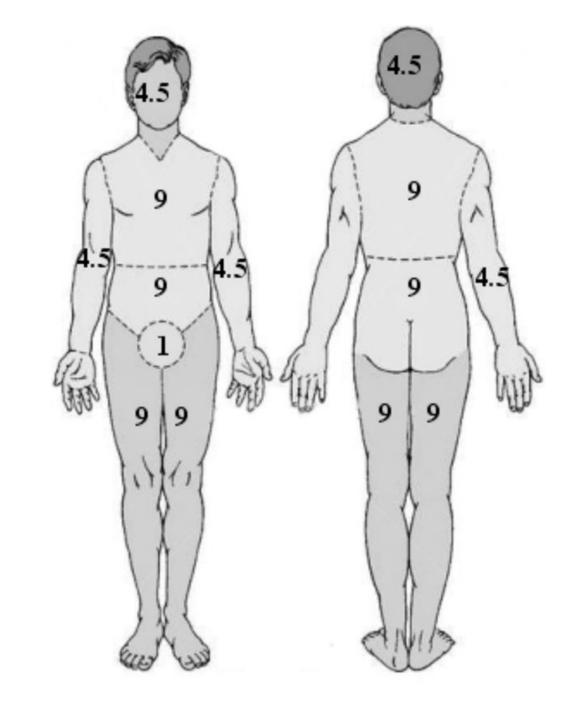


Fig. 2. Lund & Browder charts These are used to calculate the % of body surface area affected by burns and determine the prognosis and susceptibility to infection.

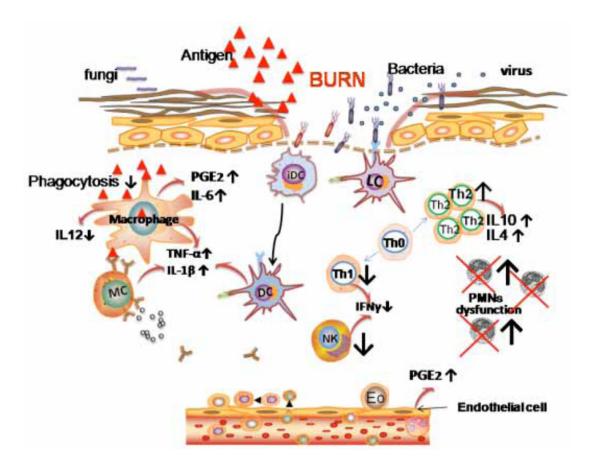


Fig. 3. Inflammatory signaling after burn that causes immunosuppression

Interleukin-1 (IL-1) and tumor necrosis factor alpha (TNF α) are produced by a wide variety of cells. The production of prostaglandin E2 (PGE2) and IL6 are upregulated by endothelial cells and macrophages, while the latter secrete decreased amounts of IL-12. T helper cells begin to preferentially differentiate into Th-2 cells, which produce the anti-inflammatory cytokines IL-4 and IL-10. Neutrophil dysfunction occurs despite higher numbers after significant thermal injuries, while macrophage phagocytosis is lower.

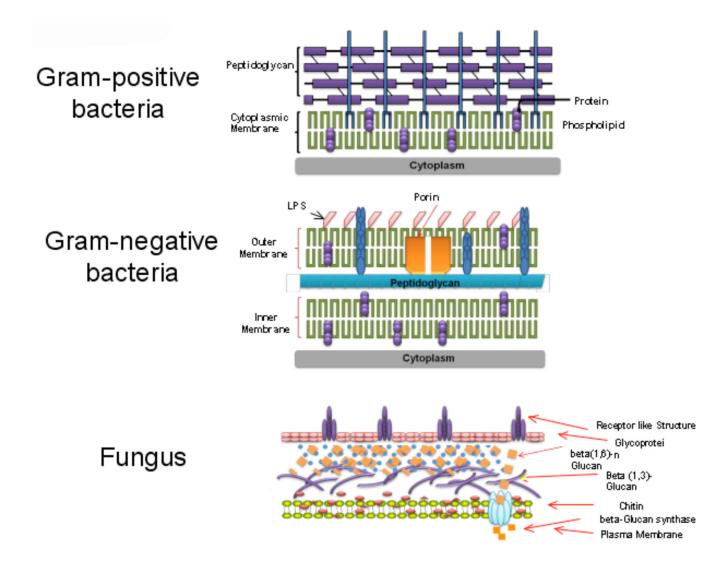
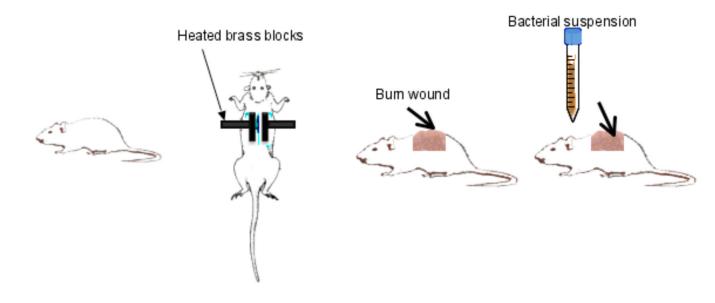
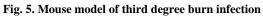


