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## The Endocannabinoid System as an Emerging Target of Pharmacotherapy

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### Abstract

The recent identification of cannabinoid receptors and their endogenous lipid ligands has triggered an exponential growth of studies exploring the endocannabinoid system and its regulatory functions in health and disease. Such studies have been greatly facilitated by the introduction of selective cannabinoid receptor antagonists and inhibitors of endocannabinoid metabolism and transport, as well as mice deficient in cannabinoid receptors or the endocannabinoid-degrading enzyme fatty acid amidohydrolase. In the past decade, the endocannabinoid system has been implicated in a growing number of physiological functions, both in the central and peripheral nervous systems and in peripheral organs. More importantly, modulating the activity of the endocannabinoid system turned out to hold therapeutic promise in a wide range of disparate diseases and pathological conditions, ranging from mood and anxiety disorders, movement disorders such as Parkinson's and Huntington's disease, neuropathic pain, multiple sclerosis and spinal cord injury, to cancer, atherosclerosis, myocardial infarction, stroke, hypertension, glaucoma, obesity/metabolic syndrome, and osteoporosis, to name just a few. An impediment to the development of cannabinoid medications has been the socially unacceptable psychoactive properties of plant-derived or synthetic agonists, mediated by CB<sub>1</sub> receptors. However, this problem does not arise when the therapeutic aim is achieved by treatment with a CB<sub>1</sub> receptor antagonist, such as in obesity, and may also be absent when the action of endocannabinoids is enhanced indirectly through blocking their metabolism or transport. The use of selective CB<sub>2</sub> receptor agonists, which lack psychoactive properties, could represent another promising avenue for certain conditions. The abuse potential of plant-derived cannabinoids may also be limited through the use of preparations with controlled composition and the careful selection of dose and route of administration. The growing number of preclinical studies and clinical trials with compounds that modulate the

endocannabinoid system will probably result in novel therapeutic approaches in a number of diseases for which current treatments do not fully address the patients' need. Here, we provide a comprehensive overview on the current state of knowledge of the endocannabinoid system as a target of pharmacotherapy.

## I. Introduction

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Marijuana, or cannabis, is the most widely used illicit drug in Western societies and also the one with the longest recorded history of human use. The popularity of marijuana as a recreational drug is due to its ability to alter sensory perception and cause elation and euphoria, most vividly described by the 19th century French poet, Charles Baudelaire, in his book *Les Paradis Artificiels* (Iversen, 2000). However, the ability of extracts of the hemp plant (*Cannabis sativa*) to cause a variety of medicinal effects unrelated to its psychoactive properties had been recognized as early as the third millennium BC, when Chinese texts described its usefulness in the relief of pain and cramps (Mechoulam, 1986). In ancient India, the anxiety-relieving effect of bhang (the Indian term for marijuana ingested as food) had been recorded more than 3000 years ago. The use of cannabis or hashish as a psychoactive substance reached Europe and the Americas through the Arab world in the 19th century. During the same period, cannabis extracts had gained widespread use for medicinal purposes until 1937, when concern about the dangers of abuse led to the banning of marijuana for further medicinal use in the United States. The rather turbulent history of marijuana and the recent resurgence of interest in its medicinal properties have been the subject of excellent reviews (Mechoulam, 1986; Iversen, 2000; Di Marzo et al., 2004; Howlett et al., 2004; Pertwee, 2005a; Piomelli, 2005; Di Marzo and Petrocellis, 2006; Mackie, 2006; Pagotto et al., 2006). Added to this interest is the emergence of the endocannabinoid system, offering not only new insights into the mechanisms underlying the therapeutic actions of plant-derived phytocannabinoids but also novel molecular targets for pharmacotherapy. In this overview, we will briefly summarize current thoughts about the role of endocannabinoids in a given physiological or pathological process and then survey attempts to exploit this role for therapeutic gain.

## II. The Pharmacology of Cannabinoids

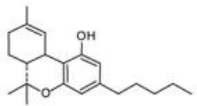
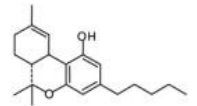
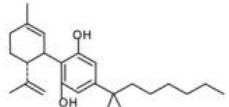
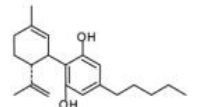
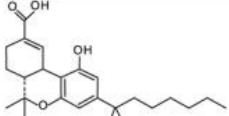
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### A. Cannabinoid Receptors and Ligands

Up until the last two decades, marijuana research was a rather esoteric field, of interest to a small number of scientists. A contributory factor was the highly lipophilic nature of the biologically active ingredients, which led to the notion that marijuana elicits its effects nonspecifically by perturbing membrane lipids (Lawrence and Gill, 1975). The first important breakthrough that ultimately led to a rejection of this concept was the identification by Gaoni and Mechoulam (1964) of the correct chemical structure of the main psychoactive ingredient of marijuana,  $\Delta^9$ -tetrahydrocannabinol (THC<sup>1</sup>), and the subsequent demonstration that bioactivity resides in the *l*-stereoisomer of this compound (Mechoulam and Gaoni, 1967), which is one of approximately 60 cannabinoids present in the plant (Dewey, 1986). This discovery stimulated the generation of a whole range of synthetic analogs in the 1970s that included not only compounds structurally similar to phytocannabinoids (Fig. 1A) but also analogs with different chemical structures, including classic and nonclassic cannabinoids and aminoalkylindoles (Fig. 1B) (Howlett et al., 2002), as well as the subsequently discovered endogenous arachidonic acid derivatives or endocannabinoids (Fig. 1C), which are discussed in more detail below. Studies of the biological effects of THC and its synthetic analogs revealed strict structural selectivity (Hollister, 1974) as well as stereo-selectivity (Jones et al., 1974), telltale signs of drug-receptor interactions. Definitive evidence for the existence of specific cannabinoid receptors was followed soon by the demonstration of high-affinity, saturable, stereospecific binding sites for the synthetic cannabinoid agonist [<sup>3</sup>H]CP-55,940 in mouse brain plasma membranes, which correlated with both the in vitro inhibition of adenylate

cyclase and the in vivo analgesic effect of the compound ([Devane et al., 1988](#)). The availability of a radioligand also allowed the mapping of cannabinoid receptors in the brain by receptor autoradiography ([Herkenham et al., 1991b](#)). This mapping turned out to be of key importance in the subsequent identification of an orphan G protein-coupled receptor (GPCR) as the brain receptor for cannabinoids ([Matsuda et al., 1990](#)), later named CB<sub>1</sub> receptor, based on the overlapping regional distribution of the mRNA for this GPCR and [<sup>3</sup>H]CP-55,940 binding sites. CB<sub>1</sub> receptors are the most abundant receptors in the mammalian brain but are also present at much lower concentrations in a variety of peripheral tissues and cells. A second cannabinoid GPCR, CB<sub>2</sub>, is expressed primarily in cells of the immune and hematopoietic systems ([Munro et al., 1993](#)) but recently were found to be present in the brain ([Van Sickle et al., 2005](#); [Gong et al., 2006](#)), in nonparenchymal cells of the cirrhotic liver ([Julien et al., 2005](#)), in the endocrine pancreas ([Juan-Pico et al., 2005](#)), and in bone ([Karsak et al., 2004](#); [Idris et al., 2005](#); [Ofek et al., 2006](#)). Two splice variants of CB<sub>1</sub> receptors have been also identified: CB<sub>1A</sub>, which has an altered amino-terminal sequence ([Shire et al., 1995](#)), and CB<sub>1B</sub>, which has an in-frame deletion of 33 amino acids at the amino terminus ([Ryberg et al., 2005](#)). The mRNAs of both splice variants are expressed at much lower levels than the CB<sub>1</sub> mRNA and, although the receptors expressed from the cDNAs have unique pharmacology ([Ryberg et al., 2005](#)), evidence for their natural expression has not been reported.

## A. Phytocannabinoids and their analogs

<p><math>\Delta^9</math>-tetrahydrocannabinol (<math>\Delta^9</math>-THC)</p> 	<p>CB<sub>1</sub> ≈ CB<sub>2</sub> agonist</p>	<p>Felder et al., 1995 Schowalter et al., 1996 Rinaldi-Carmona et al., 1994 Rhee et al., 1997</p>
<p>Cannabivarin (Cannabivarol, CBV)</p> 	<p>CB<sub>1</sub> ≈ CB<sub>2</sub> antagonist</p>	<p>Thomas et al., 2005</p>
<p>(-)-5'-(1,1-dimethylheptyl) cannabidiol (DMH-CBD)</p> 	<p>CB<sub>1</sub> ≈ CB<sub>2</sub> agonist inhibition of AEA uptake</p>	<p>Bisogno et al., 2001</p>
<p>(-)-Cannabidiol (CBD)</p> 	<p>no activity at CB<sub>1</sub> or CB<sub>2</sub> antagonism of non-CB<sub>1</sub> or non-CB<sub>2</sub> modulator of <math>\alpha_1</math>-adrenoreceptor inhibition of AEA uptake and metabolism</p>	<p>Schowalter et al., 1996 Járai et al., 1999 Pertwee et al., 2002 Bisogno et al., 2001</p>
<p>Ajulemic acid (AJA, CT-3, IP-751)</p> 	<p>CB<sub>1</sub> ≈ CB<sub>2</sub> agonist</p>	<p>Dyson et al., 2005</p>

## B. Synthetic cannabinoids

### Classical

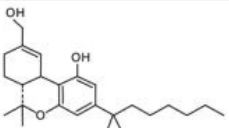
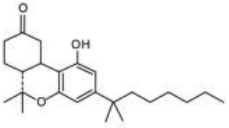
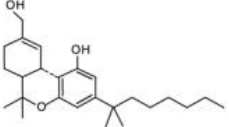
<p>(-) HU-210</p> 	<p>CB<sub>1</sub> ≈ CB<sub>2</sub></p>	<p>Felder et al., 1995 Schowalter et al., 1996</p>
<p>Nabilone</p> 	<p>CB<sub>1</sub> ≈ CB<sub>2</sub></p>	<p>Gareau et al., 1996</p>
<p>(+) HU-211 (dexanabinol)</p> 	<p>no activity at CB<sub>1</sub> or CB<sub>2</sub> noncompetitive NMDA antagonist</p>	<p>Schowalter et al., 1996 Bar-Joseph et al., 1994</p>

Fig. 1

The chemical structure and pharmacological activity of selected plant derived (A), synthetic (B), and endogenous cannabinoids (C).

An interesting twist on the steric selectivity of cannabinoid receptors has emerged through recent studies of the behaviorally inactive phytocannabinoid (–)-cannabidiol (CBD) and its synthetic analogs, which have negligible affinity for either CB<sub>1</sub> or CB<sub>2</sub> receptors. Paradoxically, some of the synthetic (+)-(-)-stereoisomers of these compounds were found to bind potently to both CB<sub>1</sub> and CB<sub>2</sub> receptors (Bisogno et al., 2001) but to display only peripheral and not centrally mediated cannabinoid-like

bioactivity, suggesting that they may act as antagonists rather than agonists at central, but not peripheral, CB<sub>1</sub> receptors (Fride et al., 2005).

Another ligand that displays central versus peripheral selectivity is ajulemic acid, a metabolite of THC that was found to have potent anti-inflammatory and analgesic properties without any overt behavioral or psychoactive effects (Burstein et al., 1992; Dyson et al., 2005; Mitchell et al., 2005). Ajulemic acid was reported to bind to both CB<sub>1</sub> and CB<sub>2</sub> receptors with reasonably high affinity ( $K_d$  100–200 nM) but only to activate the latter (Rhee et al., 1997), which may explain its unique and therapeutically attractive pharmacological profile. A more recent study indicated even higher affinities for CB<sub>1</sub> ( $K_i$  6 nM) and CB<sub>2</sub> receptors ( $K_i$  56 nM) and specified the role of CB<sub>1</sub> in mediating its antihyperalgesic activity in neuropathic pain (Dyson et al., 2005). This article also documented limited brain penetration of ajulemic acid compared with other cannabinoids, which may account for its favorable therapeutic profile. Ajulemic acid also binds to peroxisome proliferator-activated receptor  $\gamma$  receptors with low (micromolar) affinity, which was proposed to account for its effect on adipocyte differentiation (Liu et al., 2003b).

Among the 60 or so cannabinoids present in marijuana, only THC is psychoactive. However, some of the other constituents, such as cannabidiol, have well-documented biological effects of potential therapeutic interest, such as antianxiety, anticonvulsive, anti-nausea, anti-inflammatory and antitumor properties (Mechoulam et al., 2002c; Grotenhermen, 2004; Vaccani et al., 2005). Cannabidiol does not significantly interact with CB<sub>1</sub> or CB<sub>2</sub> receptors, and its actions have been attributed to inhibition of anandamide degradation or its antioxidant properties (Mechoulam and Hanus, 2002; Mechoulam et al., 2002c), or an interaction with as yet unidentified cannabinoid receptors (see below). Another marijuana constituent of potential therapeutic interest is tetrahydrocannabivarin (Markus, 1971), which has recently been shown to have CB<sub>1</sub> antagonist properties (Thomas et al., 2005).

In addition to CB<sub>1</sub> and CB<sub>2</sub> receptors, pharmacological evidence has been accumulating over the years to support the existence of one or more additional receptors for cannabinoids (reviewed in Begg et al., 2005). Two of these possibilities have been more extensively explored: an endothelial site involved in vasodilation and endothelial cell migration (Járai et al., 1999; Begg et al., 2003; Mo et al., 2004), and a presynaptic site on glutamatergic terminals in the hippocampus mediating inhibition of glutamate release (Hájos et al., 2001). Responses elicited at both of these sites were reported to survive genetic ablation of CB<sub>1</sub> receptors, yet be sensitive to inhibition by the CB<sub>1</sub> antagonist SR141716 or by pertussis toxin but not by the CB<sub>1</sub> antagonist AM251 (Járai et al., 1999; Hájos and Freund, 2002; Ho and Hiley, 2003; Offertáler et al., 2003; O'Sullivan et al., 2004a,b). However, the two sites are apparently different. The aminoalkylindol WIN 55,212-2 was found to be an agonist and capsazepine an antagonist at the hippocampal (Hájos and Freund, 2002) but not at the endothelial receptor (Wagner et al., 1999; Mukhopadhyay et al., 2002). On the other hand, certain atypical cannabinoids with no affinity for CB<sub>1</sub> or CB<sub>2</sub> receptors behave as agonists (abnormal cannabidiol, O-1602) or antagonists at the endothelial receptor (cannabidiol, O-1918) but not at the hippocampal receptor (Begg et al., 2005). Arachidonoyl-L-serine, an endogenous lipid discovered in rat brain, has been found to be a vasodilator acting at the endothelial cannabinoid receptor (Milman et al., 2006), although its activity at the hippocampal receptor has not yet been evaluated. The existence of this latter receptor has recently been called into question, as the ability of WIN 55,212-2 to suppress the same excitatory synapse as studied by Hájos et al. (2001) was found to be absent in two different strains of CB<sub>1</sub> knockout mice, yet present in their respective wild-type controls (Takahashi and Castillo, 2006). Atypical cannabinoid receptors with pharmacological properties similar to those of the endothelial receptor have been postulated to exist on microglia, where they mediate microglial migration (Walter et al., 2003), and on neurons of the mouse vas deferens (Pertwee et al., 2002, 2005c). Activation of this latter receptor by the CBD analog 7-OH-dimethylheptyl

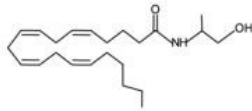
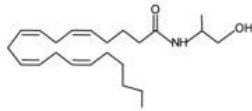
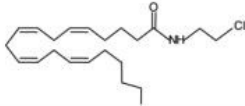
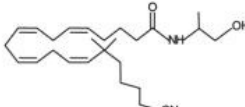
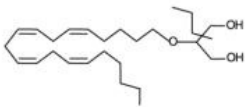
CBD, which is inactive at CB<sub>1</sub>, CB<sub>2</sub>, or transient receptor potential vanilloid type 1 (TRPV<sub>1</sub>) receptors, inhibits electrically evoked contractions of the vas deferens, and the effect is selectively inhibited by CBD itself. A brain cannabinoid receptor distinct from CB<sub>1</sub> was also indicated by the ability of anandamide and WIN 55,212-2, but not other agonists, to stimulate GTPγS binding in brain plasma membranes from CB<sub>1</sub> knockout mice ([Breivogel et al., 2001](#)).

Of interest are recent findings reported in the patent literature that the orphan receptor GPR-55 ([Sawzdargo et al., 1999](#)) recognizes a variety of cannabinoid ligands, but not WIN 55,212-2 ([Brown and Wise, 2003](#); [Drmota et al., 2004](#)). However, GPR-55 is apparently not expressed in the vascular endothelium and is sensitive to HU-210 ([Drmota et al., 2004](#)), a potent synthetic cannabinoid devoid of vasorelaxant properties ([Wagner et al., 1999](#)). Furthermore, it couples to G<sub>12</sub>/G<sub>13</sub> and ρ kinase, which have been linked to vasoconstrictor rather than vasodilator responses. This suggests that GPR-55 is not the abnormal cannabidiol-sensitive endothelial receptor. Mice deficient in GPR-55 will help in defining the biological functions of this novel cannabinoid-sensitive receptor.

Anandamide has been found to be an agonist ligand for the TRPV<sub>1</sub> ion channel, although its affinity in the low micromolar range is lower than its affinity for CB<sub>1</sub> receptors (reviewed by [van der Stelt and Di Marzo, 2004](#)). An in vitro study in rat mesenteric arteries provided evidence that the endothelium-independent component of anandamide-induced vasodilation is mediated via activation of capsaicin-sensitive TRPV<sub>1</sub> in sensory nerve terminals. This triggers the release of CGRP, which then dilates the artery by activation of calcitonin gene-related peptide receptors on the vascular smooth muscle ([Zygmunt et al., 1999](#)). However, this mechanism does not contribute to the in vivo hypotensive action of anandamide, which is similar in wild-type and TRPV<sub>1</sub><sup>-/-</sup> mice ([Pacher et al., 2004](#)).

Both CB<sub>1</sub> and CB<sub>2</sub> receptors are G protein-coupled receptors. Surprisingly, they share little sequence homology, only 44% at the protein level or 68% in the transmembrane domains, which are thought to contain the binding sites for cannabinoids ([Lutz, 2002](#)). Despite this, THC and most synthetic cannabinoids have similar affinities for the two receptors, and only recently did synthetic ligands that discriminate between CB<sub>1</sub> and CB<sub>2</sub> receptors emerge. These include agonists as well as antagonists, as listed in [Fig. 2](#). The development of potent and highly selective CB<sub>1</sub> and CB<sub>2</sub> receptor antagonists ([Rinaldi-Carmona et al., 1994, 1998](#)) is particularly noteworthy as it provided critically important tools to explore the physiological functions of endocannabinoids. For example, as it will be discussed later in this review, the appetite-reducing effects of the CB<sub>1</sub> antagonist SR141716 in various rodent models was the first sign to suggest that endocannabinoids may be tonically active orexigenic agents, representing the endogenous counterpart of the “munchies” caused by marijuana smoking.

## A. Cannabinoid receptor agonists

CB <sub>1</sub> receptor selective		K <sub>i</sub> (nM) for		Reference
		CB <sub>1</sub>	CB <sub>2</sub>	
<b>R-(+)-methanandamide</b>		17.9 20 28.3	868 815 868	Lin et al., 1998 Khanolkar et al., 1996 Goutopoulos et al., 2001
<b>Arachidonoyl 2'-chloroethylamide (ACEA)</b>		1.4 5.29	>2,000 195	Hillard et al., 1999 Lin et al., 1998
<b>O-1812</b>		3.4	3,870	Di Marzo et al., 2001a
<b>2-Arachidonoyl glycerol ether</b>		21.2	>3,000	Hanus et al., 2001

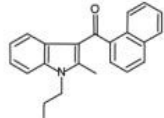
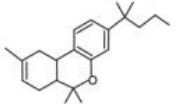
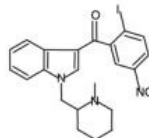
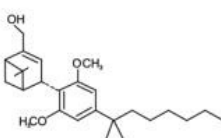
CB <sub>2</sub> receptor selective				
<b>JWH 015</b>		383	13.8	Showalter et al., 1996
<b>JWH 133</b>		677	3.4	Huffman et al., 1999
<b>AM 1241</b>		280	3.4	Ibrahim et al., 2003
<b>HU-308</b>		>10,000	22.7	Hanus et al., 1999

Fig. 2

Selective agonists (A) and antagonists (B) of CB<sub>1</sub> and CB<sub>2</sub> receptors.

However, these antagonists, as well as most of the other CB<sub>1</sub> and CB<sub>2</sub> antagonists developed to date, have inverse agonist properties (Bouaboula et al., 1997, 1999), so their effects do not necessarily reflect reversal of the tonic action of an endocannabinoid. For this reason, the development of CB<sub>1</sub> and CB<sub>2</sub> receptor-deficient mouse strains (Ledent et al., 1999; Zimmer et al., 1999; Buckley et al., 2000; Marsicano et al., 2002b; Robbe et al., 2002) was similarly important, as the use of these animals in combination with receptor antagonists can reinforce the putative regulatory roles of endocannabinoids.

More recently, the development of conditional mutant mice that lack the expression of CB<sub>1</sub> receptors only in certain types of neurons represents another milestone, as it allows linking of specific neuronal populations with a well-defined cannabinoid-modulated behavior ([Marsicano et al., 2003](#)).

## B. Cannabinoid Receptor Signaling

CB<sub>1</sub> and CB<sub>2</sub> receptors couple primarily to the G<sub>i/o</sub> subtypes of G protein, and their signaling is remarkably complex. Although coupling to adenylate cyclase through G<sub>i/o</sub> usually results in inhibition of cyclase activity through the release of G<sub>iα</sub> isoforms, cannabinoids can also stimulate isoforms 2, 4, or 7 of adenylate cyclase via the release of βγ subunits ([Rhee et al., 1998](#)). Activation of adenylate cyclase also occurs when CB<sub>1</sub> and dopamine D<sub>2</sub> receptors are simultaneously activated ([Glass and Felder, 1997](#)), probably as a result of heterodimerization of these two types of receptors ([Kearn et al., 2005](#)). Although direct evidence for the coupling of CB<sub>1</sub> receptors to G<sub>q/11</sub> had until recently been lacking ([Howlett, 2004](#)), the agonist WIN 55,212-2, but not other cannabinoids, was recently reported to increase intracellular calcium in cultured hippocampal neurons and in human embryonic kidney 293 cells via coupling to G<sub>q/11</sub> proteins ([Lauckner et al., 2005](#)). Receptor dimerization may facilitate such coupling, which may account for CB<sub>1</sub>-mediated mobilization of intracellular calcium in NG108-15 neuroblastoma glioma cells ([Sugiura et al., 1999](#)). Cannabinoids can also inhibit different types of calcium channels ([Mackie and Hille, 1992](#); [Gebremedhin et al., 1999](#)) and activate certain potassium channels ([Mackie et al., 1995](#)) via G protein βγ subunits ([Ikeda, 1996](#)). Cannabinoids can activate members of all three families of multifunctional mitogen-activated protein kinases, including p44/42 MAP kinase ([Wartmann et al., 1995](#); [Davis et al., 2003](#)), p38 kinase ([Liu et al., 2000](#); [Derkinderen et al., 2001](#)), and JUN-terminal kinase ([Liu et al., 2000](#); [Rueda et al., 2000](#)) and activate the phosphatidylinositol-3-kinase pathway ([Gómez Del Pulgar et al., 2002a](#)). These effects could be via G protein activation ([Galve-Roperh et al., 2002](#); [Davis et al., 2003](#)) or pathways independent of G proteins via other adaptor proteins ([Sánchez et al., 2001b](#)). Another G protein-independent pathway activated by cannabinoids involves G protein-coupled receptor kinase-3 and β-arrestin-2, which are required for desensitization, but not for internalization, of CB<sub>1</sub> receptors, and the related development of tolerance ([Jin et al., 1999](#)). Cannabinoids can also regulate the activity of phosphatases, as exemplified by the CB<sub>1</sub>-mediated regulation of calcineurin (protein phosphatase 2b) ([Cannich et al., 2004](#)) or the activation of mitogen-activated protein kinase phosphatase 1, which plays an important role in the anti-inflammatory action of anandamide ([Eljaschewitsch et al., 2006](#)).

Different structural classes of cannabinoid receptor agonists have the unique ability to activate different signaling cascades which, in turn, influences agonist efficacy. Using an in situ receptor/G protein reconstitution technique, CB<sub>1</sub> receptors were found to efficiently couple and activate both G<sub>i</sub> and G<sub>o</sub>, whereas CB<sub>2</sub> receptors only activated G<sub>o</sub>. Furthermore, the efficacy of a given agonist was different whether CB<sub>1</sub> receptors coupled to G<sub>i</sub> or G<sub>o</sub>, demonstrating agonist-selective G protein signaling ([Glass and Northup, 1999](#)). [Prather et al. \(2000\)](#) found that the aminoalkylindol agonist WIN 55,212-2 activated different G<sub>iα</sub> subunits with markedly different potencies. Even more striking is the recent finding that demonstrates cannabinoid agonist-selective activation of different G<sub>iα</sub> subunits ([Mukhopadhyay and Howlett, 2005](#)). A possible practical implication of such findings is that unique therapeutic profiles may be achieved through the use of different agonists for the same receptor, and such profiles may differ from one target tissue to the other, depending on the pattern of G protein subunit expression.

At least part of this agonist selectivity in G protein activation may be related to the existence of distinct binding sites on CB<sub>1</sub> receptors for different classes of ligands, as documented by site-directed mutagenesis and molecular modeling studies (see [Reggio, 2003](#)). These studies indicate that a K3.28A



mutation in the third transmembrane domain caused a more than 1000-fold loss in affinity and loss of efficacy for anandamide and nonclassic cannabinoids, without affecting the affinity for WIN 55,212-2 ([Song and Bonner, 1996](#)). In contrast, mutations at different sites in the third, fifth, and sixth transmembrane helices (F3.36A, W5.43A, and W6.48A) affected the binding of WIN 55,212-2 and SR141716, but not anandamide ([McAllister et al., 2003](#)).

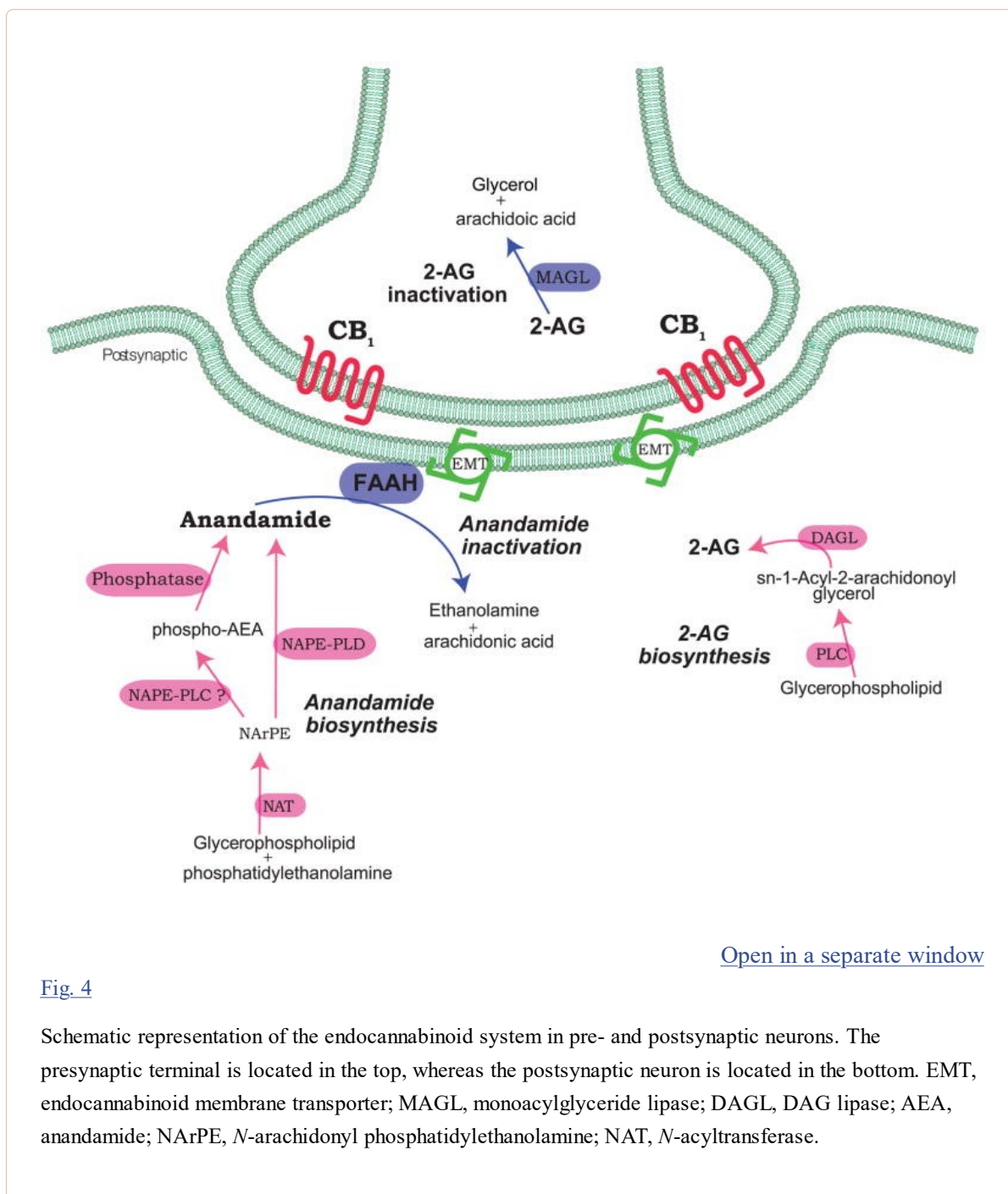
Another important feature of cannabinoid signaling in the brain is the lack of correlation between the density of CB<sub>1</sub> receptors in a given brain region and the efficiency of receptor coupling, as determined by GTPγS binding ([Breivogel et al., 1997](#)), which may explain why functionally important responses can be triggered in brain regions with very sparse CB<sub>1</sub> receptor expression, such as the brainstem ([Rademacher et al., 2003](#)) or the hypothalamus ([Jamshidi and Taylor, 2001](#)). [Selley et al. \(2001\)](#) have shown that the reduction in CB<sub>1</sub> receptor density in CB<sub>1</sub> heterozygote mice was compensated for by an increase in receptor/G protein coupling efficiency for some, but not other, agonists. Although the underlying mechanisms for such compensation are not clear, differences in the degree of receptor multimerization ([Mackie, 2005](#)), or changes in signal amplification are possibilities. Recent observations indicate that a considerable proportion of the psychomotor effect of cannabinoids can be accounted for by a signaling cascade in striatal projection neurons involving protein kinase A-dependent phosphorylation of DARPP-32, achieved via modulation of dopamine D<sub>2</sub> and adenosine A<sub>2A</sub> transmission ([Andersson et al., 2005](#)). This represents a unique form of amplification of CB<sub>1</sub> signaling, as phosphorylation of DARPP-32 at Thr-34 amplifies downstream signaling via inhibition of protein phosphatase-1 ([Greengard, 2001](#)). It would be interesting to test whether the efficiency of CB<sub>1</sub> coupling to DARPP-32 is affected by cellular receptor density.

### C. Endocannabinoids

The existence of specific receptors in mammalian cells that recognize a plant-derived substance rekindled the question raised two decades earlier, after brain receptors for morphine had been first described, i.e., is there an endogenous ligand? A positive answer was provided in 1992 by the report by Devane et al. describing the isolation from porcine brain of the lipid arachidonoyl ethanolamide, named anandamide, which bound to the brain cannabinoid receptor with reasonably high affinity and mimicked the behavioral actions of THC when injected into rodents ([Devane et al., 1992](#)). Three years later a second endocannabinoid, 2-arachidonoylglycerol (2-AG), was discovered independently by [Mechoulam et al. \(1995\)](#) and [Sugiura et al. \(1995\)](#). Since then, a number of related endogenous lipids with endocannabinoid-like activity have been reported ([Fig. 1c](#)), but follow-up studies about biosynthesis, cellular transport, metabolism, and biological function have focused on anandamide and 2-AG, with much less information available about the other compounds with endocannabinoid-like properties. The biochemical aspects of endocannabinoids have been recently reviewed by [Bisogno et al. \(2005\)](#).

Anandamide is a partial or full agonist of CB<sub>1</sub> receptors, depending on the tissue and biological response measured. Although it also binds CB<sub>2</sub> receptors, it has very low efficacy and may act as an antagonist ([Gonsiorek et al., 2000](#)). The in vivo biosynthesis of anandamide ([Fig. 4](#)) is believed to occur through the enzymatic hydrolysis catalyzed by a phospholipase D of a membrane lipid precursor, *N*-arachidonoyl phosphatidylethanolamide (NAPE) ([Schmid et al., 1983](#)), which itself is generated by the enzymatic transfer of arachidonic acid in the *sn*-1 position in phosphatidylcholine to the amide group of PE ([Di Marzo et al., 1994](#); [Cadas et al., 1997](#)). Although a specific transacylase for the latter reaction has not yet been identified, a NAPE-specific PLD has recently been cloned ([Okamoto et al., 2004](#)). It is not yet known, however, whether NAPE-PLD is obligatory for the biosynthesis of anandamide, which could make it an attractive target of drug therapy when reduction of tissue anandamide would be of benefit. Indeed, there may be parallel pathways for the generation of anandamide from NAPE. A secretory PLA<sub>2</sub> that can catalyze the hydrolysis of *N*-acyl-PE to *N*-acyl-lysoPE, which is then acted on by a

lysoPLD to generate *N*-acyl-ethanolamides, including anandamide, was recently identified in the stomach (Sun et al., 2004). An alternative parallel pathway has been identified in our laboratory in RAW246.7 macrophages. This involves hydrolysis of NAPE to phosphoanandamide by a PLC, followed by dephosphorylation through a phosphatase (Liu et al., 2006). This latter pathway rather than PLD is the target of regulation by bacterial endotoxin, which increases anandamide synthesis in macrophages (Varga et al., 1998; Liu et al., 2003a). The existence of this pathway may also account for the recent finding that anandamide tissue levels are unchanged in NAPE-PLD knockout compared with wild-type mice (Leung et al., 2006).



**Fig. 4**

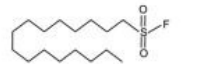
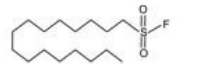
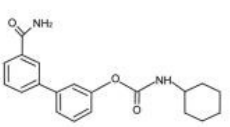
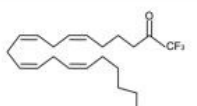
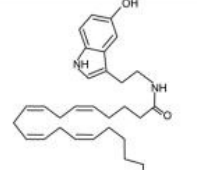
Schematic representation of the endocannabinoid system in pre- and postsynaptic neurons. The presynaptic terminal is located in the top, whereas the postsynaptic neuron is located in the bottom. EMT, endocannabinoid membrane transporter; MAGL, monoacylglyceride lipase; DAGL, DAG lipase; AEA, anandamide; NArPE, *N*-arachidonoyl phosphatidylethanolamine; NAT, *N*-acyltransferase.

2-AG is generated from diacylglycerol (DAG) by DAG lipase selective for the *sn*-1 position (Fig. 4). DAG, an intracellular second messenger that activates protein kinase C, can be generated from

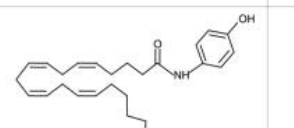
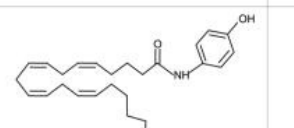
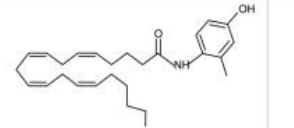
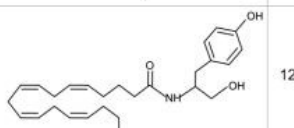
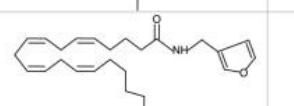
phosphoinositides by a phosphoinositide-specific PLC or from phosphatidic acid by phosphatidic acid phosphohydrolase (Bisogno et al., 2005). Two DAG lipase isozymes,  $\alpha$  and  $\beta$ , have been cloned (Bisogno et al., 2003). In the adult brain they are localized in the postsynaptic plasma membrane, in line with their putative role in generating 2-AG involved in retrograde transmission.

Basal levels of 2-AG in the brain are approximately 2 orders of magnitude higher than the levels of anandamide. Despite this, stimulus-induced release resulting in detectable extracellular levels could be demonstrated only for anandamide and not for 2-AG in an *in vivo* microdialysis study (Giuffrida et al., 1999). This finding illustrates that, despite growing interest in endocannabinoids and their roles as retrograde neurotransmitters (Wilson and Nicoll, 2002; Chevalleyre et al., 2006), the mechanism of their release is not well understood. Like prostanoids, endocannabinoids are not stored but generated on demand in response to a depolarization-induced rise in intracellular calcium or activation of various metabotropic receptors (Varma et al., 2001; Kim et al., 2002; Witting et al., 2004; Di et al., 2005a,b). A putative membrane endocannabinoid transporter involved in the cellular uptake of endocannabinoids (see below) may also be involved in their release. This is suggested by the ability of a transport inhibitor to prevent the release of intracellularly applied anandamide (Maccarrone et al., 2000a; Gerdeman et al., 2002).

Anandamide present in the extracellular space is accumulated by neurons and other cells by facilitated diffusion. This process is driven by its transmembrane concentration gradient, is saturable and temperature-dependent, and does not require ATP or sodium ions. Most importantly for the topic of the present review, anandamide uptake is selectively inhibited by a variety of structural analogs, which suggests the existence of a saturable cellular component involved in anandamide transport (Beltramo et al., 1997; Bisogno et al., 1997; Hillard and Jarrahian, 2000; Maccarrone et al., 2000a). However, a specific anandamide transporter protein has yet to be cloned, and it has been proposed that intracellular degradation of anandamide by fatty acid amide hydrolase (FAAH) is sufficient to account for anandamide uptake in long incubation periods (Glaser et al., 2003). Studies with cells isolated from FAAH<sup>+/+</sup> and FAAH<sup>-/-</sup> mice did not resolve this issue, as the absence of FAAH was found not to affect anandamide uptake (Fegley et al., 2004) or to reduce it substantially (Ortega-Gutierrez et al., 2004), albeit under different experimental conditions. Nevertheless, a FAAH-independent component of anandamide uptake, inhibited by the compound UCM707, was detected in the latter study, supporting the notion of a protein other than FAAH being involved. This notion is also supported by the emergence of a number of synthetic transport inhibitors, the potencies of which to inhibit anandamide uptake does not correlate with their affinities for CB<sub>1</sub>, CB<sub>2</sub>, or TRPV<sub>1</sub> receptors or their potencies to inhibit FAAH (Fig. 3). However, in view of the important role of FAAH in generating the transmembrane concentration gradient for anandamide, the possibility that a noncatalytic region of FAAH or a FAAH-associated protein may act as anandamide transporter cannot be excluded. Interestingly, the elucidation of the crystal structure of FAAH revealed several channel-like regions in the enzyme, granting it simultaneous access to both the cytosolic and membrane domains (Bracey et al., 2002). Against this possibility, however, is the recent report that the novel, high affinity anandamide transport inhibitor LY2318912 binds with similar  $K_d$  and  $b_{max}$  values to membranes from HeLa cells devoid of FAAH or transfected with FAAH, pointing to a binding site independent of the FAAH molecule (Moore et al., 2005). Arguments for and against the existence of a bidirectional anandamide transporter have been recently reviewed (Hillard and Jarrahian, 2003; Fowler et al., 2004; Mc-Farland and Barker, 2004; Glaser et al., 2005).

FAAH inhibitors (FAAHI)		$K_d$ (nM) for					Reference	
		CB <sub>1</sub>	CB <sub>2</sub>	FAAHI	ACUI	TRPV <sub>1</sub>		MGL
<b>Palmitylsulphonyl fluoride (AM 374)</b>		520		13-50				Deutsch et al., 1997a
<b>URB 597</b>				113 0.5-4.6		>1 mM	>1 mM >30 mM	Lichtman et al., 2004a Kathuria et al., 2003
<b>Arachidonyl Trifluoromethyl Ketone (AATFMK)</b>		0.65-2.5		900 1,000 4,000			2,500 30,000	Deutsch et al., 1997a,b Ueda et al., 1995 Beltramo et al., 1997 Koutek et al., 1994 Dinh et al., 2002b Goparaju et al., 1999
<b>Arachidonoyl Serotonin</b>		inactive		560-1200				Bisogno et al., 1998

AEA cellular uptake inhibitors (ACUI)		$K_d$ (nM) for					Reference	
		CB <sub>1</sub>	CB <sub>2</sub>	FAAHI	ACUI	TRPV <sub>1</sub>		MGL
<b>AM404</b>				inhibit	4,000	agonist		Beltramo et al., 1997 De Petrocellis et al., 2000 Jarrahian et al., 2000
<b>VDM 11</b>				inhibit	10,000			De Petrocellis et al., 2000
<b>OMDM 1</b>		12,000			2,4000	>50,000	>10,000	Ortar et al., 2003 Fowler et al., 2004
<b>UCM 707</b>			67	30,000	80 25,000			Lopez-Rodriguez et al., 2001; 2003 Fowler et al., 2004

[Open in a separate window](#)

**Fig. 3**

The structure and pharmacological specificity of inhibitors of FAAH and of endocannabinoid membrane transport.

In some *in vivo* studies, treatment with transport inhibitors unmasked cannabinoid-like tonic effects on pain sensitivity, anxiety-like behaviors, locomotor activity, and muscle spasticity, which is an indication of the potential therapeutic usefulness of such compounds (Moore et al., 2005; Bortolato et al., 2006; La Rana et al., 2006). Similar and more pronounced effects have been reported in response to treatment with FAAH inhibitors, as discussed below.

In contrast to the unsettled status of anandamide transport and a putative transporter protein, the unique role of FAAH in the *in vivo* degradation of anandamide has been extensively documented (reviewed in [McKinney and Cravatt, 2005](#)). Initial evidence for a membrane-associated enzyme in the liver that hydrolyzes *N-N*-acyl ethanolamides ([Schmid et al., 1985](#)) was followed by the cloning of FAAH ([Cravatt et al., 1996](#)) and the identification of its crystal structure in complex with an active site-directed inhibitor ([Bracey et al., 2002](#)). The unique role of FAAH in terminating signaling by anandamide was indicated by the phenotype of FAAH knockout mice, which displayed 10 to 15 times elevated levels of anandamide across the brain, supersensitivity to the actions of exogenous anandamide, and the appearance of tonic signaling by endogenous anandamide, resulting in CB<sub>1</sub> receptor-mediated hypoalgesia ([Cravatt et al., 2001](#); [Lichtman et al., 2004b](#)), reduced anxiety ([Kathuria et al., 2003](#)), antidepressant activity (Gobbi et al., 2006), and lowering of blood pressure in different models of experimental hypertension ([Bátkai et al., 2004b](#)). [Cravatt et al. \(2004\)](#) were able to resolve the relative roles of central versus peripheral fatty acid amides by generating mice deficient in FAAH in peripheral tissues only. These mice did not display the hypoalgesia observed in mice with global deficiency in FAAH, but had a similar anti-inflammatory phenotype, indicating that the latter was mediated by elevated fatty acid amides in peripheral tissues ([Cravatt et al., 2004](#)). Interestingly, another amidohydrolase catalyzing the same reaction as FAAH but at acidic pH was recently identified and cloned ([Tsuboi et al., 2005](#)). This lysosomal enzyme is structurally unrelated to FAAH and is widely distributed in tissues, with highest levels in the lung, and has been recently shown to contribute to the physiological degradation of anandamide in macrophages but not in the brain ([Sun et al., 2005](#)).

Although 2-AG is also hydrolyzed by FAAH under *in vitro* conditions ([Goparaju et al., 1998](#); [Lang et al., 1999](#)), *in vivo* it is not a substrate of FAAH, as indicated by the unchanged brain levels of 2-AG in wild-type and FAAH<sup>-/-</sup> mice ([Osei-Hyiaman et al., 2005a](#)). 2-AG is hydrolyzed *in vivo* by a monoacylglyceride lipase (MGL) ([Dinh et al., 2002a,b](#); [Saario et al., 2004](#)). A study of the ultrastructural distribution of FAAH and MGL revealed that in the hippocampus, cerebellum, and amygdala, FAAH is located postsynaptically, whereas MGL is localized in presynaptic axon terminals, including terminals of GABAergic interneurons ([Gulyas et al., 2004](#)). Correspondingly, functional studies in hippocampus indicate that depolarization-induced suppression of inhibition (DSI) is unaffected by pharmacological blockade of FAAH ([Kim and Alger, 2004](#)), but it is potentiated by blocking MGL ([Kim and Alger, 2004](#); [Makara et al., 2005](#)), in agreement with an earlier study implicating 2-AG rather than anandamide in synaptic plasticity in the hippocampus ([Stella et al., 1997](#)). Further evidence supporting the role of 2-AG as the retrograde transmitter involved in synaptic plasticity is the preferential postsynaptic distribution of the major 2-AG biosynthetic enzyme, diacylglycerol lipase  $\alpha$ , in hippocampus and cerebellum (Katona et al., 2006; [Yoshida et al., 2006](#)).

However, the behavioral consequences of DSI and its modulation remain unclear: selective knockout of CB<sub>1</sub> receptors from GABAergic interneurons was found to abolish DSI and long-term depression (LTD) of inhibitory synapses, whereas the classic behavioral responses to THC remained unaffected in these animals ([Monory et al., 2005](#)). Therefore, at this point it is difficult to predict the potential therapeutic usefulness of selective MGL inhibitors.

### III. The Endocannabinoid System as Therapeutic Target in Pathophysiological Conditions

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#### A. Diseases of Energy Metabolism

**1. Endocannabinoids and Appetite Regulation** It has been known since antiquity that use of cannabis in its various forms increases appetite, particularly for palatable foods, and can also result in significant weight gain ([Donovan, 1845](#); [Berry and Mechoulam, 2002](#)). Following the identification of THC as the

main psychoactive principle in marijuana, the appetite-promoting effect of smoked marijuana could be attributed to THC even before the identification of specific cannabinoid receptors ([Hollister, 1971](#); [Greenberg et al., 1976](#)). Animal studies also documented the ability of THC to promote food intake, although consistent effects were only seen with relatively low doses ([Abel, 1975](#)), most likely because the significant sedation and motor impairment seen with higher doses interferes with the animals' ability to initiate feeding. Variability in the observed changes in THC-induced food intake may also relate to the feeding state of the animal, the orexigenic effect being optimal in presatiated animals with low basal levels of food intake ([Williams et al., 1998](#)). After the discovery of specific cannabinoid receptors and the introduction of selective antagonists, the increase in food intake caused by THC could be linked to CB<sub>1</sub> receptors, as it was blocked by the selective CB<sub>1</sub> antagonist SR141716, but not by the CB<sub>2</sub> antagonist SR144528 ([Williams and Kirkham, 2002](#)).

The discovery of endocannabinoids has raised the question of their potential involvement in the physiological control of appetite and energy metabolism. This subject has been the topic of a number of recent reviews ([Di Marzo and Matias, 2005](#); [Kirkham, 2005](#); [Sharkey and Pittman, 2005](#); [Pagotto et al., 2006](#)), and only a brief summary is provided here. The first indication of a role for endocannabinoids in appetite control was the documented ability of low doses of anandamide to increase food intake, when administered either systemically ([Williams and Kirkham, 1999](#); [Hao et al., 2000](#)) or into the ventromedial hypothalamus ([Jamshidi and Taylor, 2001](#)), and this effect could be attributed to stimulation of CB<sub>1</sub> receptors ([Williams and Kirkham, 1999](#)). Similar increases in food intake can be elicited by 2-AG administered systemically or into the nucleus accumbens shell region ([Kirkham et al., 2002](#)) or into the lateral hypothalamus ([Kirkham and Williams, 2001a](#)). Sites for the orexigenic actions of endocannabinoids in both the hypothalamus and the limbic forebrain suggest their involvement in both the homeostatic and hedonic control of eating ([Harrold and Williams, 2003](#); [Vickers and Kennett, 2005](#)). Interestingly, endocannabinoid activation of hypothalamic centers, such as the paraventricular nucleus, may also occur indirectly via CB<sub>1</sub> receptors on peripheral afferent nerve terminals ([Gomez et al., 2002](#)), most likely located in the gastrointestinal tract. Such an "indirect" pathway is compatible with recent findings that CB<sub>1</sub> mRNA is present in cholecystokinin-containing neurons in the nodose ganglion, where CB<sub>1</sub> mRNA expression is up-regulated by fasting and down-regulated by refeeding ([Burdyga et al., 2004](#)).

Studies with antagonists provide more direct support for a regulatory function of endocannabinoids on feeding. Treatment of rats with SR141716 and the closely related CB<sub>1</sub> antagonist AM251 reduced food intake under free-feeding ([Arnone et al., 1997](#); [Colombo et al., 1998a](#); [Simiand et al., 1998](#); [Chambers et al., 2003](#); [Shearman et al., 2003](#)) or operant conditions ([Freedland et al., 2000](#); [McLaughlin et al., 2003](#)), suggesting antagonism of the tonic orexigenic effect of an endocannabinoid. However, SR141716 and AM251 are inverse agonists ([Gifford and Ashby, 1996](#); [Bouaboula et al., 1997](#)), which may be an alternative mechanism by which they reduce food intake.

Definitive evidence for the involvement of endocannabinoids in the control of food intake has been provided through the use of CB<sub>1</sub> receptor-deficient mice. In a study from our laboratory, food-deprived CB<sub>1</sub> knockout mice were found to eat less than their wild-type littermates, and their food intake was unaffected by SR141716 treatment, whereas in wild-type mice SR141716 reduced food intake to the levels seen in the knockout mice ([Di Marzo et al., 2001b](#)). Similar findings have been subsequently reported by others ([Wiley et al., 2005](#)). This indicates that part of the hunger-induced increase in food intake is mediated by endocannabinoids acting at CB<sub>1</sub> receptors. CB<sub>1</sub> knockout mice are also resistant to overeating caused by neuropeptide Y (NPY) ([Poncelet et al., 2003](#)), and SR141716 inhibits the hyperphagia of leptin-deficient mice even in the absence of temporary food deprivation ([Di Marzo et al., 2001b](#)). This latter finding suggests that the absence of leptin results in increased endocannabinoid

activity. Indeed, hypothalamic levels of endocannabinoids were elevated in leptin-deficient mice and rats and reduced after leptin treatment, suggesting that endocannabinoids are part of the leptin-regulated neural circuitry involved in appetite regulation ([Di Marzo et al., 2001b](#)). Endogenous leptin may similarly suppress endocannabinoid levels, as indicated by our recent unpublished findings using mice with obesity induced by a high-fat diet, which have elevated plasma leptin levels proportional to their increased fat mass. Anandamide levels were significantly lower in the obese mice compared with their lean controls in the hypothalamus, limbic forebrain, and amygdala, with no difference in the cerebellum. Furthermore, there was a significant inverse correlation between plasma leptin levels and anandamide levels in the above three brain regions involved in appetite control but not in the cerebellum.