Fig. 4. Cell wall structure

Gram-positive bacteria, Gram-negative bacteria and fungi have different cell wall structures that govern their susceptibility to different antimicrobial agents.





Two pre-heated brass blocks are pressed to the opposing sides of an elevated skin fold on the backs of shaved mice, followed by topical application of a suspension of bacteria in saline.

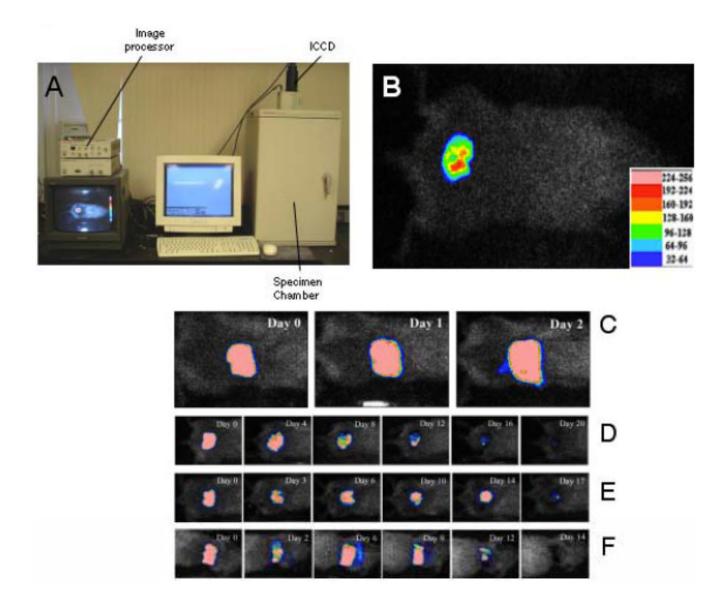


Fig. 6. Use of bioluminescent imaging of genetically engineered bacteria and fungi in burn infections (**A**) Picture of the low-light camera suitable for *in vivo* bioluminescence imaging of small animals. (**B**) Example of 3^{rd} degree burn on mouse back infected with bioluminescent *P. aeruginosa* showing color look-up-table for photon intensity. Successive bioluminescence images of representative mouse burns infected with bioluminescent strains of (**C**) *P. aeruginosa*, (**D**) *Acinetobacter baumannii*, (**E**) *Staphylococcus aureus*, and (**F**) *Candida albicans*, respectively.

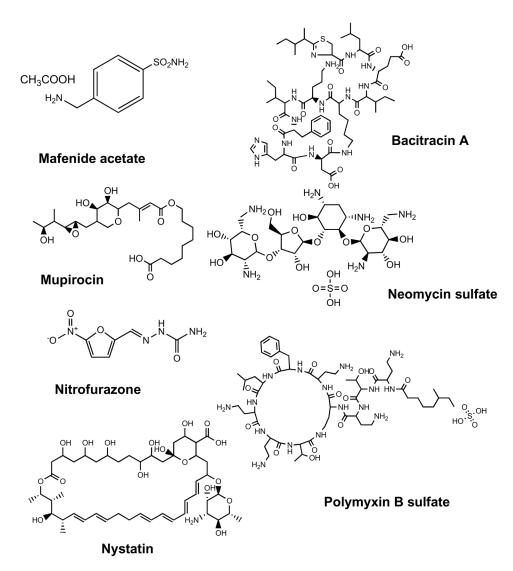


Fig. 7. Chemical structures of antibiotics used in topical antimicrobial therapy of burns Mafenide acetate, bacitracin, mupirocin, neomycin, polymyxin B, nitrofurazone, nystatin.