A possible hypothalamic site for an interaction between leptin and endocannabinoids is the lateral hypothalamus, where CB<sub>1</sub> receptors are present in orexin- and melanin-concentrating hormone (MCH)-containing neurons ([Cota et al., 2003](#)), which also express functional leptin receptors ([Hübschle et al., 2001](#); [Iqbal et al., 2001](#)). These neurons project to dopaminergic neurons in the ventral tegmental area ([Fadel and Deutch, 2002](#)), where they modulate the mesolimbic dopaminergic pathway involved in food reward. Thus, they could also represent a site of integration of hypothalamic and extrahypothalamic structures involved in the orexigenic effect of endocannabinoids. The MCH-containing neurons are tonically inhibited by GABAergic interneurons. [Jo et al. \(2005\)](#) recently demonstrated that this inhibitory tone can be suppressed by the depolarization-induced release of endocannabinoids from the MCH neurons and their retrograde activation of presynaptic CB<sub>1</sub> receptors on the GABAergic interneurons. The resulting increase in the activity of MCH neurons may contribute to the *in vivo* appetitive effect of endocannabinoids. Furthermore, this DSI could be blocked by leptin through inhibition of voltage-gated calcium channels in the MCH neurons, whereas it was increased 6-fold in leptin-deficient mice ([Jo et al., 2005](#)), mirroring the changes in hypothalamic endocannabinoid content by leptin and leptin deficiency reported earlier ([Di Marzo et al., 2001b](#)). Another hypothalamic site where a leptin/endocannabinoid interaction may occur is the paraventricular nucleus. At this site, glucocorticoids have been shown to induce endocannabinoid synthesis and endocannabinoid-induced suppression of synaptic excitation via a cAMP-dependent mechanism, and leptin was found to block these effects by a phosphodiesterase 3B-mediated decrease in intracellular cAMP ([Malcher-Lopes et al., 2006](#)). These effects may underlie the orexigenic action of glucocorticoids.

Another recent study indicates the importance of lateral hypothalamic orexin neurons in reward-seeking behavior in general ([Harris et al., 2005](#)), suggesting that they may also be targets of the effects of endocannabinoids on drug reward (see section III. B.11.). Additionally, cannabinoids can increase the intake of palatable foods by acting at sites in the brainstem ([Miller et al., 2004](#)), which also have reciprocal neural connections with forebrain limbic structures ([Saper, 2002](#)). From a behavioral point of view, cannabinoids are involved in both the appetitive and consummatory aspects of feeding behavior ([Chaperon et al., 1998](#); [Thornton-Jones et al., 2005](#)), in line with their multiple sites of action in the brain. Such multiple sites of action are also indicated by findings that in THC-naïve rats, rimonabant suppressed food-maintained operant responses and metabolic activity in the limbic forebrain, measured by 2-deoxyglucose uptake, whereas in rats made tolerant to THC, an additional metabolic inhibition was detected in the hypothalamus ([Freedland et al., 2003](#)). Exposure of rats to a palatable diet containing sucrose and condensed milk resulted in down-regulation of CB<sub>1</sub> receptors in limbic structures involved in the hedonic aspects of feeding, but not in the hypothalamus ([Harrold et al., 2002](#)). In the hypothalamus, the very low density of CB<sub>1</sub> receptors is offset by their increased coupling ([Breivogel et al., 1997](#)), which may be an alternative target of regulation ([Basavarajappa and Hungund, 1999](#); [Wang et al., 2003](#)) that needs to be explored.

Within the appetitive neural circuitry, endocannabinoids have been shown to interact with both orexigenic factors such as endogenous opioids, NPY, orexins, and ghrelin, and anorexigenic factors

including  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH), corticotropin-releasing hormone (CRH), and the peptide product of the cocaine and amphetamine-related transcript (CART). Inhibition of food intake by opioid  $\mu$  receptor antagonists and CB<sub>1</sub> receptor antagonists is supra-additive ([Kirkham and Williams, 2001b](#); [Rowland et al., 2001](#); [Chen et al., 2004](#)), suggesting a synergism between the endogenous opioid and cannabinoid systems in mediating the reinforcing effect of food ([Solinas and Goldberg, 2005](#)). Indeed, CB<sub>1</sub>-deficient mice fail to self-administer morphine ([Ledent et al., 1999](#); [Cossu et al., 2001](#)) or to release dopamine in the nucleus accumbens in response to morphine ([Mascia et al., 1999](#)), suggesting that the site of this synergism is in the mesolimbic dopaminergic pathway, which is involved in both drug and food reward ([Le Foll and Goldberg, 2005](#)). The observation that SR141716 inhibits the orexigenic effect of morphine microinjected into the hypothalamic paraventricular nucleus but not the nucleus accumbens shell suggests additional interactions between the two systems, unrelated to the hedonic aspects of feeding ([Verty et al., 2003](#)). A further intriguing parallel between the two systems is that opiate  $\mu$  receptor knockout mice, just as CB<sub>1</sub><sup>-/-</sup> mice (see below), are resistant to diet-induced obesity ([Tabarin et al., 2005](#)).

As for interactions with NPY, the similar effectiveness of SR141716 to inhibit food intake in wild-type and NPY<sup>-/-</sup> mice indicates that endocannabinoids are unlikely to be the primary compensatory factor that accounts for the lack of a lean phenotype in NPY<sup>-/-</sup> mice ([Di Marzo et al., 2001b](#)). However, anandamide was found to increase and AM251 to decrease depolarization-induced NPY release in rat hypothalamic explants, suggesting that NPY may contribute to the orexigenic effects of cannabinoids ([Gamber et al., 2005](#)). A possible role of orexins in the appetitive effects of endocannabinoids is suggested by the finding that coexpression of the CB<sub>1</sub> and orexin 1 receptors results in a marked potentiation of orexin A-induced signaling ([Hilaret et al., 2003](#)). An important site of action of the orexigenic peptide ghrelin is the hypothalamic paraventricular nucleus, where its hyperphagic effect can be blocked by SR141716, suggesting that ghrelin may act via the release of endocannabinoids ([Tucci et al., 2004](#)). Endocannabinoids, in turn, may be involved in ghrelin release, at least in the periphery, as suggested by an SR141716-induced decrease in plasma ghrelin levels in rats ([Cani et al., 2004](#)).

The proopiomelanocortin-derived peptide  $\alpha$ -MSH acting at MC-4 melanocortin receptors is part of the leptin-regulated appetitive circuitry as a major anorectic mediator. The observations that SR141716 inhibits the feeding response induced by blocking MC-4 receptors, whereas  $\alpha$ -MSH does not affect THC-induced feeding, suggest that CB<sub>1</sub> receptors are downstream from MC-4 receptors and have an obligatory role in  $\alpha$ -MSH effects on food intake ([Verty et al., 2004](#)). The peptide product of CART is also a tonically active anorectic mediator ([Kristensen et al., 1998](#)) and, unlike  $\alpha$ -MSH, may be a downstream mediator of the effect of endocannabinoids. Such an arrangement is suggested by the finding that SR141716 loses its ability to reduce food intake in CART<sup>-/-</sup> mice ([Osei-Hyiaman et al., 2005a](#)). Furthermore, mice deficient in FAAH have reduced levels of CART immunoreactivity in various hypothalamic and extrahypothalamic regions involved in appetite control, which is returned to normal levels by chronic SR141716 treatment ([Osei-Hyiaman et al., 2005a](#)). These findings suggest that inhibition of CART release by CB<sub>1</sub> activation may be involved in the orexigenic effect of anandamide. Finally, an interaction between endocannabinoids and CRH is indirectly suggested by coexpression of the mRNA for the CB<sub>1</sub> receptor with the mRNA for CRH ([Cota et al., 2003](#)) or the CRH type 1 receptor ([Hermann and Lutz, 2005](#)).

**2. Endocannabinoids and Peripheral Energy Metabolism** It is generally accepted that energy intake and utilization are regulated in a coordinated fashion, and factors involved in the central regulation of appetite may also affect peripheral energy metabolism ([Seeley and Woods, 2003](#)). The first indirect indication that cannabinoids may affect energy homeostasis through a mechanism other than food intake came from a study of marijuana smokers tested in a hospital inpatient setting ([Greenberg et al., 1976](#)). In



this study, the marijuana-induced increase in caloric intake leveled off after a few days, whereas weight gain continued throughout the rest of the 21-day observation period, suggesting independent effects on appetite and peripheral energy metabolism. After the introduction of SR141716 as the first selective CB<sub>1</sub> receptor antagonist ([Rinaldi-Carmona et al., 1994](#)), a similar conclusion was reached in normal rats treated with SR141716 for 14 days. Tolerance to the anorectic effect of SR141716 developed within 5 days, whereas the reduction in body weight was maintained throughout the treatment period ([Colombo et al., 1998a](#)). Later, similar observations were reported in mice with diet-induced obesity, in which food intake was reduced transiently whereas the reduction in body weight was maintained when the animals were chronically treated with SR141716 ([Ravinet Trillou et al., 2003](#)) or AM251 ([Hildebrandt et al., 2003](#)). These results suggested that factors other than appetite must be involved in the weight-reducing effect of CB<sub>1</sub> antagonists.

Peripheral targets of endocannabinoids include adipocytes, which express CB<sub>1</sub> receptors ([Bensaid et al., 2003](#); [Cota et al., 2003](#)). Stimulation of CB<sub>1</sub> receptors on adipocytes can affect lipid metabolism through regulating the level of adiponectin production ([Bensaid et al., 2003](#)), by increasing lipoprotein lipase activity ([Cota et al., 2003](#)), or by inhibiting AMP-activated protein kinase (AMPK) ([Kola et al., 2005](#)), which leads to increased lipogenesis and decrease in fatty acid  $\beta$ -oxidation through reducing the phosphorylation and thus disinhibiting acetyl CoA carboxylase-1 (ACC1), the rate-limiting enzyme in fatty acid synthesis. The work by [Cota et al. \(2003\)](#) provided the first clear evidence of peripheral metabolic targets of endocannabinoids in vivo in a mouse model of diet-induced obesity. By careful analysis of body composition, they were able to establish the lean phenotype of CB<sub>1</sub>-deficient mice that had escaped earlier attention. Furthermore, the use of a pair-feeding paradigm revealed that hypophagia accounts for the lean phenotype only in young and not in adult animals, which clearly indicated the involvement of peripheral metabolic target(s) in the latter. The additional documentation of functional CB<sub>1</sub> receptors in primary cultured adipocytes and their role in regulating lipogenesis provided one of the likely peripheral targets for the anabolic effects of endocannabinoids. The lean phenotype of CB<sub>1</sub><sup>-/-</sup> mice in this study was more prominent in male than in female animals, which could suggest that endocannabinoid regulation of adiposity may be subject to modulation by sex hormones.

Although earlier studies failed to detect CB<sub>1</sub> receptors in the liver, more recently they have been identified in the mouse liver using a combination of methods including reverse transcription-polymerase chain reaction, in situ hybridization, immunohistochemistry, and Western blotting. In the same study, treatment of mice with the cannabinoid agonist HU-210 increased de novo lipogenesis and the expression of the transcription factor sterol regulatory element binding protein 1c (SREBP1c) as well as of its targets, ACC1 and fatty acid synthase ([Osei-Hyiaman et al., 2005b](#)). The role of CB<sub>1</sub> receptors in these effects was indicated by the ability of SR141716 to block them and by their absence in CB<sub>1</sub> knockout mice ([Osei-Hyiaman et al., 2005b](#)). The hepatic lipogenic pathway may be also directly activated through a cannabinoid-induced decrease in AMPK phosphorylation and activity in the liver ([Kola et al., 2005](#)). CB<sub>1</sub> receptors have been also detected in rat hepatocytes ([Michalopoulos et al., 2003](#)), in whole mouse liver ([Biecker et al., 2004](#)), and in rat and human hepatic stellate cells ([Siegmund et al., 2005](#); [Teixeira-Clerc et al., 2006](#)).

Fatty acid metabolism in hypothalamic neurons acts as a sensor of nutrient availability ([Obici et al., 2003](#)), and its pharmacological modulation influences food intake ([Kim et al., 2004](#)). CB<sub>1</sub> activation was reported to increase *SREBP1c* and *FAS* gene expression in the hypothalamus, and the increased expression of these genes by fasting/refeeding ([Paulaskis and Sul, 1988](#)) could be inhibited by SR141716 treatment at the beginning of the refeeding period, which also reduced food intake ([Osei-Hyiaman et al., 2005b](#)). Although fatty acid synthesis was not measured directly in the hypothalamus, these findings suggest that the increase in food intake after fasting may involve a CB<sub>1</sub>-mediated

modulation of the fatty acid synthetic pathway. Modulation of AMPK activity by cannabinoids was documented not only in liver and adipose tissue but also in hypothalamus (Kola et al., 2005), where it has been linked to appetite control (Minokoshi et al., 2004). Thus, the AMPK/ACC1/FAS pathway may represent a common molecular pathway involved in both the central appetitive and the peripheral metabolic effects of endocannabinoids.

Because total caloric intake is similar in wild-type and  $CB_1^{-/-}$  mice on a high-fat diet (Ravinet Trillou et al., 2004; Osei-Hyiaman et al., 2005b), the resistance of  $CB_1$ -deficient mice to diet-induced obesity must be associated with increased energy expenditure. Exposing wild-type C57BL6/J mice to a high-fat diet decreases energy expenditure, as documented by indirect calorimetry (Hu et al., 2004), which may account for the increase in feed efficiency observed in such animals, whereas in  $CB_1^{-/-}$  mice feed efficiency was unaffected by a high-fat diet (Osei-Hyiaman et al., 2005b). This suggests that the high-fat diet-induced decrease in energy expenditure is mediated by endocannabinoid activation of  $CB_1$  receptors. Accordingly, HU-210 treatment of wild-type mice decreased and SR141716 treatment increased the activity of carnitine palmitoyl transferase-1, the rate-limiting enzyme in fatty acid  $\beta$ -oxidation (D. Osei-Hyiaman and G. Kunos, unpublished observations).

One of the factors involved in this effect in vivo could be adiponectin, the adipocyte-derived hormone that promotes fatty acid  $\beta$ -oxidation (Yamauchi et al., 2002). Indeed, exposure to a high-fat diet resulted in a significant decline in plasma adiponectin in wild-type but not in  $CB_1^{-/-}$  mice (Osei-Hyiaman et al., 2005a), and  $CB_1$  receptor activation in isolated adipocytes was found to suppress adiponectin expression (Perwitz et al., 2005; Matias et al., 2006). Expression of the thermogenic uncoupling protein-1 was also down-regulated by  $CB_1$  activation, whereas the expression of the insulin-mimetic adipokine visfatin was increased (Perwitz et al., 2005). Conversely, rimonabant increases adiponectin secretion by adipocytes (Bensaid et al., 2003) and adiponectin plasma levels in obese human subjects (Després et al., 2005), which should lead to increased lipid  $\beta$ -oxidation and thermogenesis in vivo. Chronic treatment of *ob/ob* mice with SR141716 increased thermogenesis, as indicated by increased oxygen consumption at a thermoneutral temperature measured by whole body calorimetry (Liu et al., 2005). Glucose uptake, subsequently measured in the isolated soleus muscle of these animals, was significantly increased in the SR141716-pretreated group. A similar effect in humans may account for the increased glucose tolerance observed in obese patients treated with rimonabant (Van Gaal et al., 2005). These observations could suggest the presence of  $CB_1$  receptors in skeletal muscle, which was recently documented (Pagotto et al., 2006). Alternatively, increased glucose tolerance may be secondary to an effect of SR141716 on  $CB_1$  receptors in the liver. It has been proposed that increased lipid synthesis in the liver may produce insulin resistance in other tissues such as muscle (McGarry, 1992), and  $CB_1$  receptor activation has been shown to contribute to the development of hepatic steatosis in diet-induced obesity (Osei-Hyiaman et al., 2005b). Endocannabinoids may also influence insulin secretion directly in islet  $\beta$ -cells via  $CB_1$  (Matias et al., 2006) or  $CB_2$  receptors (Juan-Pico et al., 2005).

The ability of rimonabant to increase energy expenditure may not be limited to an effect on adiponectin secretion, as indicated by an analysis of the effect of rimonabant treatment on gene expression profiles in lean and diet-induced obese mice as well as  $CB_1^{-/-}$  mice (Jbilo et al., 2005). Rimonabant-induced decreases in body weight and adipose tissue mass in obese mice was accompanied by a near-complete reversal of obesity-induced changes in the expression of a wide range of genes. These included genes involved in adipocyte differentiation, lipolysis, generation of futile cycles, and glycolysis. These broad-based targets may underlie the ability of rimonabant to correct symptoms of the metabolic syndrome, as discussed below. They also raise the intriguing possibility that if a  $CB_1$  antagonist that does not cross the blood-brain barrier were available, it could be effective in the treatment of the metabolic syndrome

without the risk of adverse CNS side effects (Horvath, 2006).

**3. Obesity and Associated Metabolic Abnormalities** Genetic manipulation of the expression of endogenous proteins has been instrumental in uncovering their regulatory role in normal and pathological phenotypes. When CB<sub>1</sub> knockout mice were first introduced, no change in body mass or feeding pattern had been noted ([Ledent et al., 1999](#); [Zimmer et al., 1999](#)). However, in a subsequent study, CB<sub>1</sub> knockout mice were found to have a life-long, small, but significant, weight deficit compared with their wild-type littermates, which could be attributed to a selective deficit in adipose tissue mass ([Cota et al., 2003](#)) and was confirmed by others ([Ravinet Trillou et al., 2004](#); [Osei-Hyiaman et al., 2005b](#)). Parallel to their decreased fat mass, CB<sub>1</sub><sup>-/-</sup> mice have lower plasma leptin levels and an increased sensitivity to the anorectic effect of exogenous leptin ([Ravinet Trillou et al., 2004](#)).

The possibility that an increase in the activity of the endocannabinoid system may contribute to at least some forms of obesity was suggested by three sets of findings. First, CB<sub>1</sub> antagonists were significantly more efficacious in reducing caloric intake and body weight in rodents with diet-induced or genetic obesity than in their respective lean controls ([Di Marzo et al., 2001b](#); [Hildebrandt et al., 2003](#); [Ravinet Trillou et al., 2003](#); [Vickers et al., 2003](#)).

Second, CB<sub>1</sub><sup>-/-</sup> mice are resistant to diet-induced obesity ([Ravinet Trillou et al., 2004](#); [Osei-Hyiaman et al., 2005b](#)). In both of these studies, overall caloric intake was not different between wild-type compared with CB<sub>1</sub><sup>-/-</sup> mice receiving the high-fat diet, suggesting that peripheral mechanisms play a dominant role in the control of body weight by CB<sub>1</sub> receptors. CB<sub>1</sub><sup>-/-</sup> mice are also resistant to the metabolic changes that accompany diet-induced obesity in normal mice, including hypertriglyceridemia and elevated plasma leptin and insulin levels, indicative of leptin and insulin resistance, respectively ([Ravinet Trillou et al., 2004](#); [Osei-Hyiaman et al., 2005b](#)). These metabolic changes, collectively defined by some as the “metabolic syndrome”, could also be reversed by SR141716 treatment ([Ravinet Trillou et al., 2004](#); [Poirier et al., 2005](#)).

As a third line of evidence, recent findings indicate that endocannabinoids and CB<sub>1</sub> receptors are up-regulated in the liver and adipose tissue in various forms of experimental as well as in human obesity. In wild-type mice on a high-fat diet for 3 weeks, the basal rate of de novo hepatic fatty acid synthesis was markedly increased, and the increase was partially reversed by SR141716 treatment ([Osei-Hyiaman et al., 2005b](#)). After 3 weeks of diet, the mice were not yet overweight but showed significant hepatic steatosis. Their hepatic content of anandamide was increased 3-fold, and the level of CB<sub>1</sub> receptor protein in liver plasma membranes was also markedly increased ([Osei-Hyiaman et al., 2005b](#)). These findings indicate that intake of a high-fat diet activates the hepatic endocannabinoid system, which contributes to increased lipogenesis and the subsequent development of hepatic steatosis and, ultimately, the development of obesity. Exposure of C57BL/6/J mice to a high-fat diet has been reported to induce changes characteristic of the metabolic syndrome and also to rapidly induce the expression of SREBP1c and its downstream target lipogenic enzymes ([Biddinger et al., 2005](#)). CB<sub>1</sub> receptor knockout mice are resistant to these diet-induced changes, which indicates that endocannabinoids have a major role in mediating them ([Osei-Hyiaman et al., 2005b](#)).

An up-regulation of CB<sub>1</sub> receptors has been also reported in adipose tissue of genetically obese compared with lean mice ([Bensaid et al., 2003](#)), and elevated endocannabinoid levels have been detected in adipose tissue of obese compared with lean patients ([Matias et al., 2006](#)). In a study involving 40 women ([Engeli et al., 2005](#)), circulating levels of anandamide and 2-AG were significantly increased in 20 obese versus 20 lean subjects, and remained elevated after a 5% diet-induced weight reduction. Although these plasma levels were much too low to exert hormone-like activity, they probably originate from overflow from tissues and thus may reflect functionally relevant changes in

endocannabinoid content at or near sites of action. In the same study, FAAH expression was markedly reduced in the adipose tissue of obese subjects and correlated negatively with circulating endocannabinoid levels. Furthermore, the expression of both CB<sub>1</sub> and FAAH increased in mature adipocytes compared with preadipocytes. These findings suggest that the endocannabinoid system is activated in human obesity ([Engeli et al., 2005](#)).

A genetic missense polymorphism in the *FAAH* gene predicting a proline to threonine substitution at position 129, which was reported to result in reduced cellular expression and activity of the enzyme ([Chiang et al., 2004](#)), had been earlier found to be significantly associated with problem drug use ([Sipe et al., 2002](#)). The same polymorphism has been linked to overweight and obesity in both Caucasian and African-American subjects ([Sipe et al., 2005](#)). Interestingly, the elevated hepatic levels of anandamide in mice receiving a high-fat diet could be attributed to a decrease in FAAH activity ([Osei-Hyiaman et al., 2005b](#)), suggesting that FAAH may play a key role in regulating endocannabinoid “tone” in both experimental and human obesity. Although this finding could suggest the targeting of FAAH in the treatment of eating/metabolic disorders, such an approach will be complicated by the fact that oleylethanolamide, an anorectic lipid that acts on the peroxisome proliferator-activated receptor  $\alpha$  (PPAR $\alpha$ ) ([Fu et al., 2003](#)), is also a substrate for FAAH. The opposing effects of elevated levels of both anandamide and oleylethanolamide after pharmacological blockade of FAAH may therefore result in no net change in appetite and energy metabolism.

That increased endocannabinoid activity may also contribute to obesity and its metabolic consequences in humans was indicated by the highly promising results of recent clinical trials with rimonabant. As in the animal models of diet-induced obesity, rimonabant was effective both in reducing body weight and in reversing many of the associated metabolic abnormalities in obese subjects. In a multicenter, phase III study involving 1507 obese European subjects with a body mass index  $>30$  kg/m<sup>2</sup> or a body mass index  $>27$  kg/m<sup>2</sup> with dyslipidemia and moderate hypertension, rimonabant (20 mg/day) treatment for 1 year, combined with a moderately hypocaloric diet, not only reduced body weight but also reduced plasma triglycerides, increased HDL cholesterol, and decreased plasma insulin and insulin resistance ([Van Gaal et al., 2005](#)). Blood pressure was not significantly affected. The parallel reduction in body weight and waist circumference suggested that the weight loss was predominantly due to loss of visceral fat, which is known to be a predisposing factor for the metabolic syndrome. Rimonabant was well tolerated, with mild to moderate nausea, diarrhea, and mood disorders occurring slightly more in the treatment group than in the placebo group ([Van Gaal et al., 2005](#)).

Essentially similar findings were reported in another large-scale, phase III study (RIO-North America) involving 3045 randomized, obese or overweight subjects. At the end of the 1st year, rimonabant-treated subjects were re-randomized to receive rimonabant or placebo, whereas the placebo group continued onto receive the placebo. During the 2nd year, rimonabant-treated patients retained the improvements achieved during the 1st year, whereas those who switched to placebo regained their original weight ([Pi-Sunyer et al., 2006](#)).

In a third study (RIO-Lipids) involving 1036 overweight/obese subjects, 20 mg/day rimonabant taken for 1 year significantly reduced body weight ( $-6.3 \pm 0.5$  kg), weight circumference ( $-5.7 \pm 0.6$  cm), and plasma triglycerides ( $-12.4 \pm 3.2\%$ ), increased HDL cholesterol by  $8.1 \pm 1.5\%$  and increased LDL particle size, improved glucose tolerance, and significantly elevated plasma adiponectin levels, resulting in a 50% decrease in the prevalence of the metabolic syndrome in the study population ([Després et al., 2005](#)). In contrast with the other two studies, a statistically significant, small decrease in systolic and diastolic blood pressure was evident in the group receiving 20 mg of rimonabant, and the decrease was greater for patients with initial hypertension (blood pressure  $>140/90$  mm Hg). Although the reason for the lack of a blood pressure change in the other studies is not clear, the proportion of females was lower

in RIO-Lipids (~60%) than in the other two studies where they represented ~80% of subjects. It is possible that a modest reduction in blood pressure by rimonabant occurs preferentially in males. The cumulative finding that blood pressure reduction, if present, is less than expected based on a similar level of weight reduction alone ([Appel et al., 2003](#)), is noteworthy. As discussed in section D.1., rimonabant at an i.v. dose of 3 mg/kg causes a pressor response in anesthetized, hypertensive rats, which are supersensitive to the hypotensive effect of endogenous or exogenous anandamide ([Bátkai et al., 2004](#)). Although the pressor effect is much smaller at lower doses of rimonabant comparable with the 20-mg oral dose used in humans or in the absence of anesthesia (S. Bátka, P. Pacher, and G. Kunos, unpublished observations), careful monitoring of blood pressure, particularly in the early stages of rimonabant treatment, may be advisable. A polymorphism in the *FAAH* gene is associated with obesity ([Sipe et al., 2005](#)), and because of the reduced enzyme activity resulting from this polymorphism, some of the affected individuals may have an elevated endocannabinoid tone, reversal of which by rimonabant could increase blood pressure.

It is noteworthy that part of the rimonabant-induced improvements in the hormonal and lipid abnormalities in the three clinical studies appeared to be independent of weight reduction and, based on the preclinical findings discussed above, are most likely mediated via peripheral sites of action. An interesting alternative mechanism is suggested by the results of a recent meta-analysis of the effects of low carbohydrate, nonenergy-restricted diets on weight loss and cardiovascular risk factors ([Nordmann et al., 2006](#)). Such diets were found to lead to significant weight loss for up to 1 year. Surprisingly, they were more favorable than low-fat diets in reducing plasma triglycerides and increasing HDL cholesterol levels, without a favorable effect on total or LDL cholesterol. The pattern of these metabolic changes is similar to that of those caused by 20 mg of rimonabant in the three clinical trials. Rimonabant has been shown to preferentially suppress the preference for sweet compared with normal ([Simiand et al., 1998](#)) or high-fat reinforcers ([Ward and Dykstra, 2005](#)) and can cause longer lasting suppression of intake of sweet compared with normal food ([Gessa et al., 2006](#)). It is very possible that obese subjects treated with rimonabant unwittingly altered their diet by reducing carbohydrate intake, which may have contributed to the observed effects on triglycerides and HDL cholesterol. Detailed analyses of the effects of rimonabant on dietary habits are warranted.

Overall, the findings in these three large, multicenter clinical trials strongly support a pathogenic role of increased endocannabinoid activity in obesity and the associated metabolic abnormalities and highlight the unique therapeutic potential of CB<sub>1</sub> blockade. Additional benefits may be gained by combination therapies. The efficacy of statins to preferentially lower LDL cholesterol may be effectively complemented by the ability of rimonabant to increase HDL cholesterol. In the case of insulin, the ability of rimonabant ability to increase insulin sensitivity could reduce the dose requirement for insulin in obese diabetic subjects and could also counteract the tendency of insulin treatment to cause weight gain. Nevertheless, further large-scale studies are warranted in view of the high nonadherence rate observed in the three clinical trials to date, which may have resulted in overestimation of the benefits of treatment ([Simons-Morton et al., 2006](#)).

**4. Cachexia and Anorexia** A negative energy balance resulting from decreased appetite and food intake and increased energy expenditure, leading to weight loss, can be the consequence of wasting diseases such as AIDS or metastatic cancer, or it could be associated with aging, chemotherapy of cancer, or neuropsychiatric conditions such as anorexia nervosa or various forms of dementia including Alzheimer's disease. Although there is a growing body of evidence documenting the therapeutic effectiveness of synthetic THC or even smoked marijuana as appetite boosters in some of these conditions ([Regelson et al., 1976](#); [Gorter et al., 1992](#); [Nelson et al., 1994](#); [Beal et al., 1995, 1997](#); [Timpone et al., 1997](#)) ([Table 1](#)), there is only limited information on the potential involvement of the endocannabinoid system in their pathogenesis.

**TABLE 1**

Clinical trials with cannabinoid-related medications in human disease

[Open in a separate window](#)

DB, double blind; PL, placebo-controlled; EMG, electromyography; SB, single blind; C, crossover; OL, open-label; max, maximum; R, randomized; M, multicenter; VAS, visual analog scale; P, prospective; PL, placebo; FMF, familial Mediterranean fever; NIB, nitrogen analog of THC; NK<sub>3</sub> R, neurokinin 3 receptor; SR142801, (*R*)-(*N*)-[1-[3-[1-benzoyl-3-(3,4-dichlorophenyl)piperidin-3-yl]propyl]-4-phenylpiperidin-4-yl]-*N*-methylacetamide; SR48692, 2-[(1-(7-chloro-4-quinolinyl)-5-(2,6-dimethoxyphenyl)pyrazol-3-yl)carbonylamino]tricyclo(3.3.1.1)decan-2-carboxylic acid; SR463493B, 1(*Z*)-[2-(dimethylamino)ethoxyimino]-1(2-fluorophenyl)-3-(4-hydroxyphenyl)-2(*E*)-propene; 5-HT<sub>2A/2C</sub> R, serotonin 2A/2C receptor, WC, waist circumference; BP, blood pressure.

A few studies have reported the effectiveness of THC in stimulating appetite and weight gain in cancer patients, but these therapeutic effects have been more extensively documented in AIDS patients (reviewed by [Kirkham, 2004](#); [Martin and Wiley, 2004](#); [Hall et al., 2005](#)) (see also [Table 1](#)). Although concerns have been voiced about the potential immunosuppressive effect of cannabinoids in immunocompromised individuals ([Klein et al., 1998](#)), repeated THC administration in a randomized, prospective, controlled trial was found to have few if any consistent effects on various immune functions in AIDS patients receiving antiviral treatment ([Bredt et al., 2002](#)).

Anorexia may also be associated with normal aging. A number of hormonal factors have been implicated in the loss of appetite in the elderly, including growth hormone, cholecystokinin, leptin, and various cytokines ([Morley, 2001](#)). In a recent study in mice, an age-related decline in food and alcohol intake was accompanied by the loss of ability of the CB<sub>1</sub> antagonist SR141716 to reduce food and alcohol intake and a decrease in CB<sub>1</sub> receptor-stimulated GTP $\gamma$ S labeling in the limbic forebrain ([Wang et al., 2003](#)). These findings suggest that, at least in this animal model, an age-dependent decrease in CB<sub>1</sub> receptor signaling in the limbic forebrain may be related to the parallel decline in appetite for both food and alcohol. Anorexia can also accompany debilitating diseases such as Alzheimer's disease, in which the effectiveness of THC to stimulate appetite has been documented ([Volicer et al., 1997](#)). Anorexia nervosa is a psychiatric condition that occurs predominantly in younger women and is characterized by self-starvation, weight loss, and a disturbed body image. Plasma anandamide levels have been reported to increase in patients with restricting anorexia nervosa, which may be secondary to a marked decrease in plasma leptin levels in such patients ([Monteleone et al., 2002](#)). Although the relationship between brain and plasma levels of anandamide is not clear, a parallel increase in anandamide in brain regions involved in reward may mediate the rewarding effect of self-starvation in anorexic patients ([Monteleone et al., 2005](#)). A recent family-based study examined the possible association of a CB<sub>1</sub> receptor gene polymorphism consisting of differences in a trinucleotide repeat with anorexia nervosa. Although no difference was found between parental alleles transmitted or not transmitted to the affected siblings, preferential transmission of different alleles could be established when the patients were subdivided into restricting and bingeing/purging subgroups ([Siegfried et al., 2004](#)).

Endocannabinoids have been also implicated in a unique form of food intake: milk suckling in newborn animals. In an elegant series of studies, [Fride et al. \(2005\)](#) have proposed a role for 2-AG in the brain to stimulate the suckling response in mouse pups. In their model, endogenous 2-AG in the pup's brain initiated the suckling response via CB<sub>1</sub> receptors, with continued suckling depending on milk-derived 2-AG ([Fride, 2004](#)). As predicted by this model, treatment of pups with SR141716 inhibits suckling and leads to death due to failure to thrive, a condition analogous to a human condition known as nonorganic failure to thrive, in which an oral motor defect resulting in deficient suckling ([Reilly et al., 1999](#)) is similar to the condition in mouse produced by pharmacological blockade or genetic ablation of CB<sub>1</sub> ([Fride et al., 2005](#)). The relatively high dose of SR141716 to inhibit suckling and its residual effectiveness in CB<sub>1</sub> knockout mice suggested the additional involvement of a receptor distinct from CB<sub>1</sub> or CB<sub>2</sub> ([Fride et al., 2003](#)).

## B. Pain and Inflammation

One of the earliest uses of cannabis was to treat pain. Historical documents reveal the use of cannabis for surgical anesthesia in ancient China and to relieve pain of diverse origin in ancient Israel, Greece, Rome, and India (reviewed in [Mechoulam, 1986](#); [Iversen, 2000](#); [Mechoulam and Hanus, 2000](#)). Numerous early studies have also demonstrated beneficial effects of cannabinoids in animal models of pain (reviewed in [Walker and Huang, 2002](#); [Fox and Bevan, 2005](#)). In acute pain, anandamide, THC, cannabidiol, and synthetic cannabinoids such as CP55,940 and WIN 55,212-2 are effective against

chemical (Sofia et al., 1973; Formukong et al., 1988; Calignano et al., 1998; Costa et al., 2004a,b; Guindon et al., 2006; Ulugol et al., 2006), mechanical (Sofia et al., 1973; Martin et al., 1996; Smith et al., 1998; Guindon and Beaulieu, 2006), and thermal pain stimuli (Buxbaum, 1972; Bloom et al., 1977; Lichtman and Martin, 1991a,b; Fride and Mechoulam, 1993; Guindon and Beaulieu, 2006). Recent animal studies indicate that anandamide and cannabinoid ligands are also very effective against chronic pain of both neuropathic (Herzberg et al., 1997; Bridges et al., 2001; Fox et al., 2001; Guindon and Beaulieu, 2006) and inflammatory origin (Tsou et al., 1996; Richardson et al., 1998a,b,c; Li et al., 1999; Martin et al., 1999b; Guindon et al., 2006). Moreover, endocannabinoids and synthetic cannabinoids exert synergistic antinociceptive effects when combined with commonly used nonsteroid anti-inflammatory drugs, which may have utility in the pharmacotherapy of pain (Guindon and Beaulieu, 2006; Guindon et al., 2006; Ulugol et al., 2006). Interestingly, a recent study has implicated the endocannabinoid system in the analgesic activity of paracetamol (acetaminophen), the most widely used painkiller (Ottani et al., 2006), and there is also evidence for endocannabinoid involvement in the action of some general anesthetics, such as propofol (Patel et al., 2003; Schelling et al., 2006).

Cannabinoids exert their antinociceptive effects by complex mechanisms involving effects on the central nervous system (Martin et al., 1993; Hohmann et al., 1995, 1998, 1999; Martin et al., 1995, 1996, 1998, 1999a,b; Richardson et al., 1997, 1998a,b; Meng et al., 1998; Strangman et al., 1998; Hohmann and Walker, 1999; Fox et al., 2001), spinal cord (Yaksh, 1981; Lichtman and Martin, 1991a,b; Welch and Stevens, 1992; Richardson et al., 1997, 1998a,b; Hohmann et al., 1998; Chapman, 1999; Drew et al., 2000; Naderi et al., 2005; Suplita et al., 2006), and peripheral sensory nerves (Calignano et al., 1998; Richardson et al., 1998c; Hohmann and Herkenham, 1999; Fox et al., 2001; Johaneck et al., 2001; Kelly et al., 2003; Johaneck and Simone, 2004; Jordt et al., 2004; Amaya et al., 2006). This is consistent with the anatomical location of CB<sub>1</sub> receptors in areas relevant to pain in the brain, spinal dorsal horn, dorsal root ganglia, and peripheral afferent neurons (Herkenham et al., 1990, 1991a; Hohmann and Herkenham, 1998, 1999; Hohmann et al., 1999; SañudoPeña et al., 1999a).

In addition to the role of CB<sub>1</sub> receptors, there is recent evidence implicating CB<sub>2</sub> receptors in the antihyperalgesic activity of cannabinoids in models of acute and chronic, neuropathic pain, especially of inflammatory origin (Calignano et al., 1998; Hanus et al., 1999; Malan et al., 2001; Clayton et al., 2002; Ibrahim et al., 2003, 2005; Nackley et al., 2003a,b, 2004; Quartilho et al., 2003; Elmes et al., 2004; Hohmann et al., 2004; Scott et al., 2004; Whiteside et al., 2005; Ibrahim et al., 2006). Cannabinoid agonists may also release endogenous opioids, and a functional interplay between the endocannabinoid and opioid systems in modulating analgesic responses has been suggested by numerous studies (Pugh et al., 1997; Manzanares et al., 1999a,b; Houser et al., 2000; Ibrahim et al., 2005; Tham et al., 2005; Vigano et al., 2005a,b; Williams et al., 2006).

As discussed before, anandamide is also a ligand for TRPV<sub>1</sub> receptors, albeit with an affinity lower than its affinity for CB<sub>1</sub> receptors. The potential involvement of TRPV<sub>1</sub> in the analgesic effect of endogenous anandamide has been raised by the findings that the analgesic response to microinjection of a FAAH antagonist into the periaqueductal gray of rats could be inhibited by a similar local microinjection of 6 nmol of capsazepine (Maione et al., 2006). However, others reported that systemic administration of 10 mg/kg capsazepine, which blocked capsaicin-induced analgesia, failed to inhibit endocannabinoid-mediated, stress-induced analgesia, which could be enhanced by a FAAH inhibitor and completely blocked by the CB<sub>1</sub> antagonist rimonabant (Suplita et al., 2006).

The analgesic response to exogenous cannabinoids suggested a role for the endocannabinoid system in regulating pain sensitivity, which has received experimental support (reviewed in Walker et al., 2000, 2002; Cravatt et al., 2004; Boger et al., 2005). For example, Walker et al. (1999) have demonstrated increased anandamide levels in some brain areas involved in nociception after peripheral nociceptive



input in the rat. The functional role of endogenous anandamide was further supported by the predominantly CB<sub>1</sub>-mediated analgesic response to FAAH or endocannabinoid transport inhibitors in animal models of acute and chronic pain ([Lichtman et al., 2004a](#); [Chang et al., 2006](#); [Jayamanne et al., 2006](#); [La Rana et al., 2006](#); [Suplita et al., 2006](#)). Similarly, FAAH knockout mice had elevated brain levels of anandamide and displayed analgesic behavior in acute inflammatory, but not in chronic neuropathic models of pain ([Lichtman et al., 2004b](#)). Formation of anandamide and 2-AG is also increased in response to stress in the periaqueductal gray matter, in which inhibition of endocannabinoid degradation was found to enhance stress-induced analgesia in a CB<sub>1</sub> receptor-dependent manner ([Hohmann et al., 2005](#); [Suplita et al., 2006](#)), confirming and extending an earlier finding that implicated CB<sub>1</sub> receptors and endocannabinoids in stress-induced analgesia ([Valverde et al., 2000](#)).

In humans, the analgesic activity of THC and other cannabinoids is less clear-cut. There are numerous case reports on the beneficial effects of cannabis or synthetic derivatives of THC in pain associated with multiple sclerosis, cancer, neuropathies, and HIV infection ([Noyes et al., 1975a,b](#); [Martyn et al., 1995](#); [Consroe et al., 1997](#); [Hamann and di Vadi, 1999](#); [Ware et al., 2003](#); [Rudich et al., 2003](#); [Ware and Beaulieu, 2005](#); [Ware et al., 2005](#); [Berlach et al., 2006](#); reviewed in [Burns and Ineck, 2006](#)) ([Table 1](#)). The results of randomized studies conducted before 1999 on the analgesic effect of orally administered synthetic cannabinoids in patients with postoperative, post-traumatic, cancer, or spastic pain had been subjected to a meta-analysis. The authors concluded that cannabinoids were not more effective than codeine in controlling pain, and their use was associated with numerous undesirable, dose-limiting CNS side effects ([Campbell et al., 2001](#)).

Recent clinical trials with THC or cannabis extracts containing a 1:1 mixture of THC and cannabidiol (Sativex, GW-1000) have provided mixed results. In a randomized, double-blind, placebo-controlled crossover study of 48 patients suffering from central neuropathic pain due to brachial plexus avulsion, oromucosally administered THC or Sativex was ineffective in reducing the pain severity score recorded during the last 7 days of treatment ([Berman et al., 2004](#)). Similarly, oral THC (dronabinol) did not improve postoperative ([Buggy et al., 2003](#)) and neuropathic pain ([Attal et al., 2004](#)) in trials involving small numbers of patients. However, numerous lessons have been learned from these initial human studies on optimal trial design, dose and route of administration of cannabinoids, and more recent larger-scale studies allow reason for more optimism, as outlined below.

THC or Sativex reduced neuropathic pain in patients with traumatic nerve injury or multiple sclerosis in randomized, double-blind, placebo-controlled, crossover trials ([Wade et al., 2003](#); [Notcutt et al., 2004](#)). Modest, but clinically relevant analgesic effects were reported in 21 multiple sclerosis patients treated with dronabinol, in a randomized, controlled clinical trial ([Svendsen et al., 2004](#)). Effective pain relief by orally administered cannabis extract or THC was also reported in a randomized, controlled, multicenter trial involving 611 multiple sclerosis patients ([Zajicek et al., 2003](#)). Moreover, in a recent study of 66 multiple sclerosis patients, Sativex was effective in reducing central neuropathic pain ([Rog et al., 2005](#)). A preview of as-yet-unpublished human studies gave an account of a significant benefit of Sativex over placebo in peripheral neuropathic pain characterized by allodynia, in central pain associated with multiple sclerosis, and in opiate-resistant, intractable pain due to cancer ([Russo, 2006](#)). A multicenter dose-escalation study of the analgesic and adverse effects of an oral cannabis extract (Cannador) in patients with postoperative pain demonstrated significant dose-related improvements in rescue analgesia requirements and significant trends across the escalating dose groups for decreasing pain intensity ([Holdcroft et al. 2006](#)). THC (Marinol) was found to suppress otherwise intractable cholestatic pruritus in a case report ([Neff et al., 2002](#)). An analysis of pain questionnaires from 523 patients with HIV infections revealed that 90 to 94% of the subjects using cannabis experienced improvement in muscle and neuropathic pain ([Woolridge et al., 2005](#)). The therapeutic potential of cannabinoids in pain associated with trigeminal neuralgia and migraine has also been the subject of

several recent reviews ([Liang et al., 2004](#); [Russo, 2004, 2006](#)). Preclinical studies ([Burstein et al., 1998, 2004](#); [Burstein, 2000, 2005](#); [Dyson et al., 2005](#); [Mitchell et al., 2005](#); [Salim et al., 2005](#)) and a recent clinical trial of 24 patients with neuropathic pain of varying etiologies demonstrated that ajulemic acid, a major metabolite of THC with CB<sub>1</sub> agonist activity, was effective in reducing pain without causing cannabinoid-like CNS side effects, the first evidence for the separability of the psychotropic and analgesic effects of a THC analog in humans ([Karst et al., 2003](#)). Numerous additional human studies are ongoing to determine the effectiveness of THC or cannabis-based extracts against various forms of pain (reviewed in [Ware and Beaulieu, 2005](#) reviewed in Ware and Beaulieu, 2006).

Multiple lines of evidence support the important role of the cannabinoid signaling system in the modulation of immune function and inflammation (reviewed in [Klein et al., 1998, 2003](#); [Walter and Stella, 2004](#); [Klein, 2005](#)). First, cannabinoid receptors are present on immune cells, where their expression is modulated by microbial antigens or other stimuli that induce immune activation. Second, stimulation of immune cells by bacterial toxins such as lipopolysaccharide (LPS) increases the cellular levels of endocannabinoids and their degrading enzyme(s). Third, cannabinoid agonists modulate immune function both in vitro and in vivo via cannabinoid receptor-dependent and -independent mechanisms.

The anti-inflammatory effects of cannabinoids are complex and may involve modulation of cytokine (e.g., TNF- $\alpha$ , IL-12, IL-1, IL-6, and IL-10) and chemokine production (e.g., CCL2, CCL5, CXCL8, and CXCL10), modulation of adenosine signaling ([Carrier et al., 2006](#)), expression of adhesion molecules (e.g., ICAM-1, P- intercellular adhesion molecule-1 and P-selectin), and the migration, proliferation, and apoptosis of inflammatory cells (reviewed in [Klein et al., 1998, 2003](#); [Walter and Stella, 2004](#); [Klein, 2005](#)). To the extent that pain and inflammation accompany many of the disorders discussed in the rest of this review, cannabinoids would be expected to provide significant benefit due to their analgesic and anti-inflammatory properties.

### C. Central Nervous System Disorders

The emerging role of the endocannabinoid system in a variety of CNS disorders should not come as a surprise given the very high level of expression of CB<sub>1</sub> receptors in the brain. The particularly high density of CB<sub>1</sub> receptors in the cortex, cerebellum, hippocampus, and basal ganglia had drawn early attention to diseases affecting movement, mood and anxiety disorders, and conditions related to altered brain reward mechanisms, as well as processes of memory and learning. The classic behavioral effects of marijuana also provided early clues about potential therapeutic targets, such as the control of pain or appetite. The role of the endocannabinoid system in the pathogenesis and treatment of specific CNS diseases is discussed below.

**1. Neurotoxicity and Neurotrauma** The endocannabinoid system plays an important role in neuroprotection both in acute neuronal injury (e.g., traumatic brain injury, stroke, and epilepsy) and in chronic neurodegenerative disorders, such as multiple sclerosis, Parkinson's disease, Huntington's disease, amyotrophic lateral sclerosis, and Alzheimer's disease (reviewed in [Glass, 2001](#); [Mechoulam et al., 2002a,b](#); [Grundy, 2002](#); [Croxford, 2003](#); [Drysdale and Platt, 2003](#); [Jackson et al., 2005a](#); [Ramos et al., 2005](#)). Although the underlying mechanisms are not fully understood, multiple cannabinoid receptor-dependent as well as receptor-independent processes have been implicated. These include, but are not limited to 1) modulation of excitatory glutamatergic transmissions and synaptic plasticity via presynaptic CB<sub>1</sub> receptors ([Molina-Holgado et al., 1997](#); [Marsicano and Lutz, 1999](#); [Gerdeman et al., 2002](#); reviewed in [Alger, 2002](#); [Robbe et al., 2002](#); [Azad et al., 2003](#); [Freund et al., 2003](#); [Gerdeman and Lovinger, 2003](#); [Piomelli, 2003](#); [Mato et al., 2004](#)), 2) modulation of immune responses and the release of inflammatory mediators by CB<sub>1</sub>, CB<sub>2</sub>, and non CB<sub>1</sub>/CB<sub>2</sub> receptors on neurons, astrocytes, microglia,

macrophages, neutrophils and lymphocytes ([Watzl et al., 1991](#); [Zheng et al., 1992](#); [Fischer-Stenger et al., 1993](#); [Cabral and Fischer-Stenger, 1994](#); [Kusher et al., 1994](#); [Burnette-Curley and Cabral, 1995](#); [Cabral et al., 1995](#); reviewed in [Friedman et al., 1995](#); [Zheng and Specter, 1996](#); [Shohami et al., 1997](#); [Newton et al., 1998](#); [Srivastava et al., 1998](#); [Gallily et al., 2000](#); [Klein et al., 2000a,b, 2003](#); [Smith et al., 2000](#); [Carlisle et al., 2002](#); [Germain et al., 2002](#); [Killestein et al., 2003](#); [Kaplan et al., 2005](#); [Ramirez et al., 2005](#); reviewed in [Friedman et al., 1995](#); [Stella, 2004](#); [Walter and Stella, 2004](#); [Correa et al., 2005](#); [Croxford and Yamamura, 2005](#); [Klein, 2005](#)); 3) activation of cytoprotective signaling pathways ([Grigorenko et al., 2002](#)), such as protein kinase B/Akt ([Molina-Holgado et al., 2002](#)), protein kinase A ([Kim et al., 2005](#)), or neurotrophic factors ([Khaspekov et al., 2004](#)), 4) modulation of excitability and calcium homeostasis via effects on  $\text{Ca}^{2+}$ ,  $\text{K}^{+}$ , and  $\text{Na}^{+}$  channels, *N*-methyl D-aspartate (NMDA) receptors, gap junctions, and intracellular  $\text{Ca}^{2+}$  stores ([Caulfield and Brown, 1992](#); [Mackie and Hille, 1992](#); [Mackie et al., 1993](#); [Nadler et al., 1995](#); [Venance et al., 1995](#); [Shohami et al., 1997](#); [Hampson et al., 2000b](#); [Oz et al., 2000, 2004](#); [Chemin et al., 2001](#); [Maingret et al., 2001](#); [Nogueron et al., 2001](#); [Robbe et al., 2001](#); [Wilson and Nicoll, 2001](#); [Wilson et al., 2001](#); [Nicholson et al., 2003](#); [Guo and Ikeda, 2004](#); [del Carmen et al., 2005](#); [del Carmen Godino et al., 2005](#); [Zhuang et al., 2005](#)), 5) antioxidant properties of cannabinoids ([Eshhar et al., 1995](#); [Hampson et al., 1998](#); [Chen and Buck, 2000](#); reviewed in [Hampson et al., 2000a](#); [Marsicano et al., 2002a](#)), and 6)  $\text{CB}_1$  receptor-mediated hypothermia, possibly by reducing metabolic rate and oxygen demand ([Leker et al., 2003](#)).

Excitotoxicity, the toxic effects of an overactivation of glutamate receptors, and the resulting oxidative stress may contribute to the pathological processes eventually leading to cellular dysfunction or death in both acute and chronic forms of neurodegeneration ([Coyle and Puttfarcken, 1993](#); [McNamara, 1999](#); [Lutz, 2004](#)). Dexamabinol (HU-211), a behaviorally inactive cannabinoid and noncompetitive antagonist of NMDA receptors, protects primary rat neuronal cultures against NMDA and glutamate exposure in vitro ([Eshhar et al., 1993](#); [Nadler et al., 1993a,b](#)). THC protects primary cultured neurons against kainate-mediated toxicity in a  $\text{CB}_1$ -dependent manner ([Abood et al., 2001](#)), similar to protection by WIN 55,212-2 against low extracellular magnesium-induced cell death ([Shen and Thayer, 1998](#)). Palmitoylethanolamide also improves neuronal survival in a glutamate-induced cell death model ([Skaper et al., 1996](#)). Intracerebral injection of NMDA in neonatal rats results in a 13-fold increase of cortical anandamide concentrations ([Hansen et al., 2001a,b](#)). Both THC and anandamide exerted  $\text{CB}_1$ -mediated neuroprotective effects in an ouabain-induced rat model of in vivo excitotoxicity ([van de Stelt et al., 2001a,b](#)). Anandamide and synthetic agonists of  $\text{CB}_1$  receptors also protected the newborn brain against AMPA-kainate receptor-mediated excitotoxic damage in mice ([Shouman et al., 2006](#)).

Traumatic brain injury (TBI) is one of the leading causes of disability and mortality in young individuals ([Holm et al., 2005](#)), yet the available therapy is very limited ([Faden, 2002](#); [Maas et al., 2004](#)). TBI is characterized by cerebral edema, axonal and neuronal injury, increased permeability of the blood-brain barrier, and post-traumatic changes in cognitive and neurological functions ([Bayir et al., 2003](#)). TBI can trigger glutamate-induced excitotoxicity, oxidative stress, release of inflammatory cytokines from brain-resident cells (microglia, neurons, and astrocytes), programmed cell death, and cortical blood flow dysregulation (reviewed in [Wang and Feuerstein, 2000](#); [Gentleman et al., 2004](#)).