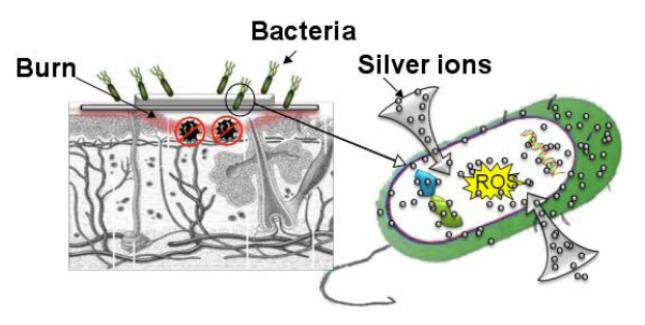


Fig. 8. Use of silver as a topical antimicrobial agent in burns

Silver has multiple mechanisms of action as an antimicrobial agent including permeabilization of bacterial cell walls and generation of intracellular reactive oxygen species.

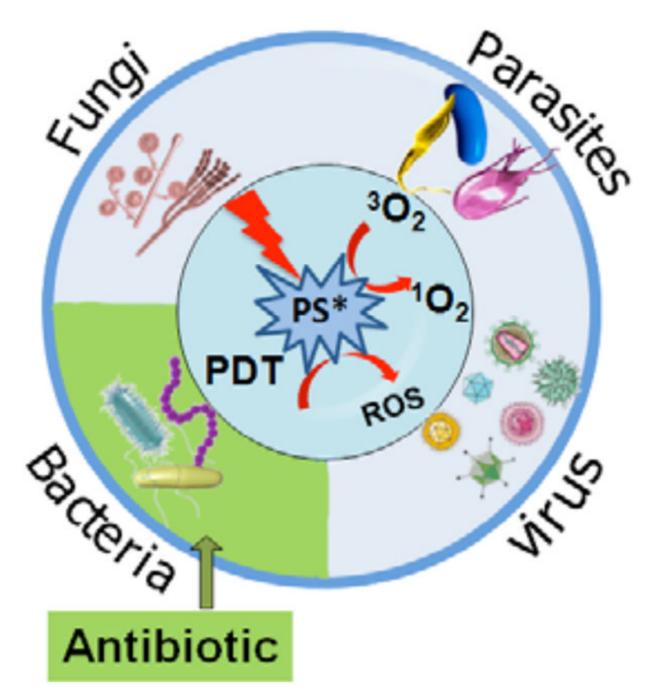
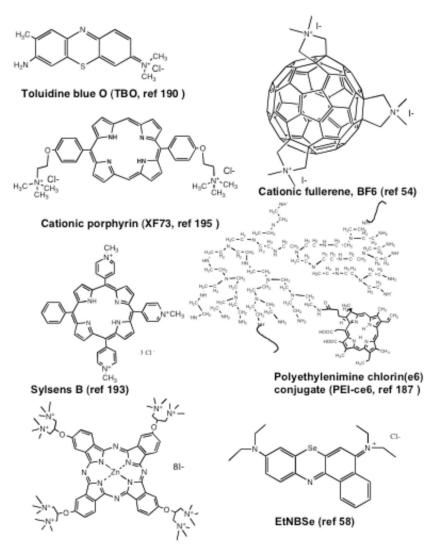


Fig. 9. Concept of antimicrobial photodynamic therapy

The photosensitizer is excited to its triplet state by light of the correct wavelength and then transfers its energy to molecular oxygen forming reactive oxygen species that are able to kill all classes of pathogenic micro-organisms.



Cationic phthalocyanine (RLP068, ref 57)

Fig. 10. Chemical structures of photosensitizers used in antimicrobial applications

These molecules possess either constitutive cationic charges or basic amino groups. Toluidine blue O (TBO), cationic fullerene (BF6), cationic porphyrin (XF73), meso-mono-phenyl-tri(N-methyl-4-pyridyl)-porphyrin (Sylsens B), polyethylenimine chlorin(e6) conjugate (PEI-ce6), cationic phthalocyanine (RLP068), selenium benzo(A)phenoxazinium chloride (EtNBSe).