The protective effect of cannabinoids in traumatic brain injury was first indicated in studies with the nonpsychotropic cannabinoid dexamabinol (HU-211) ([Fig. 1b](#)). These studies have demonstrated reduced brain damage and improved motor and cognitive function in HU-211-treated animals in a rat model of TBI. The favorable effects of a single injection of HU-211 on learning and neurological deficits lasted up to 30 days and could be achieved within a therapeutic window of 6 h ([Shohami et al., 1993, 1995](#)). Beneficial effects of HU-211 were also demonstrated in an axonal injury model ([Yoles et al., 1996](#); [Zalish and Lavie, 2003](#)). These protective effects were attributed, at least in part, to NMDA receptor blockade, attenuation of  $\text{Ca}^{2+}$  influx and decreased TNF- $\alpha$  levels ([Nadler et al., 1995](#); [Shohami](#)

et al., 1997; reviewed in [Mechoulam et al., 2002a,b](#); [Biegon, 2004](#)). In mice with closed head injury, brain levels of 2-AG increased, and exogenous 2-AG administered 1 h after the head injury reduced infarct size and improved neurological outcome ([Panikashvili et al., 2001](#)). Neuroprotection by 2-AG was attributed to CB<sub>1</sub> receptor-mediated inhibition of nuclear factor- $\kappa$ B and of the early expression of proinflammatory cytokines TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 ([Panikashvili et al., 2005, 2006](#)). In a rat model of TBI, BAY 38-7271, a CB<sub>1</sub>/CB<sub>2</sub> agonist with predominant action at CB<sub>1</sub> receptors, caused a marked, 70% reduction in infarct volume when administered as a 4-h infusion immediately after induction of subdural hematoma, and even when it was applied with a 3-h delay, infarct volume was reduced by 59% ([Mauler et al., 2002](#)).

A multicenter, double-blind, randomized, placebo-controlled phase II trial conducted in 67 patients with severe closed head injury found dexanabinol to be safe and well tolerated. The treated patients achieved significantly better intracranial pressure/cerebral perfusion pressure control without jeopardizing blood pressure, and a trend toward faster improvement and better neurological outcome was also observed ([Knoller et al., 2002](#)). However, a double-blind, randomized, placebo-controlled phase III clinical trial of dexanabinol, conducted in 15 countries in 86 specialized centers and involving 861 patients failed to demonstrate any favorable effects ([Maas et al., 2006](#)).

**2. Stroke** Ischemic stroke is the most common form of stroke, mostly caused by a transient interruption of blood supply to the brain by thrombotic occlusion of blood vessels. It is an important cause of death and disability in industrialized countries, affecting up to 0.2% of the population each year ([Klijn and Hankey, 2003](#); [Pinto et al., 2004](#)). One in six patients die in the 1st month after ischemic stroke, and half of the survivors are permanently disabled despite the best efforts to rehabilitate them and to prevent complications ([Klijn and Hankey, 2003](#)).

One of the first indications of the neuroprotective effect of cannabinoids came from the field of stroke research, using various in vitro and in vivo models of ischemic injury. Anandamide, 2-AG, and WIN 55,212-2 protected cultured cortical neurons against hypoxia and glucose deprivation ([Nagayama et al., 1999](#); [Sinor et al., 2000](#)). The effects of various cannabinoid ligands were also investigated in in vivo models of global cerebral ischemia induced by two-vessel occlusion with hypotension or by four-vessel occlusion, or in focal ischemia induced by occlusion of the middle cerebral artery (MCAo), with or without reperfusion. Dexanabinol at doses of 2 to 10 mg/kg decreased infarct size and histological damage and improved neurological score in rat and gerbil models of both global and focal cerebral ischemia ([Bar-Joseph et al., 1994](#); [Vered et al., 1994 a,b](#); [Belayev et al., 1995a,b,c](#); [Leker et al., 1999](#); [Lavie et al., 2001](#); [Teichner et al., 2003](#)). Importantly, this protective effect was observed even when the drug was administered 60 to 180 min after the insult ([Vered et al., 1994](#); [Belayev et al., 1995a,b,c](#); [Leker et al., 1999](#); [Lavie et al., 2001](#); [Teichner et al., 2003](#)).

WIN 55,212-2, at doses of 0.03 and 1 mg/kg but not 3 mg/kg decreased hippocampal neuronal loss after transient global cerebral ischemia in rats. It also reduced infarct size after permanent focal cerebral ischemia induced by MCAo, when given 40 min before or 30 min after the occlusion, and these effects were prevented by SR141716 ([Nagayama et al., 1999](#)). WIN 55,212-2 also protected cultured cerebral cortical neurons from in vitro hypoxia and glucose deprivation, but in contrast to the receptor-mediated neuroprotection observed in vivo, this in vitro effect was not stereoselective and was insensitive to CB<sub>1</sub> and CB<sub>2</sub> receptor antagonists ([Nagayama et al., 1999](#)). BAY38-7271 also decreased infarct size in rats with permanent MCAo even when given intravenously 4 h after the occlusion ([Mauler et al., 2002](#)). Similarly, HU-210 reduced infarct size by up to 77% and improved motor disability in a rat model of permanent MCAo ([Leker et al., 2003](#)). The protective effect of HU-210 was partially reversed by pretreatment with SR141716, indicating CB<sub>1</sub> receptor involvement. Surprisingly, all protection could be abolished by warming the animals to the body temperature of controls, indicating that CB<sub>1</sub>-mediated

hypothermia contributed to the neuroprotection (Leker et al., 2003). Likewise, CB<sub>1</sub>-mediated hypothermia was responsible for the neuroprotective effects of THC in a mouse transient MCAo model (Hayakawa et al., 2004) and perhaps also in a rat model of global cerebral ischemia (Louw et al., 2000). Consistent with these findings, CB<sub>1</sub> knockout mice had increased mortality from permanent focal cerebral ischemia, increased infarct size, more severe neurological deficits after transient focal cerebral ischemia, and decreased cerebral blood flow in the ischemic penumbra during reperfusion, compared with wild type controls subjected to the same insult (Parmentier-Batteur et al., 2002). NMDA neurotoxicity was also increased in CB<sub>1</sub><sup>-/-</sup> mice compared with wild-type littermates (Parmentier-Batteur et al., 2002). Further evidence for a role of CB<sub>1</sub> receptors is their increased expression on neurons in the arterial boundary zone of cortical infarction (Jin et al., 2000). Finally, brain levels of endocannabinoids are increased during ischemic (Schmid et al., 1995; Schabitz et al., 2002; Berger et al., 2004; Muthian et al., 2004) and other types of brain injury (Sugiura et al., 2000; Hansen et al., 2001a,b; Panikashvili et al., 2001).

Other studies do not support the neuroprotective role of CB<sub>1</sub> receptor activation. For example, CB<sub>1</sub> antagonists by themselves had no effect on the outcome of injury, and in two recent reports, SR141716 and LY320135 were found to actually reduce infarct size and to improve neurological function in a rat model of MCAo (Berger et al., 2004; Muthian et al., 2004), whereas low doses of WIN 55,212-2 had no protective effect (Muthian et al., 2004). Thus, it appears that both CB<sub>1</sub> agonists and antagonists can be neuroprotective in cerebral ischemia. The reason for the opposite effects of pharmacological blockade versus genetic knockout of CB<sub>1</sub> receptors is not clear and may be related to the CB<sub>1</sub> receptor-independent effects of antagonists (Begg et al., 2005; Pertwee, 2005b,c). Clearly, evaluating the potential usefulness of cannabinoid ligands in the treatment of stroke warrants future studies.

**3. Multiple Sclerosis and Spinal Cord Injury** Multiple sclerosis (MS) is a complex, immune-mediated, inflammatory disease of the white matter of the brain, which compromises impulse conduction due to the loss of the myelin sheath of neurons and the secondary axonal loss (Sospedra and Martin, 2005). MS affects 2 to 5 million people worldwide and commonly presents with an unpredictable, relapsing-remitting course and a range of clinical symptoms depending on where the demyelination and axonal loss have occurred (Compston and Coles, 2002). Some patients become disabled within a short period of time, whereas others can live their entire lives with only negligible or no disability. The symptoms of MS typically involve tremor, ataxia, visual loss, double vision, weakness or paralysis, difficulty in speaking, loss of bladder control and constipation, cognitive impairment, and painful muscle spasms. Muscle spasticity often leads to reduced mobility, considerable distress from pain, and interference with daily living activities. Spasticity, neuropathic and nociceptive pain, and some of the above symptoms are also common in spinal cord injury (SCI). Although there are numerous drugs available that target the immune system to slow down the progression of the disease, they are only moderately effective, and the treatment of MS remains mostly symptomatic and far from satisfactory (Killestein and Polman, 2005).

Cannabis had been used in ancient Greece, Rome, China, and India for relieving muscle cramps, spasm, and pain (reviewed in Mechoulam, 1986; Mechoulam et al., 1998; Mechoulam and Hanus, 2000) and its therapeutic application in MS is a topic of recent lively debate (Grundy, 2002; Pertwee, 2002; Baker and Pryce, 2003; Croxford, 2003; Killestein et al., 2004; Sirven and Berg, 2004; Jackson et al., 2005a; Pryce and Baker, 2005; Robson, 2005; Smith, 2005). Lyman et al. (1989) examined the effects of parenteral THC in experimental autoimmune encephalomyelitis (EAE) in rats, a laboratory model of MS. THC treatment not only reduced CNS inflammation and improved neurological outcome but also improved survival compared with placebo. Δ<sup>8</sup>-THC, a less psychotropic and more stable analog of THC, also reduced the severity and incidence of neurological deficits in rats with EAE (Wirguin et al., 1994). The nonpsychotropic dexamabinol also suppressed inflammatory responses in the brain and spinal cord

of rats with EAE and improved their neurological symptoms ([Achiron et al., 2000](#)). Although CB<sub>1</sub> receptor density is decreased in the striatum and cortex of EAE rats, this is compensated for by increased coupling to G protein-mediated signaling, ensuring the effectiveness of treatment with cannabinoid agonists ([Berrendero et al., 2001](#)).

In a mouse model of chronic relapsing EAE, intravenous administration of THC, WIN 55,212-2, JWH-133, or methanandamide reduced spasticity and tremor, whereas the same symptoms were exacerbated by treatment with either CB<sub>1</sub> or CB<sub>2</sub> antagonists ([Baker et al., 2000](#)). These mice with EAE had increased levels of anandamide, 2-AG, and palmitoylethanolamide (PEA) in areas associated with nerve damage ([Baker et al., 2001](#)). Furthermore, spasticity could be relieved not only by administration of exogenous anandamide, 2-AG, or PEA but also by selective inhibitors of endocannabinoid transport or hydrolysis, which suggests tonic control of muscle tone by the endocannabinoid system in EAE ([Baker et al., 2001](#); [Ligresti et al., 2006a](#)). Additional evidence for this has emerged through the use of CB<sub>1</sub>-deficient mice, which tolerated inflammatory and excitotoxic insults poorly and developed substantial neurodegeneration after the induction of EAE ([Pryce et al., 2003](#)). [Jackson et al. \(2005b\)](#) reported that the absence of CB<sub>1</sub> receptors was associated with increased caspase activation and a greater loss of myelin and axonal/neuronal proteins after the induction of chronic EAE. Interestingly, CB<sub>1</sub> knockout mice had increased caspase 3 levels before the induction of EAE, suggesting a neuroprotective tone mediated by CB<sub>1</sub> receptors ([Jackson et al., 2005a,b](#)). In mice with EAE, WIN 55,212-2 inhibited leukocyte/endothelial interactions via activation of CB<sub>2</sub> receptors ([Ni et al., 2004](#)). Interestingly, a recent study suggests that the high levels of IFN- $\gamma$  present in the CNS of mice with EAE can counteract endocannabinoid-mediated neuroprotection by disrupting P2X7 purinergic receptor signaling, a key step in endocannabinoid production by glia ([Witting et al., 2006](#)).

Another murine model of MS is Theiler's murine encephalomyelitis virus-induced demyelinating disease. In mice with Theiler's murine encephalomyelitis virus-induced demyelinating disease, treatment with WIN 55,212-2 slowed the progression of symptoms, down-regulated delayed-type hypersensitivity reactions and interferon- $\gamma$  production, and inhibited the expression of proinflammatory cytokines in the CNS ([Croxford and Miller, 2003](#)). In another study using this model, treatment with WIN 55,212-2, ACEA, or JWH-015 caused long-lasting improvements in neurological deficits in the established disease and reduced microglial activation, abrogated major histocompatibility complex class II antigen expression, and decreased the number of CD4<sup>+</sup> infiltrating T cells in the spinal cord. These changes were paralleled by extensive remyelination ([Arevalo-Martin et al., 2003](#)). Treatment of Theiler's murine encephalomyelitis virus-infected mice with the transport inhibitors OMDM1 and OMDM2 enhanced anandamide levels, down-regulated inflammatory responses in the spinal cord, and ameliorated motor symptoms ([Mestre et al., 2005](#)), and similar findings were reported using the transport inhibitor UCM707 ([Ortega-Gutierrez et al., 2005](#)). In these two studies, the treatments were also shown to reduce the surface expression of major histo-compatibility complex class II molecules, the production of the proinflammatory cytokines (TNF $\alpha$ , IL-1 $\beta$ , and IL-6), and the expression of inducible NO synthase.

Consistent with the animal data, cannabinoids have shown promise in the treatment of MS in humans ([Table 1](#)). A possible underlying mechanism is suggested by a recent study in which the endocannabinoid system was found to be highly activated during CNS inflammation in MS patients and to protect neurons from inflammatory damage by activating a negative feedback loop in microglial cells via CB<sub>1/2</sub>-mediated epigenetic regulation of mitogen-activated protein kinase phosphatase 1 expression ([Eljaschewitsch et al., 2006](#)).

There have been anecdotal reports of the effectiveness of marijuana smoking in relieving symptoms of MS and SPI ([Grinspoon and Bakalar, 1993, 1998](#)), which were supported by the results of early open or single-blind observations with orally given THC or smoked marijuana, involving small numbers of

patients ([Dunn and Davis, 1974](#); [Petro, 1980](#); [Petro and Ellenberger, 1981](#); [Clifford, 1983](#); [Meinck et al., 1989](#); [Brenneisen et al., 1996](#); [Schon et al., 1999](#)). The most consistent finding was a subjective improvement in spasticity, although benefits for mobility, tremor, nystagmus, mood, and bladder control were also reported. In a double-blind crossover study of a single MS patient, nabilone treatment improved muscle spasms, nocturia, and general well-being ([Martyn, 1995](#)). In contrast, [Greenberg et al. \(1994\)](#) reported impairments of both balance and posture after a single dose of smoked cannabis in a placebo-controlled study of 10 MS patients and 10 normal subjects. In an anonymous survey of 112 MS patients who self-medicated with cannabis, 30 to 97% of the subjects reported relief from a wide variety of symptoms by smoking marijuana ([Consroe et al., 1997](#)). These encouraging reports have triggered numerous larger, population-based clinical trials of cannabis-based medicines in MS, which have yielded mixed results.

Using a randomized, double-blind, placebo-controlled, crossover design, [Killestein et al. \(2002\)](#) have evaluated the effects of oral THC, two doses of 2.5 to 5 mg/day or a *Cannabis sativa* plant extract administered over a 4-week period, in 16 MS patients with severe spasticity. Spasticity and disability, quantified using the objective Ashworth scale ([Ashworth, 1964](#)) and the Expanded Disability Status Scale were not improved. However, a significant improvement in the subjective rating of spasm frequency and trends toward improved mobility were noted, with no effect on tremor, sleep quality, or lower urinary tract symptoms. Both THC and the plant extract worsened the patients' global impression of their conditions, with plant extracts causing more adverse side effects. It should be mentioned, however, that the dose of THC used was lower than that in subsequent studies with more positive outcome, and as was noted in an accompanying editorial ([Thompson and Baker, 2002](#)), the study was not powered to detect efficacy.

A large multicenter study involving 33 clinical centers and 660 MS patients in the United Kingdom and United States and supported by the UK Medical Council aimed to explore the effects of cannabis extract (Cannador) or synthetic THC (Marinol) versus placebo on spasticity, pain, tremor, bladder function, and cognitive function [Cannabinoids in Multiple Sclerosis (CAMS) study; [Zajicek et al., 2003, 2004](#)]. There was no change in Ashworth score, tremor, irritability, depression, or tiredness after 15 weeks of treatment with Marinol or Cannador. However, there were significant improvements in patient-reported spasticity, pain, and sleep quality. Unexpectedly, there was also a reduction in hospital admissions for relapse in the two active treatment groups. Adverse side effects were generally minor and similar to those with placebo. Remarkably, in the 12-month follow-up of the original CAMS study of 657 patients, muscle spasticity measured by the Ashworth scale was significantly improved in the THC-treated group. The Rivermead Mobility Index was also improved, indicative of reduced disability. The effect of Cannador on tremor was also studied in a randomized, double-blind, placebo-controlled, crossover trial in 14 patients with MS. Consistent with an earlier report ([Zajicek et al., 2003](#)), no significant therapeutic effects were noted ([Fox et al., 2004](#)). In another study of similar design, administration of oral capsules containing 2.5 mg of  $\Delta^9$ -THC plus 0.9 mg of CBD (maximal dose of 30 mg of  $\Delta^9$ -THC/day) caused improvements in spasm frequency and mobility in 37 MS patients who received at least 90% of their prescribed dose ([Vaney et al., 2004](#)).

In a double-blind, placebo-controlled study involving 18 patients with MS, THC and CBD decreased self-reported spasticity and pain and improved bladder symptoms, whereas spasticity measured by the Ashworth scale was not significantly improved ([Wade et al., 2003](#)). The therapeutic effect of Sativex delivered by oromucosal spray (2.7 mg of THC and 2.5 mg of CBD at each actuation) was evaluated in 160 outpatients with MS ([Smith, 2004](#)). Patients were allowed to self-titrate the dose to achieve optimal effects, up to a maximal daily dose of 120 mg of THC and CBD. Efficacy was assessed by using a modified Ashworth scale to assess spasticity, whereas daily living, mobility, cognitive function, and tremor were quantified through the use of various scales and questionnaires ([Wade et al., 2004](#)). There

was no significant difference in the Ashworth scale, tremor, and pain at 6 weeks between the Sativex and placebo groups. However, visual analog scales showed significant improvement in patients whose primary symptom had been spasticity ([Wade et al., 2004](#)). Sativex was well tolerated and effective against central neuropathic pain and sleep disturbances associated with MS in a randomized, controlled trial involving 66 patients ([Rog et al., 2005](#)). Sativex was approved and launched in Canada in 2005 for the treatment of neuropathic pain associated with MS and is currently being investigated for various other therapeutic indications ([Russo, 2004, 2006](#)).

In a recent case report, a 46-year-old woman was diagnosed with MS after having entered treatment with the CB<sub>1</sub> receptor antagonist rimonabant for obesity, and recovery to near normal was noted within weeks after discontinuation of the treatment ([van Oosten et al., 2004](#)). This report, coupled with the more severe neurodegenerative process when MS is induced in CB<sub>1</sub> knock-out mice or in mice treated with a CB<sub>1</sub> receptor antagonist, could suggest that CB<sub>1</sub> antagonism may exacerbate inflammatory demyelinating diseases in humans ([van Oosten et al., 2004](#)). However, the occurrence of MS in this one patient may have been purely coincidental.

Although the results of the above clinical studies ([Table 1](#)) are somewhat equivocal, patients treated with cannabis experienced improvements in the most disturbing symptoms including pain and spasticity compared with those receiving placebo, without experiencing significant side effects. These studies also suggest that the Ashworth scale as a primary measure of spasticity in MS does not accurately assess the complex collection of symptoms associated with spasticity, which may be more accurately evaluated using subjective measures. Indeed, the use of the Ashworth scale as a primary measure of spasticity in MS has often been criticized, and many commonly used antispasticity medications have also failed to generate statistically significant improvements according to this scale ([Hinderer and Gupta, 1996](#); [Shakespeare et al., 2003](#)). Accurate assessment of the clinical effectiveness of cannabinoids in MS may be complicated by the difficulty in achieving the most appropriate individual oral dose ([Table 1](#)). Peak plasma concentrations and their timing vary greatly because of the low water solubility of cannabis components and the large variability in their absorption from the gastrointestinal tract. An additional disadvantage of oral administration is the hepatic first-pass effect. This can result in the formation of large quantities of the psychoactive metabolite 11-OH-THC, which may be responsible for some of the side effects observed ([Table 1](#)). Delivery of cannabis-based extracts as an oromucosal spray may minimize these drawbacks and may allow patients to better optimize their individual daily dose by self-titration ([Russo, 2006](#)).

In conclusion, controlled clinical trials with cannabinoids have demonstrated their efficacy in eliciting symptomatic improvements in MS patients. These results suggest that there is place for the use of cannabis in the treatment of MS, which should be confirmed in further larger-scale clinical trials.

**4. Movement Disorders (Basal Ganglia Disorders)** Endocannabinoid involvement in the central regulation of motor functions and in movement disorders is based on multiple lines of evidence. First, CB<sub>1</sub> receptors are highly expressed in the basal ganglia, especially in the substantia nigra and in the cerebellum ([Herkenham et al., 1990, 1991a,b](#); [Mailleux and Vanderhaeghen, 1992](#); [Tsou et al., 1998](#); [Hohmann and Herkenham, 2000](#); [Moldrich and Wenger, 2000](#); [Howlett et al., 2002](#)), areas involved in motor control. Second, endocannabinoids are also abundant in these brain regions ([Bisogno et al., 1999a](#); [Di Marzo et al., 2000](#)). Third, endogenous, plant-derived, and synthetic cannabinoids have potent, mostly inhibitory, effects on motor activity ([Crawley et al., 1993](#); [Fride and Mechoulam, 1993](#); [Wickens and Pertwee, 1993](#); [Smith et al., 1994](#); [Romero et al., 1995a,b, 2002b](#); reviewed in [Sañudo-Peña et al., 1999b](#)). Fourth, CB<sub>1</sub> receptor and endocannabinoid levels are altered in the basal ganglia both in experimental models ([Zeng et al., 1999](#); [Page et al., 2000](#); [Romero et al., 2000](#); [Lastres-Becker et al., 2001a,b, 2002a,b](#); [Gonzalez et al., 2006](#)) and human forms of movement disorders ([Glass et al., 1993](#),



2000, 2004; Lastres-Becker et al., 2001a; reviewed in Romero et al., 2002b). Fifth, the endocannabinoid system interacts with several neurotransmitter pathways at various levels of the basal ganglia circuitry (Glass et al., 1997a; Miller et al., 1998; Sañudo-Peña and Walker, 1998; Giuffrida et al., 1999; Rodriguez De Fonseca et al., 2001; Brotchie, 2003; van der Stelt and Di Marzo, 2003; de Lago et al., 2004a).

*a. Parkinson's disease and levodopa-induced dyskinesia* Parkinson's disease (PD) is the second most common neurodegenerative disease of adult onset, with incidence of 16 to 19/100,000 people worldwide (Twelves et al., 2003). PD is caused by a severe loss of dopaminergic neurons in the substantia nigra pars reticulata (SNr), resulting in reduced dopamine levels and a loss of dopaminergic neurotransmission in the striatum, which interferes with motor function and coordination. Although excitotoxicity, oxidative stress, inflammation, mitochondrial dysfunction, and environmental and hereditary factors have all been implicated in the pathogenesis of PD, the exact cause of the loss of dopaminergic neurons remains elusive (Hattori and Mizuno, 2004; Eriksen et al., 2005). Clinically, PD is characterized by the classic triad of resting tremor, muscular rigidity, and bradykinesia/akinesia (slowness of movement or postural immobility). Current therapies include oral dopamine replacement via the dopamine precursor levodopa, anti-cholinergic agents, and monoamine oxidase B inhibitors (Horn and Stern, 2004). Although dopamine replacement therapy can be effective in most patients by controlling the symptoms in the short term, their long-term use is associated with diminishing efficacy and severe side effects such as levodopa-induced dyskinesia (LID) (involuntary movements), which often lead to treatment discontinuation and severe disability.

In PD, there are secondary abnormalities in nondopaminergic transmission within the basal ganglia that are thought to contribute to the inhibition of motor function. Inhibitory GABAergic transmission from the striatum to the external region of the globus pallidus (GPe) is increased, making the GPe hypoactive. This results in decreased GABAergic input from the GPe to the subthalamic nucleus which, together with increased activity of glutamatergic efferents to this nucleus, results in its hyperactivity. In turn, the hyperactive subthalamic nucleus increases the activity of the SNr and internal globus pallidus (GPi) through glutamatergic efferents. Because both the SNr and GPi provide inhibitory output to motor nuclei outside the basal ganglia (e.g., motor thalamus and brain stem locomotor regions), this mechanism is thought to contribute to the excessive motor inhibition in PD (Obeso et al., 2000; Bezard et al., 2001). In general, changes opposite to those described above are likely to be involved in LID. The final outcomes of the dysregulation of neuronal circuits are abnormal patterning, firing rate, and synchronization of basal ganglia outputs (Obeso et al., 2000; Bezard et al., 2001). Importantly, nondopaminergic mechanisms may counterbalance the loss of dopamine and are probably responsible for the lack of parkinsonian symptoms until the loss of >80% of striatal dopamine. They may also attenuate the severity of symptoms once symptoms develop. As discussed below, the endocannabinoid system may play an important regulatory role in PD, PD and LID as well as in the compensatory mechanisms.

Overactivity of endocannabinoid transmission, as reflected by increased tissue levels of endocannabinoids and CB<sub>1</sub> receptors as well as decreased rates of endocannabinoid transport and degradation by FAAH, have been found in the basal ganglia in the 6-hydroxydopamine-lesioned or reserpine-treated rat models of PD (Mailleux and Vanderhaeghen, 1993; Romero et al., 2000; Gubellini et al., 2002; Centonze et al., 2005; Fernandez-Espejo et al., 2005; Gonzalez et al., 2006). In basal ganglia from 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-lesioned marmosets, a primate model of PD, and in basal ganglia of PD patients, the density of striatal CB<sub>1</sub> receptors and CB<sub>1</sub> receptor-G-protein coupling were found to be increased (Lastres-Becker et al., 2001a). The above changes in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-treated marmosets and 6-hydroxydopamine-lesioned rats were reversible by chronic L-dopa treatment, which indicates that the similar changes observed in PD

patients were unlikely to have been induced by the replacement therapy ([Lastres-Becker et al., 2001a](#); [Maccarrone et al., 2003](#)). There is broad agreement that the endocannabinoid system becomes overactive in the basal ganglia in PD (reviewed in [Brotchie, 2003](#)), although some studies report a reduction ([Silverdale et al., 2001](#)) or no change in CB<sub>1</sub> receptor expression ([Herkenham et al., 1991a](#)) or a dependence on L-DOPA cotreatment of the increased CB<sub>1</sub> receptor expression in the basal ganglia of animals with experimental PD ([Zeng et al., 1999](#)).

If the enhanced CB<sub>1</sub> receptor signaling in the striatum is viewed as an attempt of the dopamine-deficient brain to normalize striatal function, the pharmacological amplification of this signaling might alleviate symptoms of PD, e.g., by reducing striatal glutamate release ([Gerdeman and Lovinger, 2001](#); [Gerdeman et al., 2002](#); [Gubellini et al., 2002](#)). On the other hand, enhanced CB<sub>1</sub> receptor signaling, if focused on the other part of the circuitry (e.g., GPe), can enhance GABA transmission, leading to inhibition of GPe and thereby contributing to the symptoms of PD. Likewise, CB<sub>1</sub> antagonism could have either pro-parkinsonian effects, if it targets the striatum, or antiparkinsonian effects, if it targets the GPe. Accordingly, both agonists and antagonists might have therapeutic potential, both in PD and LID (reviewed in [Brotchie, 2003](#)).

Treatment with CB<sub>1</sub> receptor agonists can decrease the tremor associated with overactivity of the subthalamic nucleus ([Sañudo-Peña et al., 1998, 1999b](#)), improve motor impairment seen with dopaminergic agonists ([Anderson et al., 1995](#); [Maneuf et al., 1997](#); [Sañudo-Peña et al., 1998](#)), protect against dopaminergic cell death ([Lastres-Becker et al., 2005](#)), and delay or reduce the incidence of LID ([Sieradzan et al., 2001](#); [Fox et al., 2002a](#); [Ferrer et al., 2003](#); [Gilgun-Sherkiet et al., 2003](#)). However, cannabinoid agonists are unlikely to be used for reducing bradykinesia in PD because of their hypokinetic profile both in primates and humans ([Consroe, 1998](#); [Müller-Vahl et al., 1999a](#); [Romero et al., 2002](#); [Brotchie, 2003](#); [Croxford, 2003](#); [Croxford and Miller, 2003](#)).

On the other hand, dysfunction of nigrostriatal dopaminergic neurons can be associated with overactivity of endocannabinoid transmission in the basal ganglia (see above). CB<sub>1</sub> receptor antagonists may therefore be useful for alleviating the bradykinesia of PD or LID, because they attenuate CB<sub>1</sub> signaling in GPe or GPi. ([Mailleux and Vanderhaeghen, 1993](#); [Di Marzo et al., 2000](#); [Lastres-Becker et al., 2001a,b](#); [Gubellini et al., 2002](#); reviewed in [Brotchie, 2003](#); [Fernandez-Espejo et al., reviewed in Brotchie, 2003; 2005](#); [Fernandez-Ruiz and Gonzalez, 2005](#)). Notwithstanding the above, studies using SR141716 in rat ([Di Marzo et al., 2000](#)) and primate models of PD or LID ([Meschler et al., 2001](#); [van der Stelt et al., 2005](#)) provided conflicting results. Rimonabant treatment also failed to influence dyskinesia in the first small-scale, randomized, double-blind, placebo-controlled human study ([Mesnage et al., 2004](#)). However, the dose used in this human study was approximately 10-fold lower (0.3 mg/kg versus 3 mg/kg), than in a recent primate study with positive outcome ([van der Stelt et al., 2005](#)). As suggested by a recent report ([Fernandez-Espejo et al., 2005](#)), it is also possible that CB<sub>1</sub> receptor blockade is effective only at the very advanced stages of the disease. More recently, using Park-2 knockout mice, a genetic model of early PD, [Gonzalez et al. \(2005\)](#) observed gender-dependent differences for both the levels of CB<sub>1</sub> receptors and motor responses to agonists or antagonists, extending earlier data obtained in humans and in animal models of PD.

Taken together, although the above studies do not offer a complete understanding of the role of endocannabinoids and cannabinoid receptors in PD and LID, they support the notion that the endocannabinoid system plays an important role in movement disorders, including PD, and may provide the framework for novel therapeutic approaches in the future.

**b. Huntington's disease** Huntington's disease (HD) is an inherited, autosomal dominant, progressive neuropsychiatric disorder of the midlife, caused by an unstable expansion of a trinucleotide

polyglutamine repeat in the N-terminal domain of a protein termed huntingtin, which leads to degeneration of neurons in the basal ganglia and cortical regions. The disease is characterized by motor disturbances, such as chorea (involuntary movements) and dystonia, psychiatric symptoms, and dementia ([Melone et al., 2005](#)). The prevalence of HD is similar to that of ALS (see below), but much lower than that of most of the other neurodegenerative illnesses discussed above or below. The therapy of HD is very limited and includes antidopaminergic drugs to reduce the hyperkinesias and antiglutamatergic agents to reduce excitotoxicity ([Melone et al., 2005](#)).

It has been clearly demonstrated, both in postmortem human tissue ([Glass et al., 1993, 2000](#); [Richfield and Herkenham, 1994](#)) and in chemically induced and transgenic animal models ([Denovan-Wright and Robertson, 2000](#); [Page et al., 2000](#); [Lastres-Becker et al., 2001, 2002a,b](#); [Sieradzan and Mann, 2001](#); [Behrens et al., 2002](#); [Glass et al., 2004](#); [McCaw et al., 2004](#)) that a decrease in CB<sub>1</sub> receptor level and signaling activity in the basal ganglia is one of the earliest changes in HD, preceding nerve loss and clinical symptoms. Furthermore, decreased levels of anandamide and 2-AG in the striatum and an increase of anandamide in the ventral mesencephalon, where the substantia nigra is located, have been documented in a rat model of HD ([Lastres-Becker et al., 2001](#)). Thus, it appears that endocannabinoid signaling in the basal ganglia is hypofunctional in HD, which probably contributes to the hyperkinesia associated with the disease. These studies also suggest that the endocannabinoid system is involved in the pathogenesis and/or progression of HD, and cannabinoid agonists could be of significant therapeutic benefit in HD because of their antihyperkinetic and neuroprotective effects (reviewed in [Lastres-Becker et al., 2003b](#)). A recent study identified a novel population of progenitor cells expressing CB<sub>1</sub> receptors in the subependymal layer of the normal and Huntington's diseased human brain. This finding raises the intriguing possibility that these cells could be a source of replacement of cells lost due to neurodegenerative disease ([Curtis et al., 2006](#)).

Indeed, data from animal models demonstrated that both CB<sub>1</sub> agonists and inhibitors of endocannabinoid transport are able to reduce hyperkinesia ([Lastres-Becker et al., 2002b, 2003a](#)). Interestingly, direct agonists of CB<sub>1</sub> receptors, such as CP55,940, only produced a very modest effect compared with the anandamide transport inhibitor AM404, which also exhibits affinity for the VR<sub>1</sub> receptor ([Zygmunt et al., 2000](#)). This latter property of AM404 may account for its ability to reduce hyperkinesia ([Lastres-Becker et al., 2002b, 2003a](#)), as other transport inhibitors such as VDM11 and AM374, which are not active at TRPV<sub>1</sub> receptors, were devoid of antihyperkinetic effects in HD rats ([Lastres-Becker et al., 2003a](#)), and the most potent transport inhibitor to date, UCM707, only produced a modest effect ([de Lago et al., 2002, 2004b, 2006](#)). Arvanil, a hybrid endocannabinoid and vanilloid compound, was also reported to alleviate hyperkinesias in a rat model of HD ([de Lago et al., 2005](#)). These results suggest that TRPV<sub>1</sub> receptors alone, or in combination with CB<sub>1</sub> receptors, might represent novel therapeutic targets in HD (reviewed in [Lastres-Becker et al., 2003b](#)).

There have been few human trials on the effects of cannabinoid agonists in HD, and the results do not live up to the promise of the animal data. Small trials with the synthetic THC analog nabilone and with the nonpsychoactive cannabidiol showed no efficacy or even increased choreic movements in HD patients ([Consroe et al., 1991](#); [Müller-Vahl et al., 1999b](#)). These negative results could be related to dosing issues, to the lack of TRPV<sub>1</sub> receptor activity of the compounds tested, or to the advanced stage of the disease. Nevertheless, further studies are warranted to explore the therapeutic potential of cannabinoids in HD.

*c. Gilles de la Tourette's syndrome, tardive dyskinesia, and dystonia* Based on its ubiquitous presence in motor regions of the brain, the endocannabinoid system might be involved in other extrapyramidal disorders such as Gilles de la Tourette's syndrome (TS), tardive dyskinesia, and dystonia. TS is a neurological syndrome that becomes evident in early childhood and is characterized by multiple motor

and vocal tics lasting for more than 1 year. Plant-derived cannabinoids have been found to be effective in the treatment of tics and behavioral problems in TS (Müller-Vahl et al., 1997, 1998, 1999c, 2002, 2003a, Müller-Vahl et al., b; Müller-Vahl, 2003). Beneficial effects of cannabinoids have been also reported in dystonia, both in animal models (Richter and Löscher, 1994, 2002) and in humans (Fox et al., 2002b; Jabusch et al., 2004). In addition, as described in the sections above, cannabinoids have potential in the management of the LID in PD and of the spasticity and tremor in MS. On the other hand, in patients chronically treated with neuroleptic drugs, a correlation between chronic cannabis use and the presence of tardive dyskinesia has been described previously (Zaretsky et al., 1993).

**5. Amyotrophic Lateral Sclerosis** ALS (also known as Lou Gehrig's disease) is the most common adult-onset human motor neuron disease with a prevalence of 5 to 7/100,000. It is characterized by rapid, progressive degeneration of motor neurons in the brain and spinal cord, which ultimately leads to progressive weakness, paralysis, and premature death (Rowland and Shneider, 2001). Although weak, patients are cognitively intact and thus are completely aware of their progressive disability. The disease strikes adults at any age, and most patients die within 3 to 5 years after the onset of symptoms. Although most cases of ALS are sporadic and are probably acquired, approximately 10% are familial, usually inherited in an autosomal dominant pattern. Despite a variety of putative underlying mechanisms, including oxidative stress, neuroinflammation, autoimmunity, a defect in neuronal glutamate transport and glutamate toxicity, neurofilament accumulation, exogenous factors (viruses or toxins), mitochondrial dysfunction, and mutations in the superoxide dismutase (*SOD1*) gene, the pathogenesis of ALS is incompletely understood (Barnham et al., 2004). Tragically, available treatment options are limited and do not prevent disease progression and death (Rowland and Shneider, 2001).

Based on the well-known protective effect of cannabinoids against oxidative cell damage and excitotoxicity (Hampson et al., 1998; Shen and Thayer, 1999; Abood et al., 2001; van der Stelt et al., 2001a), combined with their antispastic effect in MS, Carter and Rosen (2001) have proposed the use of marijuana for the pharmacological management of ALS. Indeed, in a pilot study of the safety and tolerability of THC in ALS patients, symptomatic benefits were seen for spasticity, insomnia, and appetite (Gelinas et al., 2002). Consistent with this clinical report, studies using transgenic mice expressing a mutant form of human SOD1 (hSOD1<sup>G93A</sup> mice) as an experimental model of ALS have demonstrated that either THC or WIN55,212-2 administered after the onset of the disease or genetic ablation of FAAH delayed disease progression (Raman et al., 2004; Bilslund et al., 2006). Furthermore, THC potently reduced oxidative and excitotoxic damage in spinal cord cultures in vitro and prolonged survival in SOD1 mutant mice (Raman et al., 2004). Surprisingly, neither WIN55,212-2 nor FAAH ablation affected the life span of SOD1(G93A) mice, whereas deletion of the CB<sub>1</sub> receptor significantly extended life span without affecting the disease onset (Bilslund et al., 2006). These results suggest that cannabinoids have significant neuroprotective effects in a mouse model of ALS but that these beneficial effects may be mediated by non-CB<sub>1</sub> receptor mechanisms.

**6. Alzheimer's Disease** Alzheimer's disease (AD) is a progressive neurodegenerative disorder that accounts for the vast majority of age-related dementia and is one of the most serious health problems in the industrialized world. The disease is characterized by the formation of neuritic plaques rich in  $\beta$ -amyloid ( $A\beta$ ) peptide, neurofibrillary tangles rich in hyperphosphorylated  $\tau$  protein, gliosis, and a neuroinflammatory response involving astrocytes and microglia, inevitably leading to progressive global cognitive decline (Weksler et al., 2005). These studies have engendered new perspectives on the possible role of the endocannabinoid system in neurodegenerative processes associated with inflammation (reviewed in Walter and Stella, 2004), including those in AD (reviewed in Pazos et al., 2004).

In an in vitro cell culture model of AD, anandamide prevented  $A\beta$ -induced neurotoxicity through CB<sub>1</sub>-

mediated activation of the mitogen-activated protein kinase pathway (Milton, 2002). In rat microglia cells in culture, CB<sub>1</sub> receptor stimulation also dose dependently inhibited the release of NO, which had been implicated in the neurotoxic effects of A $\beta$  peptide (Waksman et al., 1999). In PC12 cells, protection against A $\beta$ -induced neurotoxicity was also observed with cannabidiol, which does not bind to CB<sub>1</sub>/CB<sub>2</sub> receptors (Iuvone et al., 2004). Interestingly, CB<sub>1</sub> receptor blockade by SR141716 improved the memory deficit induced by administration of A $\beta$  peptide in mice, presumably by increasing hippocampal acetylcholine levels (Mazzola et al., 2003). However, analyses of brain tissue samples obtained from AD patients (Westlake et al., 1994) or animal models of AD (Romero et al., 1998; Benito et al., 2003) indicate that CB<sub>1</sub> receptors are not dramatically affected. In contrast, CB<sub>2</sub> receptors and FAAH are overexpressed in microglia associated with neuritic plaques in the brain of AD patients (Benito et al., 2003). Senile plaques in AD patients express both CB<sub>1</sub> and CB<sub>2</sub> receptors together with markers of microglial activation, and CB<sub>1</sub>-positive neurons, present in high numbers in control cases, are greatly reduced in areas of microglial activation (Ramirez et al., 2005). CB<sub>1</sub> receptor protein levels and G protein coupling were also markedly decreased in AD brains, coupled with increased nitration of the CB<sub>1</sub> and CB<sub>2</sub> receptor proteins (Ramirez et al., 2005). Intracerebroventricular administration of WIN 55,212-2 to rats prevented A $\beta$ -induced microglial activation, cognitive impairment and loss of neuronal markers. HU-210, WIN 55,212-2, and JWH-133 blocked A $\beta$ -induced activation of cultured microglial cells, as judged by mitochondrial activity, cell morphology and TNF- $\alpha$  release, and these effects were independent of the antioxidant action of ligands. Furthermore, cannabinoids abrogated microglia-mediated neurotoxicity after addition of A $\beta$  to rat cortical cocultures (Ramirez et al., 2005). Although there are no data available on the endocannabinoid content in AD brain tissue are available, increased levels have been reported in the brain after inflammatory events and in neurodegenerative disorders associated with inflammation (reviewed in Walter and Stella, 2004 and see also sections above).

Based on the above, one might hypothesize that A $\beta$  deposition induces the release of endocannabinoids from neurons and glia, which activate CB<sub>1</sub>-mediated neuroprotective pathways and modulate the release of inflammatory mediators in microglia through CB<sub>2</sub> receptors. If this hypothesis is confirmed by future studies, the beneficial effects of CB<sub>1</sub>/CB<sub>2</sub> agonists and FAAH antagonists in AD could be explored. Intriguingly, in a recent open-label pilot study of six patients in the late stages of dementia (five patients with AD and one patient with vascular dementia), treatment with 2.5 mg of dronabinol daily for 2 weeks significantly improved the Neuropsychiatric Inventory total score and the subscores for agitation and aberrant motor and nighttime behaviors (Walther et al., 2006).

**7. Epilepsy** If the balance between inhibitory and excitatory communications among neurons is disturbed, the intensity of excitatory transmission may exceed a certain threshold, leading to epileptic seizures. Stimulation of postsynaptic neurons is known to trigger the on-demand synthesis of endocannabinoids via an increase in intracellular calcium and/or stimulation of metabotropic receptors (reviewed in Piomelli, 2003; Lutz, 2004). Thereafter, endocannabinoids are released and reach presynaptic CB<sub>1</sub> receptors retrogradely to modulate both inhibitory GABAergic and excitatory glutamatergic transmissions via multiple mechanisms (Marsicano and Lutz, 1999; Alger, 2002, 2004; Gerdeman et al., 2002; Robbe et al., 2002; Azad et al., 2003; Freund et al., 2003; Gerdeman and Lovinger, 2003; Kim and Alger, 2004; Isokawa and Alger, 2005).

Multiple pathways, eventually culminating in neuronal death, are triggered by excessive excitatory activity through a process known as excitotoxicity (McNamara, 1999). Excitotoxicity is believed to contribute to the progression of numerous degenerative central nervous system disorders such as Parkinson's disease, Alzheimer's disease, and various forms of epilepsy. More than 1% of the human population is affected by epilepsy and the incidence is highest in elderly persons or during the first years

of life (reviewed in [Holmes and Ben-Ari, 1998](#); [McCormick and Contreras, 2001](#)). Epileptic syndromes are classified as generalized seizures, which affect the entire forebrain, or partial seizures, which occur within localized brain regions. Conventional antiepileptic treatment is not fully effective in ~30% of patients, therefore justifying the search for new targets ([LaRoche and Helmers, 2004](#)).

Cannabis has been used to treat epilepsy for centuries. Hashish was reported to cure the sick son of the chamberlain of the Caliphate Council in Baghdad by the medieval Arab writer Ibn al-Badri ([Mechoulam, 1986](#); [Iversen, 2000](#)). Almost four centuries later, W. B. O'Shaughnessy, an Irish physician and scientist working at the Medical College of Calcutta, confirmed the benefit of hashish for treating pain, emesis, muscle spasms, and convulsions (reviewed in [Karler and Turkanis, 1981](#); [Mechoulam, 1986](#)). The benefit of cannabis in epilepsy was also reported by a British neurologist ([Reynolds, 1890](#)), but the medicinal use of cannabis was prohibited in the early 20th century in most countries.

After the identification of the structure of THC ([Gaoni and Mechoulam, 1964](#)), several groups investigated its antiepileptic effects (reviewed in [Gordon and Devinsky, 2001](#); [Lutz, 2004](#)). THC was originally characterized as an anticonvulsant, but it has a variety of excitatory and depressant effects, ranging from convulsions to ataxia, depending on the dose, experimental model, and the animal species used ([Karler and Turkanis, 1981](#); reviewed in [Gordon and Devinsky, 2001](#); [Lutz, 2004](#)). Further complicating the picture, animal studies also document a rebound effect to THC with enhanced CNS excitability and increased sensitivity to convulsions ([Chiu et al., 1979](#); [Karler and Turkanis, 1981](#); [Karler et al., 1986](#)). This withdrawal hypersensitivity implies that in susceptible patients, the use of marijuana may be associated with withdrawal seizures ([Karler and Turkanis, 1981](#)).

Only case reports on the effects of THC in epileptic patients are currently available. Two reports described decreased seizure frequency after marijuana use ([Consroe et al., 1975](#); [Ellison et al., 1990](#)) and an epidemiological study found that chronic marijuana use is protective against seizures ([Ng et al., 1990](#)). According to a questionnaire completed by 215 epileptic patients using marijuana regularly, 7.4% experienced a reduction, 2.3% an increase, and 90.2% no change in seizure frequency ([Gordon and Devinsky, 2001](#)). In contrast, marijuana smoking was associated with an increase in seizure frequency in another study ([Keeler and Reifler, 1967](#)). Small-scale clinical studies have shown that the nonpsychotropic cannabidiol either reduced seizure frequency or had no significant effect on it ([Cunha et al., 1980](#); [Ames and Cridland, 1986](#); [Gordon and Devinsky, 2001](#)).

As in human studies, cannabinoids were found to exert both anti- and proconvulsive activity in animal models of epilepsy, largely depending on the stimulus applied to induce seizures (chemical, electrical, light, or fever) and the species used ([Johnson et al., 1975](#); [Ten Ham et al., 1975](#); [Wada et al., 1975a,b](#); [Corcoran et al., 1978](#); [Chiu et al., 1979](#); [Duncan and Dagirmanjian, 1979](#); [Fish et al., 1981](#); [Karler and Turkanis, 1981](#); [Colasanti et al., 1982](#); [Fish and Consroe, 1983](#); [Karler et al., 1984, 1986](#); [Consroe and Mechoulam, 1987](#); [Pertwee et al., 1991](#); [Hayase et al., 2001a,b](#); reviewed in [Gordon and Devinsky, 2001](#); [Lutz, 2004](#)).

Anandamide and its metabolically stable analog, O-1812, dose dependently inhibited electroshock-induced seizures in rats, and this effect was abolished by SR141716 ([Wallace et al., 2002](#)). In a rat model of febrile seizures, the expression of presynaptic CB<sub>1</sub> receptors in hippocampal GABAergic interneurons was increased ([Chen et al., 2003](#)), and the CB<sub>1</sub> receptor-mediated DSI was enhanced ([Alger, 2002](#)), suggesting that the endogenous cannabinoid system is protective. Remarkably, in a rat model of pilocarpine-induced status epilepticus, CB<sub>1</sub> receptor agonists were more effective in reducing seizure frequency than clinically used anticonvulsants, such as phenytoin or phenobarbital. Consistently, CB<sub>1</sub> receptor blockade increased seizure frequency, and the seizure activity was associated with increased brain levels of CB<sub>1</sub> receptors and 2-AG ([Wallace et al., 2003a](#)).

With use of the kainic acid-induced excitotoxic epileptiform seizure model in wild type and CB<sub>1</sub> knockout mice, recent studies have established that the seizure-induced increase of intracellular calcium, a hallmark of epilepsy ([Raza et al., 2001](#)), triggers the synthesis of anandamide, which activates CB<sub>1</sub> receptors in glutamatergic neurons in the hippocampus and cerebral cortex ([Marsicano et al., 2003](#); [Khaspekov et al., 2004](#)). Such “on-demand” activation of CB<sub>1</sub> receptors was suggested to protect against excitotoxicity by various mechanisms, including inhibition of calcium channels and stimulation of potassium channels to decrease neuronal excitability and the activation of extracellular signal regulated kinases ([Marsicano et al., 2003](#); [Khaspekov et al., 2004](#)). In contrast to these findings, FAAH knockout mice or mice treated with a CB<sub>1</sub> agonist were found to have increased sensitivity to kainic acid-induced seizures ([Clement et al., 2003](#)). The lack of protection in this latter study may be related to the nonselective activation of CB<sub>1</sub> receptors on both inhibitory (proconvulsive effect) and excitatory neurons (anticonvulsive effect) and by the life-long rather than on-demand activation of CB<sub>1</sub> receptors present in FAAH knockout animals.

In summary, the use of cannabinoids for the treatment of epilepsy is still controversial, although recent experimental studies provide some new insight. To date, there have been no large-scale, controlled clinical trials to examine the beneficial effects of cannabinoids in various forms of epilepsy. The potential use of the nonpsychotropic cannabidiol and of inhibitors of anandamide transport or degradation warrants further investigation.

**8. Mental Disorders** The well-known psychotropic effects of cannabinoids and the distribution of cannabinoid receptors across important emotional circuits in the brain suggest that the endocannabinoid system may be involved in various psychiatric disorders such as schizophrenia and mood disorders (reviewed in [van der Stelt and Di Marzo, 2003](#); [Hall et al., 2004](#); [Leweke et al., 2004](#); [Manzanas et al., 2004](#); [Ujike and Morita, 2004](#); [Ashton et al., 2005](#); [Gambi et al., 2005](#); [Semple et al., 2005](#); [Vinod and Hungund, 2005](#)).

*a. Schizophrenia* Schizophrenia is the second most common mental disorder with a lifetime prevalence of approximately 0.2 to 2% worldwide ([Ban, 2004](#)). The disease usually begins in early adulthood or late adolescence and is characterized by psychotic episodes with positive symptoms including delusions and/or hallucinations, loose associations, and distortion of perception. The psychotic episodes are separated by periods with negative symptoms consisting of apathy, anhedonia, reduced social drive, loss of motivation, poverty of speech and thought, and blunting of affect. With disease progression, behavioral impairment can lead to complete social isolation. Although recent advances in the pharmacotherapy of schizophrenia produced great improvement in the clinical symptoms and the quality of life of patients, there is room for further improvements ([Ban et al., 2004](#); [Moller, 2005](#)).

Numerous theories have been put forth regarding the etiology of schizophrenia, ranging from developmental or neurodegenerative processes, environmental factors, neurotransmitter abnormalities (dopamine or glutamate), and infectious or autoimmune processes, but also including the cannabinoid hypothesis (reviewed in [Thaker and Carpenter, 2001](#); [Lewis et al., 2005](#)). It appears that hypoglutamatergic and hypodopaminergic transmission in the prefrontal cortex is involved in the negative symptoms, whereas hyperactivity of dopamine neurotransmission in the mesencephalic projections to the nucleus accumbens may underlie the positive symptoms ([Thaker and Carpenter, 2001](#); [Lewis et al., 2005](#)).