PS+light PDT	o "	3d'	Dye 15	12.Jem ²	36 J/cm	84.J/cm ⁷	180 J/cm ²	240.3 cm²
PS dark			Dye 15	Dye 17	Dye 21'	Dye 29	Dye 45"	Dye 55
Lightalone	•	0		12.Jem ⁴	36.kem²	84 fiem ²	180 J/cm²	240 J/cm ¹

Fig. 11. Use of PDT to treat mouse burn infection caused by bioluminescent *A. baumannii* Bacterial luminescence of a representative mouse burn infected with *A. baumannii* and treated with PDT at 30 minutes after infection (top row), a mouse burn treated with PS only (dark control, middle row), and a mouse burn treated with light only (light control, bottom row). After 240 J/cm² red light had been delivered (40 minutes irradiation time at an irradiance of 100 mW/cm²), PDT induced an approximately 3-log-unit reduction in bacterial luminescence from the mouse burn, while during the same period of time, only modest reduction of bacterial luminescence was observed in the dark control and the light control.

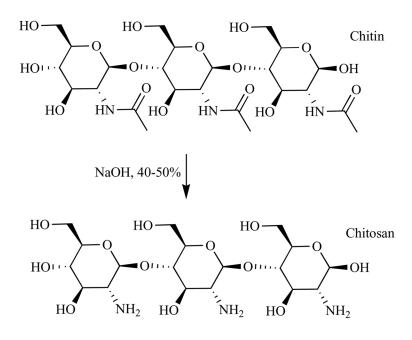


Fig. 12. Chemical structure of chitin and chitosan

Treatment of chitin (poly-N-acetyl glucosamine) with hot sodium hydroxide partially hydrolyzes the amide bonds to form chitosan (poly-glucosamine).

Day 0	Day I	Day 2	Day 3
Control	Control	Control	Control
Day 0	Day 1	Day 2	Day 3
Silver	Silver	Silver	Silver
Day 0	Day 1	Day 2	Day 3
Chitosan	Chitosan	Chitosan	Chitosan

Fig. 13. Use of a chitosan acetate bandage to treat mouse burn infection

The successive bioluminescence images from day 0 to day 3 after *P. aeruginosa* infection of an untreated burn (top row), a silver dressing-treated burn (middle row), and a chitosan acetate-treated burn (bottom row).

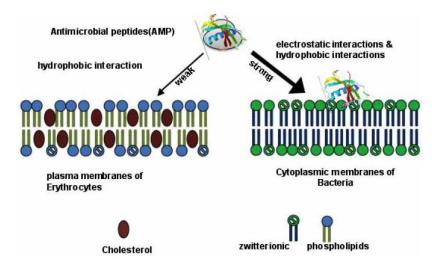


Fig. 14. Antimicrobial action of AMPs

Schematic illustration of the selectivity exhibited by antimicrobial peptides that allow them to lyse microbial cell membranes but not those of host cells.

Table 1

Microbiology of Pathogens Responsible for Burn Infections and the Occurrence of Drug Resistance

Group	Species	Drug Resistance		
Gram-positive bacteria	Staphylococcus aureus			
	Methicillin-resistant S. aureus	By definition		
	Coagulase-negative staphylococci	MRSE increasing [7]		
	Enterococcus sp.			
	Vancomycin-resistant enterococci	By definition		
Gram-negative bacteria	Pseudomonas aeruginosa	High innate resistance		
	Escherichia coli	ESBL increasing		
	Klebsiella pneumoniae	ESBL increasing		
	Serratia marcescens	increasing		
	Enterobacter sp.	ESBL increasing [8]		
	Proteus sp.	ESBL increasing [9]		
	Acinetobacter sp.	Very common		
	Bacteroides sp.	uncommon		
Fungi	Candida sp.	Azole resistance and efflux pumps increasing [1		
	Aspergillus sp.	Increasing [11]		
	Fusarium sp.	Common in agriculture [12]		
	Alternaria sp.	Common in agriculture]13]		
	Rhizopus sp.	Azole resistance [14]		
	Mucor sp.	Azole resistance [15]		
Viruses	Herpes simplex virus			
	Cytomegalovirus			
	Varicella-zoster virus			