According to the endocannabinoid hypothesis of schizophrenia, overactivity of the endocannabinoid system may lead to a hyperdopaminergic and hypoglutamatergic state, which may underlie some of the symptoms ([Emrich et al., 1997](#), reviewed in [Ujike and Morita, 2004](#); [Laviolette and Grace, 2006](#)). The endocannabinoid hypothesis is supported by multiple lines of evidence. First, the use of large amounts of cannabis and THC may produce psychotic symptoms in normal individuals, including delusions,

hallucinations, and cognitive impairment, which resemble schizophrenia ([Spencer, 1971](#); [Halikas et al., 1972](#); [Chopra and Smith, 1974](#); [McGuire et al., 1994](#); [Emrich et al., 1997](#); [Johns, 2001](#); [D'Souza et al., 2004](#)). Second, cannabis and THC may worsen psychotic symptoms in schizophrenic patients, contribute to poor outcome, increase the possibility of relapse, and decrease the effectiveness of antipsychotic drugs ([Breakey et al., 1974](#); [Treffert, 1978](#); [Negrete, 1989](#); [Turner and Tsuang, 1990](#); [Linszen et al., 1994](#); [Martinez-Arevalo et al., 1994](#); [Voruganti et al., 2001](#); [D'Souza et al., 2005](#)). Third, the use of cannabis may precipitate the onset of schizophrenia in individuals susceptible to psychosis ([Andreasson et al., 1987](#); [Miller et al., 2001](#)). Fourth, postmortem radioligand studies document increased CB<sub>1</sub> receptor density in the dorsolateral and anterior cingular regions and subregions of the prefrontal cortex in schizophrenia ([Dean et al., 2001](#); [Zavitsanou et al., 2004](#); [Newell et al., 2006](#)). Fifth, the levels of anandamide are increased in cerebrospinal fluid or blood from schizophrenic patients ([Leweke et al., 1999](#); [De Marchi et al., 2003](#); [Giuffrida et al., 2004](#)). Sixth, treatment with neuroleptics appears to normalize the imbalance in endocannabinoid signaling in blood cells in schizophrenic patients ([De Marchi et al., 2003](#)) and also decreases CB<sub>1</sub> receptor binding in the rat nucleus accumbens ([Sundram et al., 2005](#)). Last, the hebephrenic type of schizophrenia shows a strong association with polymorphisms in the *CNRI* gene encoding CB<sub>1</sub> receptors ([Leroy et al., 2001](#); [Ujike et al., 2002](#)).

Taken together, the above evidence suggests that the endocannabinoid system may be a novel therapeutic target in schizophrenia. It is also tempting to speculate that CB<sub>1</sub> antagonists may be beneficial against some, most likely the negative, symptoms of the disease. Some preclinical and clinical evidence also suggests that cannabidiol may have antipsychotic potential (reviewed in [Zuardi et al., 2006](#)).

*b. Anxiety and depression* Mood disorders such as generalized anxiety or panic disorder, major depressive disorder and bipolar disorder (manic depressive illness) are very common, often serious, and potentially life-threatening conditions. More than 20% of the adult population experiences a mood disorder at some point during their life. In up to 15% of individuals with major depressive disorder the cause of death is suicide. According to a World Health Organization forecast, by the year 2020 depression will become the second leading cause of premature death and disability worldwide ([Pacher and Kecskeméti, 2004](#)). Although significant advances have been made in the treatment of mood disorders during the past decades, ~30% of the population do not respond to current therapies, and the search for novel pharmacological approaches continues (reviewed in [Pacher and Kecskeméti, 2004](#)).

Many of the psychological effects of cannabis and THC are biphasic and bidirectional, depending on mode of administration, dose, personality, time frame, degree of tolerance, and various other environmental and individual factors ([Paton and Pertwee, 1973](#); [Ashton et al., 1981, 2005](#); [Viveros et al., 2005](#)). The acute effects in normal subjects can range from euphoria, relaxation, excitation, heightened perception, and increased motor activity to dysphoria, anxiety, sedation, perceptual distortion, and incoordination. THC, under certain conditions and at certain doses, exerts anxiolytic, antidepressant, and hypnotic effects in patients suffering from pain associated with cancer or multiple sclerosis and improves mood and general well-being in normal subjects ([Regelson et al., 1976](#); [Glass et al., 1980](#); [Ashton et al., 1981](#); [Fabre and McLendon, 1981](#); [Ilaria et al., 1981](#); [Martyn et al., 1995](#); [Ashton, 1999](#); [Wade et al., 2003](#)). However, under different conditions and at higher doses, cannabis or THC can produce dysphoric reactions, anxiety, panic paranoia, and psychosis ([Spencer, 1971](#); [Halikas et al., 1972](#); [Chopra and Smith, 1974](#); [Ashton et al., 1981, 2005](#); [McGuire et al., 1994](#); [Emrich et al., 1997](#); [Johns, 2001](#); [Patton et al., 2002](#); [Tournier et al., 2003](#); [Dannon et al., 2004](#); [D'Souza et al., 2004](#); reviewed in [Hollister, 1986](#); [Hall and Solowij, 1998](#)).

CBD also possesses anxiolytic, antipsychotic and anticonvulsant properties, which are not mediated by classic cannabinoid receptors ([Carlini et al., 1975](#); [Consroe and Wolkin, 1977](#); [Consroe et al., 1981](#);



Zuardi et al., 1982, 1995, 2006; Ames and Cridland, 1986; Martin et al., 1987; Guimaraes et al., 1990, 1994; reviewed in Mechoulam et al., 2002c; Grotenhermen, 2003; Long et al., 2006). The mode of action of CBD is not completely understood; it may involve blockade of anandamide and serotonin reuptake (Bisogno et al., 2001; McPartland and Russo, 2001), inhibition of the enzymatic hydrolysis of anandamide (Mechoulam et al., 2002), or an interaction with as yet unidentified receptors (Járai et al., 1999; Pertwee et al., 2002).

Animal studies yielded further support to the biphasic and bidirectional effects of cannabinoids on anxiety, with low doses being anxiolytic and high doses being anxiogenic. Indeed, low doses of CP55,940 (Genn et al., 2003; Marco et al., 2004), nabilone (Onaivi et al., 1990), and THC (Berrendero and Maldonado, 2002) exerted anxiolytic-like effects in the light-dark crossing test and in the elevated plus-maze in adult rodents. Low-dose CP55,940 was also anxiolytic in other models of anxiety in adult, juvenile, or infant rodents (Romero et al., 2002a; Borcel et al., 2004; Genn et al., 2004). In contrast, at medium to high doses, CP55,940 or HU-210 displayed anxiogenic effects in the same or other experimental paradigms in adult as well as in juvenile or infant animals (McGregor et al., 1996a,b; Rodriguez de Fonseca et al., 1996; Giuliani et al., 2000; Arevalo et al., 2001; Marin et al., 2002; Romero et al., 2002; Genn et al., 2003;2003;2004; Marin Marco et al., 2004). Although several hypotheses have been proposed to explain the biphasic effects of cannabinoids on anxiety, including distinct receptors (Haller et al., 2004a,b) or neuroanatomically separated CB<sub>1</sub> receptors with a differential sensitivity to the anxiolytic versus anxiogenic effects of cannabinoids, these need to be confirmed in future studies (reviewed in Viveros et al., 2005).

The high level of CB<sub>1</sub> receptors in the hippocampus, amygdala, and prefrontal and anterior cingulate cortex, key regions in the regulation of anxiety, may suggest that the endocannabinoid system plays a role in the control of anxiety (Herkenham et al., 1990, 1991a,b; Glass et al., 1997b; Katona et al., 2001; Hájos and Freund, 2002; Tzavara et al., 2003; Pistis et al., 2004). Further support of this theory came from studies using CB<sub>1</sub> receptor antagonists or CB<sub>1</sub> receptor knockout mice. SR141716 produced anxiogenic effects in the elevated plus-maze and the defensive withdrawal tests in adult rats (Navarro et al., 1997; Arevalo et al., 2001). Furthermore, SR141716 not only reversed the anxiolytic effects of the CB<sub>1</sub> agonist CP55,940 but also was anxiogenic in the ultrasonic vocalization test in rat pups when administered alone (McGregor et al., 1996a). In contrast, Haller et al. (2002) found SR141716 to be anxiolytic in the plus-maze in mice, but this effect was not mediated by CB<sub>1</sub> receptors as indicated by its presence in CB<sub>1</sub> knockout mice. Furthermore, another selective CB<sub>1</sub> receptor antagonist, AM251, increased anxiety-like behavior in wild-type mice but had no effect in the knockouts, in support of a CB<sub>1</sub> receptor-mediated anxiolysis. As discussed before, SR141716, but not AM251, also inhibits a CB<sub>1</sub>-like receptor that mediates presynaptic inhibition of glutamate release in the hippocampus (Hájos and Freund, 2002). Thus, the findings of Haller et al. (2002) could suggest that the anxiolytic effect of SR141716 is mediated by such a CB<sub>1</sub>-like receptor, activation of which would be anxiogenic.

CB<sub>1</sub> knockout mice displayed increased anxiogenic responses in the light-dark box, plus-maze, and social interaction tests, an increased aggressive response in the resident-intruder test, and marked alterations in the hypothalamic-pituitary-adrenal (HPA) axis coupled with impaired action of known anxiolytic drugs such as buspiron and bromazepam (Haller et al., 2002, 2004b; Martin et al., 2002; Urigüen et al., 2004). However, Marsicano et al. (2002) were unable to demonstrate anxiogenic-like response in CB<sub>1</sub> knockout mice in the plus-maze. This may be related to differences in the genetic background of the CB<sub>1</sub> knockout mice used and/or different experimental conditions. The importance of the latter is also indicated by the confounding effect of stress on anxiogenic behaviors and their modulation by endocannabinoids (Haller et al., 2004a; Patel et al., 2005). Stress-induced down-regulation of hippocampal endocannabinoid signaling may contribute to problems in behavioral

flexibility and may play a role in the development of perseveratory and ruminatory behaviors in stress-related neuropsychiatric disorders (Hill et al., 2005). Collectively, a majority of evidence supports a role for CB<sub>1</sub> receptors in the control of emotional behavior and suggests the existence of an anxiolytic endocannabinoid tone. Facilitation of such a tone by inhibiting the degradation of endocannabinoids in vivo may be therapeutically exploited, as indicated by the reduced anxiety-like behavior and potent antidepressant-like effects in mice and rats treated with a FAAH or anandamide transport inhibitor and the blockade of this effect by SR141716 or AM281 (Kathuria et al., 2003; Gobbi et al., 2005; Bortolato et al., 2006; Rutkowska et al., 2006).

The mechanisms responsible for the effects of cannabinoids on anxiety-related responses are complex and may involve modulation of numerous neurotransmitter systems. For example, stimulation of CB<sub>1</sub> receptors in rodents activates the HPA axis through the release of CRH (Weidenfeld et al., 1994; Wenger et al., 1997; Martin-Calderon et al., 1998; Manzanares et al., 1999a; Marco et al., 2004), which could account for the anxiogenic effects of high doses of cannabinoids (Rodriguez de Fonseca et al., 1996; Marin et al., 2002). In contrast, there are also examples of negative modulation of HPA function by endocannabinoids (Di et al., 2003; Patel et al., 2004). Cannabinoids also modulate GABAergic transmission and the release of the peptide cholecystokinin, which may contribute to both anxiolytic and anxiogenic effects (Onaivi et al., 1990; Katona et al., 1999, 2001; Marsicano and Lutz, 1999; Tsou et al., 1999; Beinfeld and Connolly, 2001; Rotzinger and Vaccarino, 2003). Furthermore, cannabinoids enhance the release of endogenous opioids and a functional interplay between the endocannabinoid and opioid systems modulates analgesic responses and is involved in antidepressant-like effects and in various addiction-related processes (Pugh et al., 1997; Manzanares et al., 1999b; Houser et al., 2000; Zimmer et al., 2001; Ghozland et al., 2002). From studies with THC and CP55,940, it appears that  $\mu$ - and  $\delta$ -opioid receptors mediate certain anxiolytic effects, whereas activation of  $\kappa$ -opioid receptors leads to increased anxiety (Pugh et al., 1997; Houser et al., 2000; Zimmer et al., 2001; Berrendero and Maldonado, 2002; Ghozland et al., 2002; Marin et al., 2003). There are also interactions between the endocannabinoid and serotonergic systems (Arevalo et al., 2001; Malone and Taylor, 2001; Fride and Shohami, 2002; Marin et al., 2003; Marco et al., 2004; Steffens and Feuerstein, 2004; reviewed in Viveros et al., 2005), although their role in anxiety-like behaviors has not been explored

In contrast to earlier dogma, recent findings indicate that neurogenesis occurs in the adult brain. Furthermore, stress and depression decrease neurogenesis, particularly in the hippocampus, whereas electroconvulsive therapy and chronic treatment with conventional antidepressants increases this process (reviewed in Pacher et al., 2004a). It has been recently demonstrated that the endocannabinoid system drives neural progenitor cell proliferation (Aguado et al., 2005, 2006), and cannabinoids promote neurogenesis (Berghuis et al., 2005; Jiang et al., 2005). Furthermore, CB<sub>1</sub> receptors appear to be required for neuronal survival in the hippocampus (Bilkei-Gorzo et al., 2005). These findings are particularly exciting, as they raise the possibility of a role for endocannabinoids in antidepressant drug action. Indeed, CB<sub>1</sub> receptor density in the hippocampus and hypothalamus is increased by chronic tricyclic antidepressant treatment (Hill et al., 2006), and the amplification of the actions of endocannabinoids by the FAAH inhibitor URB597 was found to produce antidepressant-like effects in the mouse tail-suspension and rat forced-swim tests, without eliciting reward-related effects indicative of addictive potential (Gobbi et al., 2005). It should not be surprising, however, that based on the basis of the bimodal action of cannabinoids on mood and anxiety, a case could be made for the opposite, i.e., for the antidepressant potential of CB<sub>1</sub> antagonism. CB<sub>1</sub> antagonists were reported to elicit antidepressant-like behavioral effects in rodents and can increase the synaptic concentration of biogenic amines, much like antidepressants do (reviewed in Witkin et al., 2005). Thus, pharmacological modulation of the endocannabinoid system holds considerable promise in the treatment of both anxiety-related and mood disorders.

The results of a recent study implicated endocannabinoids and CB<sub>1</sub> receptors in the extinction of aversive memories by demonstrating that CB<sub>1</sub> knockout mice show impaired extinction in auditory fear-conditioning tests, and this could be mimicked in wild-type mice by treatment with SR141716 (Marsicano et al., 2002b). These exciting findings raise the possibility that pharmacological amplification of CB<sub>1</sub> signaling, for example, by FAAH inhibitors, may have therapeutic value in obsessive-compulsive disorder or post-traumatic shock syndrome.

**9. Insomnia** Insomnia, the most common sleep disorder, is defined as difficulty with the initiation, maintenance, duration, or quality of sleep that results in the impairment of daytime functioning, despite adequate opportunity and circumstances for sleep (Silber, 2005). The cause for insomnia is often not known, but frequently it may be a consequence of a chronic disease associated with pain or depression.

Early studies documented the fact that marijuana and THC affect sleep patterns both in humans (Freeman, 1972, 1982; Pivik et al., 1972; Barratt et al., 1974; Feinberg et al., 1975, 1976) and in experimental animals (Monti, 1977; Buonamici et al., 1982). More recently, Nicholson et al. (2004) have studied the effects of cannabis extracts on nocturnal sleep, early-morning performance, memory, and sleepiness in a placebo-controlled, double-blind, crossover study in eight healthy volunteers. They found that 15 mg of THC was sedative, whereas 15 mg of CBD had alerting properties as it increased wake activity during sleep and counteracted the residual sedative activity of THC (Nicholson et al., 2004).

Anandamide was also found to modulate sleep by increasing slow-wave sleep two and rapid eye movement sleep in a CB<sub>1</sub> receptor-dependent manner in rats (Murillo-Rodriguez et al., 1998, 2001). Moreover, CB<sub>1</sub> receptor expression in the pons of rats was modulated by the light/dark cycle and by sleep (Martinez-Vargas et al., 2003), and endocannabinoids and CB<sub>1</sub> receptors were also implicated in rapid eye movement sleep rebound (Navarro et al., 2003). Interestingly, a recent study has demonstrated that anandamide not only induced sleep but also increased levels of the sleep-inducing substance adenosine in the basal forebrain, and both of these effects were blocked by SR141716 (Murillo-Rodriguez et al., 2003).

Oleamide is a fatty acid amide with a variety of in vitro effects, including inhibition of gap junction-mediated cell-cell communication (Boger et al., 1998a,b), modulation of 5-HT<sub>1</sub>, 5-HT<sub>2A,C</sub>, and 5-HT<sub>7</sub> receptors (Thomas et al., 1997, 1999; Hedlund et al., 1999), and modulation of inhibitory ionotropic receptors such as the GABA<sub>A</sub> receptor (Coyne et al., 2002). Oleamide accumulates in the cerebrospinal fluid of sleep-deprived cats (Cravatt et al., 1995) and rats (Basile et al., 1999) and induces sleep, an effect which could be blocked by SR141716 (Mendelson and Basile, 1999). Initially, it was suggested that inhibition of anandamide degradation by FAAH rather than the activation of CB<sub>1</sub> receptors was responsible for the sleep-inducing effect of oleamide (Boring et al., 1996; Mechoulam et al., 1997), but this is a matter of dispute (Fowler, 2004; Lees and Dougalis, 2004; Leggett et al., 2004).

Although little is known about the role of the endocannabinoid system in the pathophysiology of sleep disorders, clinical studies uniformly report significantly improved sleep quality in patients taking cannabinoids for symptomatic treatment of multiple sclerosis, cancer, chronic pain, or intractable pruritus. Although psychotropic cannabinoids are unlikely to gain acceptance for the treatment of insomnia, FAAH inhibitors were shown to enhance certain endocannabinoid-mediated behaviors without evidence for addictive properties (Kathuria et al., 2003). The sleep-inducing property of some potent FAAH inhibitors, such as the endogenous lipid 2-octyl  $\gamma$ -bromoacetoacetate (Boger et al., 1998a), could therefore be therapeutically exploited.

**10. Nausea and Emesis** Nausea and vomiting can present as symptoms of a variety of diseases or as secondary consequences of chemotherapy or radiotherapy of cancer. It is for this latter indication that

THC has gained acceptance as a highly efficacious therapeutic agent, often effective in cases resistant to other, more conventional, medications (reviewed by [Martin and Wiley, 2004](#); [Aapro, 2005](#); [Hall et al., 2005](#)). Emesis is thought to involve activation of specific receptors on sensory nerve endings in the gut and also in brainstem regions including the medullary chemoreceptor trigger zone and the lateral reticular formation. Activation of 5-HT<sub>3</sub> receptors appears to play a dominant role in acute emesis, whereas activation of NK<sub>1</sub> (substance P) receptors is more important in the delayed emesis after chemotherapy, as indicated by the effectiveness of the respective receptor antagonists in controlling these different stages of the emetic response ([Aapro, 2005](#)). Although the mechanism of the antiemetic action of cannabinoids is not quite clear, an interaction with 5-HT<sub>3</sub> is suggested by the colocalization of CB<sub>1</sub> and 5-HT<sub>3</sub> receptors on GABAergic neurons where they have opposite effects on GABA release ([Morales et al., 2004](#)). Also, cannabinoids may directly inhibit 5-HT<sub>3</sub>-gated ion currents by a mechanism not involving CB<sub>1</sub> receptors ([Fan, 1995](#); [Barann et al., 2002](#)). Such a CB<sub>1</sub> receptor-independent effect is also suggested by the ability of cannabidiol, a natural constituent of marijuana which does not bind to the CB<sub>1</sub> receptor, to reduce lithium-induced vomiting in the house musk shrew ([Parker et al., 2004](#)). Nevertheless, the involvement of CB<sub>1</sub> receptors is clearly indicated by the ability of SR141716 to reverse the effects of THC and synthetic agonists in suppressing vomiting caused by cisplatin ([Darmani, 2001b](#)) or lithium chloride ([Parker et al., 2004](#)), or by the ability of these agonist to reverse the emesis elicited by SR141716 in the least shrew ([Darmani, 2001a](#)). These latter findings suggest that the emetic circuitry is tonically controlled by endocannabinoids.

In line with such a possibility, a recent human study found an association between chronic marijuana use, which probably results in desensitization of cannabinoid receptors, and cyclical hyperemesis: in the 19 subjects studied, the hyperemetic episodes subsided upon discontinuation of cannabis use and reappeared upon rechallenge with cannabis ([Allen et al., 2005](#)). A meta-analysis of 30 randomized comparisons of cannabis (nabilone, dronabinol, or levonantradol) with placebo or standard antiemetics, involving a total of 1366 patients, concluded that cannabinoids are slightly more effective than conventional antiemetics, and the patients prefer them because of their mood enhancing and sedative effects. However, they were also more toxic, with dizziness, dysphoria, hallucinations, and paranoia being the most prominent undesirable side effects ([Tramér et al., 2001](#)). This led to the recommendation to limit the use of cannabinoids as antiemetics to patients with chemotherapy-related sickness, in whom their mood-enhancing effects would be of added benefit.

**11. Drug Addiction and Alcohol Disorders** The positive reinforcing effect of natural rewards, such as those derived from eating, drinking, work, or sexual activity, are mediated by the brain's reward circuitry. Neuroanatomically, this circuitry consists of three series of coupled pathways. First-order neurons project from structures in the ventral limbic forebrain (orbitofrontal cortex and anterior cingulate area) to the mesencephalic ventral tegmental area (VTA) where they synapse onto dopaminergic neurons. These second-order neurons project primarily to neurons in the shell of the nucleus accumbens (nAc), but also to cortical areas and to the amygdala. Third-order neurons in the nAc, some of which are GABAergic, project to the ventral pallidum and other regions involved in mediating reward-related behaviors (recently reviewed by [Lupica et al., 2004](#); [Gardner, 2005](#)). It is believed that addictive drugs activate or "hijack" the same pathway. Genetic vulnerability to drug addiction has been linked to a functional deficiency in the second-order dopaminergic neurons at their interface with third-order neurons in the nAc ([Nestler, 2003](#)). In human subjects prone to addiction, a deficiency in D<sub>2</sub> dopamine receptors in the nAc could be documented by brain imaging ([Volkow et al., 1997, 1999](#)).

A common denominator among different addictive drugs interacting with distinct receptors is their ability to activate the mesolimbic dopaminergic reward pathway and increase dopamine levels in the

nAc, which is believed to be responsible for their addictive properties (Koob, 1992; Wise, 2004). Similar to other drugs of abuse, THC increases extracellular dopamine levels in the nAc via activation of CB<sub>1</sub> receptors (Chen et al., 1990; Tanda et al., 1999) and also lowers the reward threshold for electrical brain stimulation (Gardner et al., 1988), a phenomenon known to involve activation of the mesolimbic dopamine system. THC also increases the firing rate of the second-order VTA-nAc dopaminergic neurons via CB<sub>1</sub> but not opiate receptors (French, 1997), and withdrawal from THC increases corticotropin-releasing factor levels in the central nucleus of the amygdala (Rodriguez de Fonseca et al., 1997), another hallmark of drugs of abuse (Koob, 1996).

THC and related synthetic cannabinoid agonists also fulfill the reward-related behavioral criteria for drugs of abuse: they support conditioned place preference (CPP) under appropriate conditions (Lepore et al., 1995; Valjent and Maldonado, 2000; Zangen et al., 2006), they are self-administered intravenously or intracerebrally in a CB<sub>1</sub> antagonist-sensitive manner (Martellotta et al., 1998; Ledent et al., 1999; Braida et al., 2001; Zangen et al., 2006), and they reinstate cocaine-or heroine-seeking behavior in rats previously extinguished from self-administration (De Vries et al., 2001).

An issue of intense interest is the location of the CB<sub>1</sub> receptors mediating these effects. Similar to cannabinoids, opiates also increase the activity of dopaminergic neurons in the VTA. This effect has been shown to result from  $\mu$  receptor-mediated inhibition of GABA release from the terminals of inhibitory GABAergic interneurons, i.e., through a “disinhibitory” mechanism (Johnson and North, 1992). A similar mechanism has been postulated for cannabinoids by Cheer et al. (2000), who reported that local application of the cannabinoid agonist HU-210 to brain slices containing the VTA increased dopaminergic neuronal activity, which could be blocked by the GABA<sub>A</sub> antagonist bicuculline. In line with this, WIN 55,212-2 was found to suppress electrically evoked, but not muscimol-induced, inhibitory postsynaptic currents via CB<sub>1</sub> receptors in brain slices containing the VTA (Szabo et al., 2002). However, cannabinoids also inhibit glutamate release in the VTA, which would have an opposite effect on dopaminergic activity (Melis et al., 2004a). There is evidence for additional sites of action, such as CB<sub>1</sub> receptors on the terminals of GABAergic projection neurons that target GABA<sub>B</sub> receptors on VTA dopamine neurons resulting in their disinhibition (Riegel and Lupica, 2004). This pathway may be activated by ethanol, as indicated by the ability of the GABA<sub>B</sub> agonist baclofen to antagonize the increase in ethanol drinking caused by WIN 55,212-2 treatment of alcohol-preferring rats (Colombo et al., 2004). Activation of CB<sub>1</sub> receptors on glutamatergic terminals in the nAc was reported to inhibit glutamate release onto GABAergic neurons in the nAc that project to the VTA, which may also result in disinhibition of VTA dopaminergic neurons (Robbe et al., 2001). Indeed, both the VTA and the nAc may be sites of the rewarding effects of cannabinoids, as documented by the propensity of rats to self-administer THC into either site (Zangen et al., 2006).

Regardless of the exact location of presynaptic CB<sub>1</sub> receptors, their natural activation occurs through retrograde transmission, with their endogenous ligands being released from postsynaptic cells (Kreutzger and Regehr, 2001; Ohno-Shosaku et al., 2001; Wilson and Nicoll, 2001). This mechanism has also been implicated in LTD (Gerdeman et al., 2002; Robbe et al., 2002), a form of synaptic plasticity that can be initiated by drugs of abuse (Thomas et al., 2001), and may be involved in certain features of compulsive drug use (Gerdeman et al., 2003). A further indication that endocannabinoids may be involved in mechanisms of drug reward is findings that the neurochemical and behavioral responses to different classes of drugs of abuse can be inhibited by the CB<sub>1</sub> receptor antagonists. These findings suggests that endocannabinoid activation of CB<sub>1</sub> receptors in the mesolimbic reward pathway may be part of a “common pathway” of drug reward (reviewed in De Vries and Schoffelmeer, 2005; Maldonado et al., 2006). Examples of this are discussed below.