Table 2

Summary of Topical Antimicrobial Therapy Agents used for Burns Covered in this Review

Agent Class	Specific Agent	Application	Ref
Skin Substitutes	Duoderm	Clinical 2 nd degree burns	[25]
	Trans-Cyte	Clinical 2 nd degree burns	[26]
	Omniderm	Clinical 2 nd degree burns	[27]
	Suprathel	Clinical 2 nd degree burns	[28]
	Epigard	Clinical 2 nd degree burns	[29]
	SYSpur-derm	Clinical 2 nd degree burns	[30]
	Biobrane	Clinical 2 nd degree burns	[31]
	Xenoderm	Clinical 2 nd degree burns	[32]
Topical antibiotics	Mafenide acetate	Clinical 2nd/3rd degree burns	[33]
	Bacitracin	Clinical 2nd/3rd degree burns	[34]
	Mupirocin	Clinical 2 nd /3 rd degree burns	[35]
	Neosporin	Clinical 2 nd /3 rd degree burns	[36]
	Polymyxin B	Clinical 2 nd /3 rd degree burns	[37]
	Nitrofurazone	Clinical 2nd/3rd degree burns	[38]
	Nystatin	Clinical 2nd/3rd degree burns, fungal infections	[35]
Silver	Silver nitrate	itrate Clinical 2 nd /3 rd degree burns	
	Silver sulfadiazine	Clinical 2 nd /3 rd degree burns	[40]
	Silver foams (Contreet, Allevyn)	Clinical 2 nd /3 rd degree burns	[41]
	Flammacerium	Clinical 2nd/3rd degree burns	[42]
	Acticoat 7	Clinical 2 nd /3 rd degree burns	[43]
	Aquacel-Ag	Clinical 2 nd /3 rd degree burns	[44]
	Silvercel	Clinical 2 nd /3 rd degree burns	[45]
	Silver amniotic membrane (Clinical 2 nd /3 rd degree burns	[46]
Iodine	Povidone-iodine	Clinical 2nd/3rd degree burns	[47]
	Cadexomer iodine (Iodosorb)	Clinical chronic wounds	[48]
	Liposomal iodine, Repithel	Clinical 2 nd degree burns	[49]
	Iocide	Oral infections	[50]
Photodynamic therapy	ТВО	Mouse wound infections	[51]
	PEI-ce6	Mouse burn infections (A. baumannii)	[52]
	XF73	In vitro, ex vivo skin infection	[53]
	BB6	In vitro, mouse wound infection	[54]
	Sylsens B	Mouse burn infection S. aureus	[55]
	DP/hemin	Guinea pig burn infections S. aureus	[56]
	Cat PC	In vitro, veterinary infections	[57]
	EtNBSe	In vitro	[58]
Chitosan	hydrogel	Clinical 2 nd degree burns	[59]

Agent Class	Specific Agent	Application	Ref
	film	2 nd degree burns in rabbits	[60]
	bandage	Mouse burn infections (Psuedomonas, Proteus)	[61]
Antimicrobial peptide	Defensins	In vitro	[62]
	Demegel	Pseudomonas infected rat burns	[63]
	Histone H1.2	Pseudomonas infected rat burns	[64]
	Cecropin B	Pseudomonas infected mouse wounds	[65]
	rBPI	Clinical trial 2 nd degree burns	[66]
	Ceragenins	In vitro	[67]
Miscellaneous	Chlorhexidine (LM Miller)	Infected rat burns, clinical trial	[68,69
	Superoxidized water	In vitro	[70]
	BCTP nanoemulsion	In vitro	[71]
	FPQC	Mice burns, Pseudomonas	[72]
	Acidified nitrite	In vitro	[73]
	Phage therapy	Mice burns, Pseudomonas	[74]
	p38MAPK inhibitor	Rat burns, Pseudomonas	[75]
	Probiotics, <i>Lactobacillus</i> Clinical trial, 2 nd , 3 rd degree	Clinical trial, 2 nd , 3 rd degree	[76]
	Honey	Clinical trial, 2 nd degree	[77]
	Essential oils	In vitro	[78]
	MEBO	Clinical trial, 2 nd degree	[79]
	Papaya	Clinical trial, 2 nd , 3 rd degree	