**a. Opiates** There is a large body of evidence indicating a reciprocal relationship between the

endocannabinoid and endogenous opioid systems in drug dependence (recently reviewed by [Fattore et al., 2005](#); [Vigano et al., 2005a,b](#)). This fact is not surprising, given that opioids and cannabinoids have a similar pharmacological profile at both the behavioral level (e.g., analgesia, hypothermia, catalepsy, and motor impairment) and cellular/molecular levels (both CB<sub>1</sub> and opiate  $\mu$  receptors are predominantly presynaptic, they are coupled to and share the same pool of G<sub>i</sub>/G<sub>o</sub> proteins, and have an overlapping brain distribution). There are numerous examples for opioid or cannabinoid reward-related effects being inhibited by both CB<sub>1</sub> and opiate  $\mu$  antagonists ([Fattore et al., 2005](#); [Gardner, 2005](#); [Vigano et al., 2005a,b](#)). The mechanisms underlying these reciprocal interactions are not clear, but they may involve heterodimerization of CB<sub>1</sub> and  $\mu$  opiate receptors, depletion of shared G protein pools and/or utilization of common postreceptor signaling pathways. In addition, the opiate/cannabinoid synergism observed in nAc/striatal neurons appears to require adenosine and A2a receptor signaling ([Yao et al., 2006](#)).

Here we will only review evidence that pertains to the potential involvement of endocannabinoids in the addictive, reward-related actions of opioids. Such evidence is based on the ability of pharmacological or genetic ablation of CB<sub>1</sub> receptors to prevent or inhibit opioid effects. CB<sub>1</sub> knockout mice were reported to be unable to acquire morphine self-administration ([Ledent et al., 1999](#); [Cossu et al., 2001](#)), to have reduced morphine withdrawal symptoms ([Ledent et al., 1999](#)), and not to develop CPP for morphine ([Martin et al., 2000](#)). A possible neurochemical correlate of these changes is the lack of morphine-induced dopamine release in the nucleus accumbens of CB<sub>1</sub> receptor knockout mice ([Mascia et al., 1999](#)), although more recently CB<sub>1</sub> blockade was found to reverse the morphine-induced decrease in ventropallidal GABA overflow without affecting the morphine-induced increase in dopamine release in the nAc ([Caillé and Parsons, 2006](#)). Treatment of wild-type mice and rats with a CB<sub>1</sub> antagonist elicits similar phenotypes ([Rubino et al., 2000](#); [Mas-Nieto et al., 2001](#); [Navarro et al., 2001, 2004](#)). These observations raise the therapeutic potential of chronic treatment with a CB<sub>1</sub> receptor antagonist in preventing or reversing the development of opiate dependence.

**b. Nicotine** Nicotine is the main neuroactive component in tobacco smoke and is responsible for its addictive properties. Nicotine's rewarding effects are mediated by the same mesolimbic dopaminergic pathway that is involved in the rewarding effects of many other addictive drugs ([Pontieri et al., 1996](#)). Therefore, it should not be unexpected that there is a positive synergism between nicotine and THC in paradigms used to reveal reinforcing effects ([Valjent and Maldonado, 2000](#)). A role of endocannabinoids in the rewarding effects of nicotine is indicated by the absence of nicotine-induced CPP in CB<sub>1</sub> knockout mice ([Castane et al., 2002](#)), although the acquisition of nicotine self-administration was not affected by the absence of CB<sub>1</sub> receptors in another study using an acute reinforcement paradigm ([Cossu et al., 2001](#)). On the other hand, SR141716 was reported to decrease nicotine operant self-administration ([Cohen et al., 2002](#)) and nicotine-induced CPP in rats ([Le Foll and Goldberg, 2004](#); [Forget et al., 2006](#)) and also to inhibit nicotine-induced dopamine release in the nucleus accumbens shell ([Cohen et al., 2002](#)). SR141716 also inhibited nicotine self-administration sustained by nicotine-associated cues in the absence of nicotine itself ([Cohen et al., 2005](#)), and chronic exposure to nicotine was reported to induce endocannabinoid release ([Gonzalez et al., 2002](#)). Furthermore, SR141716 abolished the anxiolytic effects of low-dose nicotine in mice and potentiated its anxiogenic effects at higher doses ([Balerio et al., 2006](#)). Together, these findings justified testing rimonabant in clinical trials to promote smoking abstinence. Indeed, the results of a recent multicenter phase III clinical trial in the United States indicate that a 10-week treatment of smokers with a daily oral dose of 20 mg of rimonabant with a follow-up period of 42 weeks doubled the odds of quitting smoking, was well tolerated, and also reduced the post-cessation weight gain by >80% ([Dale and Anthenelli, 2004](#)).

**c. Cocaine** Unlike THC, opiates and nicotine, cocaine does not increase the activity of dopaminergic neurons in the VTA but elevates synaptic levels of dopamine in the nAc by blocking dopamine reuptake

at the dopamine transporter (Giros et al., 1996). Therefore it is not surprising that cocaine-induced increases in dopamine in the nAc were found to be unaffected by genetic ablation of CB<sub>1</sub> receptors (Soria et al., 2005). Accordingly, CB<sub>1</sub> receptors do not appear to participate in the acute rewarding properties of cocaine, as indicated by the preserved acute cocaine self-administration and cocaine-induced CPP in CB<sub>1</sub> knockout mice (Martin et al., 2000; Cossu et al., 2001; Lesscher et al., 2005; Soria et al., 2005) or in mice treated with SR141716 (Tanda et al., 2000; De Vries et al., 2001; Caillé and Parsons, 2006). SR141716 treatment also did not affect the threshold-lowering effect of cocaine in the intracranial self-stimulation paradigm, although treatment with WIN 55,212-2 was able to achieve this, suggesting that CB<sub>1</sub> receptor stimulation might inhibit the reinforcing properties of cocaine (Fattore et al., 1999; Vlachou et al., 2003).

Other studies indicate, however, that endocannabinoid activation of CB<sub>1</sub> receptors may mediate the reinforcing effects of cocaine. SR141716 treatment decreased the sensitivity of rats to the reinforcing effects of cocaine in an intracranial self-stimulation paradigm (Deroche-Gamonet et al., 2001). The ability to acquire operant self-administration of cocaine was reduced in CB<sub>1</sub> knockout mice or in SR141716-treated wild-type mice, which also displayed a reduced maximal effort to obtain cocaine infusion in a progressive ratio schedule, compared with untreated wild-type mice (Martin et al., 2000; Soria et al., 2005). Furthermore, prior use of cannabis was found to enhance the “high” elicited by subsequent use of cocaine in humans (Foltin et al., 1993; Lukas et al., 1994) and also to hasten relapse in abstinent former cocaine users (Rawson et al., 1986). Furthermore, a recent genetic study found an association between an (AAT)<sub>n</sub> triplet repeat polymorphism in the *CNR1* gene encoding the CB<sub>1</sub> receptor with cocaine addiction in an African-Caribbean population (Ballon et al., 2006). Treatment with HU-210 promoted reinstatement of cocaine-seeking behavior in rats, whereas treatment with SR141716 prevented reinstatement (De Vries et al., 2001). Thus, the endocannabinoid system may be involved in the acquisition and consolidation of cocaine addiction as well as in relapse, through mechanisms other than an effect on the cocaine-induced increase in dopaminergic transmission in the nAc. These latter studies also predict the possible effectiveness of rimonabant in the treatment of cocaine addiction.

*d. Alcohol* Several lines of evidence indicate the involvement of the endocannabinoid system in alcohol drinking behavior (recently reviewed by Colombo et al., 2005). Chronic alcohol intake increases endocannabinoid levels in the limbic forebrain (Gonzalez et al., 2002) and decreases CB<sub>1</sub> receptor binding and signaling (Basavarajappa and Hungund, 2002). Studies in the late 1990s indicated the effectiveness of SR141716 in reducing voluntary ethanol intake in rodent models of ethanol drinking (Arnone et al., 1997; Colombo et al., 1998b; Freedland et al., 2001), whereas cannabinoid agonists promoted drinking (Gallate et al., 1999; Colombo et al., 2002). Operant self-administration of ethanol and relapse to drinking are also inhibited by SR141716 (Cippitelli et al., 2005; Economidou et al., 2006) and potentiated by chronic exposure to a cannabinoid agonist (Lopez-Moreno et al., 2005).

The possible role of the endocannabinoid system in ethanol preference was further indicated by observations of reduced voluntary ethanol drinking in CB<sub>1</sub> knockout compared with wild-type mice (Hungund et al., 2003; Poncelet et al., 2003; Wang et al., 2003; Lallemand and de Witte, 2004; Naassila et al., 2004; Thanos et al., 2005), although no difference was noted in one study (Racz et al., 2003). Sensitivity to alcohol is inversely related to the chance of becoming an alcoholic among humans (Schuckit, 1997), and the same inverse relationship was noted in CB<sub>1</sub> knockout mice and their wild-type littermates (Naassila et al., 2004). The reduced voluntary ethanol intake in CB<sub>1</sub> knockout mice was associated with reduced alcohol-induced CPP (Houchi et al., 2004; Thanos et al., 2005), a further indication of the role of CB<sub>1</sub> receptors in the rewarding effects of alcohol.

Similar to cannabinoids and other drugs of abuse, alcohol intake can also result in increased dopamine

release in the nAc ([Weiss et al., 1993](#); [Campbell and McBride, 1995](#)). The reported absence of such release in CB<sub>1</sub> knockout mice and the ability of SR141716 to block ethanol-induced dopamine release in wild-type mice further suggest the involvement of endocannabinoids in the reinforcing effects of ethanol. However, the brain site where ethanol-induced endocannabinoid release and CB<sub>1</sub> receptor activation occur is not yet known. The recent observation that microinjection of SR141716 into the prefrontal cortex of alcohol-preferring AA rats inhibited ethanol self-administration suggests that this region may be one of the sites involved ([Hansson et al., 2006](#)). In the same study, FAAH activity and CB<sub>1</sub> signaling were both reduced in the same brain region of AA rats compared with their nonpreferring ANA counterparts, and microinjection of the FAAH inhibitor URB597 increased ethanol self-administration ([Hansson et al., 2006](#)). Analogous findings in female FAAH knockout mice are their increased voluntary ethanol intake and decreased alcohol sensitivity ([Basavarajappa et al., 2006](#)). These findings suggest that increased anandamide tone secondary to decreased FAAH activity in the prefrontal cortex may be causally linked to high alcohol preference. Such a scenario would be compatible with evidence for an association between problem drug and alcohol use and a missense mutation in the human FAAH gene ([Sipe et al., 2002](#)).

A number of mediators have been implicated in the control of appetite for both food and alcohol. In the case of endocannabinoids, the regulation is “unidirectional”, i.e., endocannabinoids promote both food intake (see section III.A.3.) and alcohol drinking. Because both food intake and alcohol drinking activate the brain reward pathways, one might postulate that the role of endocannabinoids in promoting drinking behavior would be most prominent in the type of alcoholics who drink for the rewarding effects of alcohol, such as young binge-drinkers. The high alcohol preference of C57BL6 mice and the role of the endocannabinoid system mediating it were found to be age-dependent ([Wang et al., 2003](#)), which is compatible with such a possibility. In contrast, the effects of NPY and CRH on food intake and ethanol consumption are bidirectional: NPY increases food intake ([Clark et al., 1984](#)) but reduces ethanol consumption ([Thiele et al., 1998](#)), whereas CRH is anorectic ([Britton et al., 1982](#)) but promotes ethanol drinking ([George et al., 1990](#)). The effects of NPY and CRH on alcohol preference correlate with their effects on anxiety-like behaviors, NPY being anxiolytic ([Heilig et al., 1989](#)) and CRH being anxiogenic ([Koob and Thatcher-Britton, 1985](#)). We would predict that CB<sub>1</sub> antagonists will be more effective in reducing the drive to drink in younger people who drink for the rewarding effects of alcohol, whereas CRH antagonists or NPY agonists would be more effective in older, chronic alcoholics who more likely drink to suppress the negative affect and anxiety of alcohol withdrawal. This hypothesis may be tested by appropriately designed clinical trials. Studies to test the safety and efficacy of rimonabant in the treatment of alcoholism and alcohol abuse are currently underway at the National Institute on Alcohol Abuse and Alcoholism.

*e. Psychostimulants* 3,4-Methylenedioxymethamphetamine (MDMA, Ecstasy) is a psychostimulant abused for its euphoric and stimulant properties, and it is often used in combination with marijuana. Intracerebral self-administration of MDMA was found to be reduced in the presence of the cannabinoid agonist CP55,940 and increased after treatment with SR141716. These findings were interpreted to indicate synergism between the reinforcing effects of cannabinoids and MDMA and a reduction in the motivational value of MDMA by CB<sub>1</sub> blockade ([Brida and Sala, 2002](#)). In another study, the authors found that SR141716 blocked MDMA-induced CPP ([Brida et al., 2005](#)). Amphetamine-induced long-term synaptic depression in the amygdala could be blocked by the CB<sub>1</sub> antagonist AM251, mimicked by the agonist WIN 55,212-2, and occluded by the transport inhibitor AM404, suggesting that amphetamine-induced LTD and related behavioral effects may be mediated via endocannabinoid release ([Huang et al., 2003](#)). Together, these findings suggest that CB<sub>1</sub> antagonists may be of value in the treatment of addiction to psychostimulants, including amphetamine and MDMA.



## D. Cardiovascular and Respiratory Disorders

Besides their well known neurobehavioral and immunological actions, cannabinoids and their endogenous and synthetic analogs exert important cardiovascular effects. The underlying mechanisms are complex, involving direct effects on the vasculature ([Gebremedhin et al., 1999](#); [Járai et al., 1999](#); [Wagner et al., 2001b](#); [Wagner et al., 2005](#)) and myocardium ([Bonz et al., 2003](#); [Maslov et al., 2004](#); [Sterin-Borda et al., 2005](#)), as well as modulation of autonomic outflow through sites of action in the central ([Niederhoffer and Szabo, 2000](#); [Pfitzer et al., 2004](#)) and the peripheral nervous systems ([Ishac et al., 1996](#); [Malinowska et al., 1997](#); [Szabo et al., 2001](#); [Niederhoffer et al., 2003](#)). As for endogenous cannabinoids, their effects are also complicated by their rapid metabolism, which liberates arachidonic acid that can be further metabolized into vasoactive prostanoids (reviewed in [Mechoulam et al., 1998](#); [Kunos et al., 2000](#); [Randall et al., 2002](#); [Ralevic et al., 2002](#)).

Studies to date indicate that CB<sub>1</sub> receptors are much more important than CB<sub>2</sub> receptors in cardiovascular regulation, the latter so far being implicated only in ischemic preconditioning and ischemia/reperfusion (I/R) injury of the myocardium (see below). CB<sub>1</sub> receptors have been detected in the human, rat, and mouse myocardium where they mediate negative inotropy ([Bonz et al., 2003](#); [Bátkai et al., 2004b](#); [Pacher et al., 2004, b, 2005a,b,d](#); [Engeli et al., 2005](#); [Wagner et al., 2005](#)) and also in vascular tissues ([Gebremedhin et al., 1999](#); [Liu et al., 2000](#)), where their activation leads to vasodilation, and both of these effects appear to be involved in the hypotensive effect of anandamide ([Wagner et al., 2001a,b](#); [Bátkai et al., 2004a,b](#); [Pacher et al., 2004, b, 2005a,b,d](#)) in anesthetized rodents. Sympathetic nerve terminals contain presynaptic CB<sub>1</sub> receptors, stimulation of which inhibits norepinephrine release ([Ishac et al., 1996](#)), which contributes to the bradycardic effects of anandamide in vivo ([Wagner et al., 2001b](#)). Anandamide-induced cardiovascular depressor effects are devoid of a centrally mediated component ([Varga et al., 1996](#)), in contrast to the effects of certain synthetic cannabinoids, which cause centrally mediated sympathoexcitation ([Niederhoffer and Szabo, 2000](#); [Gardiner et al., 2001, 2002b](#)).

The vasorelaxant effect of endocannabinoids and synthetic cannabinoids in vitro are complex and display tissue and interspecies differences. They may involve CB<sub>1</sub> and TRPV<sub>1</sub> receptor- and NO-mediated or NO-independent mechanisms and also as yet undefined endothelial site(s) of action. A detailed discussion of these in vitro vasodilatory effects can be found in recent reviews ([Hillard, 2000](#); [Kunos et al., 2000, 2002](#); [Ralevic et al., 2002](#); [Randall et al., 2002, 2004](#); [Begg et al., 2005](#); [Pacher et al., 2005a,b](#)) and is beyond the scope of this review.

Compared with the growing body of information on the vascular effects of cannabinoids, less is known about cannabinoid-induced direct cardiac effects. Anandamide, *R*-methanandamide, and HU-210 dose dependently decrease contractile performance in isolated, electrically paced human atrial muscle, an effect blocked by the potent CB<sub>1</sub> antagonist AM251, whereas the involvement of CB<sub>2</sub> receptors, NO, or prostanoids could be excluded ([Bonz et al., 2003](#)). HU-210 also decreased left ventricular developed pressure in isolated perfused rat hearts through CB<sub>1</sub> receptor activation ([Maslov et al., 2004](#); [Krylatov et al., 2005](#)). Another study using isolated, perfused, rat Langendorff heart preparations to study the effects of anandamide, *R*-methanandamide, and palmitoylethanolamide on coronary perfusion pressure and left ventricular developed pressure suggested the involvement of a cardiac site of action distinct from CB<sub>1</sub> and CB<sub>2</sub> receptors ([Ford et al., 2002](#)).

Several studies have examined the in vivo hemodynamic effects of endocannabinoids and their synthetic analogs in rodents (recently reviewed in [Begg et al., 2005](#); [Pacher et al., 2005a,b](#)). Intravenous administration of anandamide causes a triphasic blood pressure response in anesthetized mice and rats, in which a prolonged hypotensive effect (phase III) is preceded by a transient, vagally mediated, fall in heart rate, cardiac and contractility, and blood pressure and an increase in total peripheral resistance

(phase I) followed by a brief, pressor response (phase II) associated with increased cardiac contractility (Varga et al., 1995; Lake et al., 1997b; Pacher et al., 2004, b, 2005d). Inhibition of the phase I bradycardic response by TRPV<sub>1</sub> receptor antagonists in rats (Malinowska et al., 2001) and the absence of both phase I and phase II responses in TRPV<sub>1</sub><sup>-/-</sup> mice (Pacher et al., 2004) imply that these components are mediated by TRPV<sub>1</sub> receptors. Additional central and vascular mechanisms may also be involved in the brief pressor response (phase II) in anesthetized rats (Kwolek et al., 2005). The third, prolonged hypotensive phase (phase III) is characterized by marked decreased cardiac contractility and slightly decreased total peripheral resistance, and it lasts up to 10 min in anesthetized mice (Pacher et al., 2004b, 2005d), similar to the hypotensive effect previously described in anesthetized but not conscious rats (Stein et al., 1996; Varga et al., 1996; Lake et al., 1997a,b; Gardiner et al., 2002a; Batkai et al., 2004b) and also observed with synthetic cannabinoids (Vidrio et al., 1996; Lake et al., 1997a; Pacher et al., 2005d).

The anandamide-induced phase III hypotension and decreased cardiac contractility, as well as similar hemodynamic responses to synthetic cannabinoids, are mediated by CB<sub>1</sub> receptors. First, these effects are prevented or reversed by selective CB<sub>1</sub> antagonists both in normal rodents (Varga et al., 1995, 1996; Calignano et al., 1997; Pacher et al., 2004, b, 2005a,d) and in mice lacking FAAH, which exhibit increased sensitivity to hypotensive and cardiodepressant effects of anandamide (Pacher et al., 2005d). Second, there is a positive correlation between the concentrations of various cannabinoid agonists in producing half-maximal hypotensive and bradycardic responses (EC<sub>50</sub>) and in their affinity constants for binding to CB<sub>1</sub> receptors in the brain (Lake et al., 1997a). Third, cannabinoid-induced hypotension and bradycardia are absent in mice lacking the CB<sub>1</sub> receptor (Jarai et al., 1999; Ledent et al., 1999). The involvement of the endocannabinoid system in various cardiovascular disorders is reviewed below.

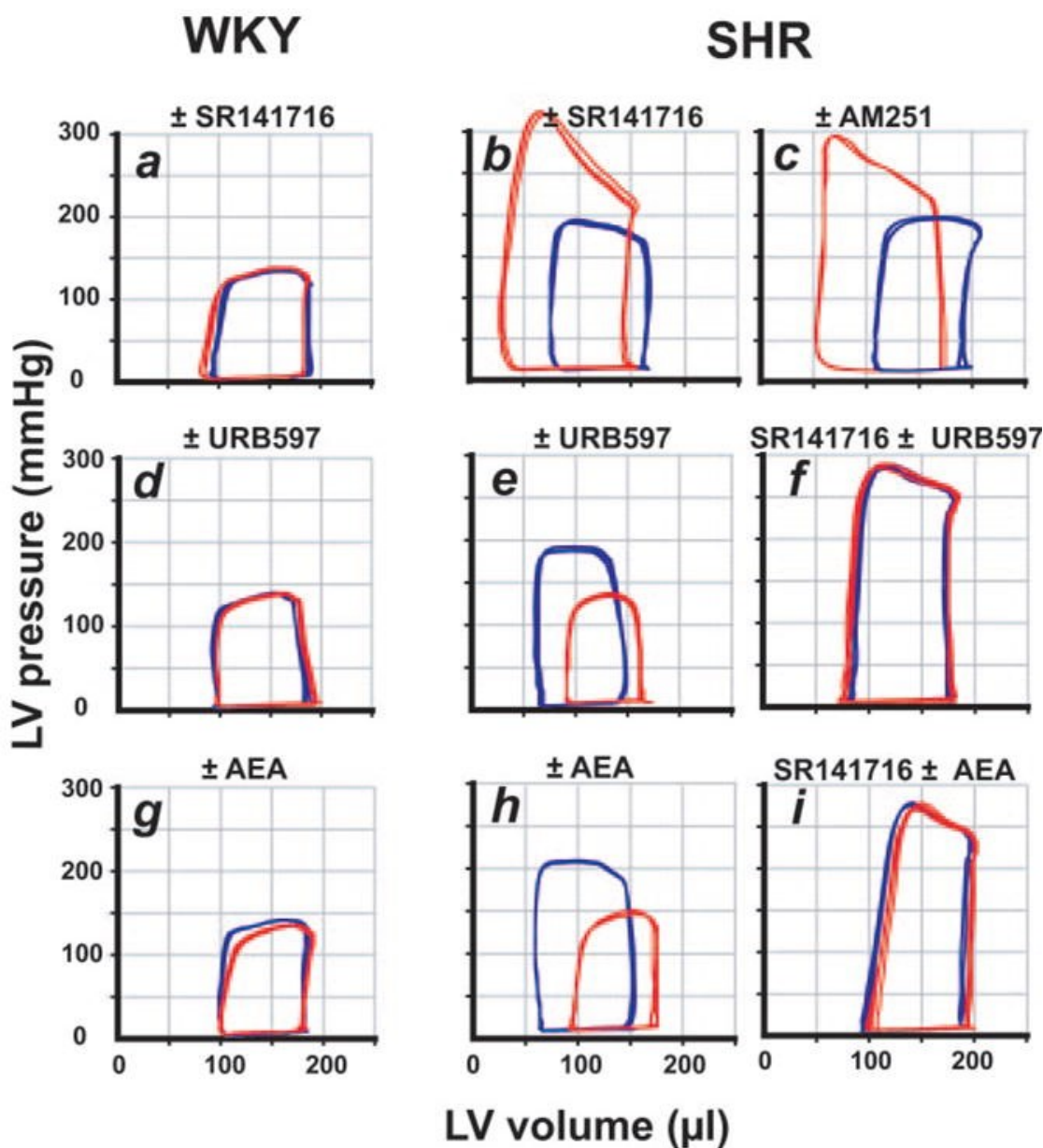
**1. Hypertension** Chronic use of cannabis in humans as well as both acute and prolonged administration of THC to experimental animals elicits a long-lasting decrease in blood pressure and heart rate (Rosenkratz and Braude, 1974; Benowitz and Jones, 1975), whereas the acute effect of smoking cannabis usually increases heart rate with no consistent change in blood pressure (Kanakis et al., 1976). In a recent study conducted in 63 male cannabis smokers, 22% of subjects experienced symptomatic hypotension, which could be reversed by the administration of 30 or 90 mg but not lower doses of rimonabant, indicating that CB<sub>1</sub> receptors mediate the hypotensive effect of cannabis smoking in humans (Gorelick et al., 2006).

More than three decades ago, several studies explored the potential use of cannabinoids to treat hypertension (Birmingham, 1973; Archer, 1974; Varma and Goldbaum, 1975; Adams et al., 1977; Crawford and Merritt, 1979; Zaugg and Kyncl, 1983). Unfortunately, the initial high anticipation was tempered by a report of the development of rapid tolerance to the hypotensive and bradycardic effects of THC (Adams et al., 1976) and by the failure to separate the cardiovascular and neurobehavioral effects of cannabinoids. Albeit a later study in spontaneously hypertensive rats (SHR) demonstrated no tolerance to the same effects during a 10-day treatment period (Kosersky, 1978), interest in this issue had vanished for the next two decades.

As with many other effects of marijuana, the discovery of endocannabinoids has focused attention on their possible role in cardiovascular regulation. Studies with SR141716 indicated that the hypotensive/bradycardic effects of exogenous anandamide, THC, and potent synthetic cannabinoids are mediated by CB<sub>1</sub> receptors (Varga et al., 1995; Lake et al., 1997a). CB<sub>1</sub> receptor knockout mice have normal blood pressure (Jarai et al., 1999; Ledent et al., 1999) and the blood pressure of normotensive mice and rats is unaffected or slightly reduced by CB<sub>1</sub> antagonists (Varga et al., 1995; Lake et al., 1997a; Varga Batkai et al., 2004b). In anesthetized rats, anandamide elicits only a modest and short-lasting hypotensive response (Varga et al., 1995; Lake et al., 1997a), whereas in conscious normotensive

rats it has no hypotensive effect at all ([Stein et al., 1996](#); [Lake et al., 1997b](#); [Gardiner et al., 2002](#)). Furthermore, inhibitors of anandamide transport or FAAH do not lower blood pressure in normotensive animals ([Calignano et al., 1997](#); [Bátkai et al., 2004b](#)), and mice deficient in FAAH have normal baseline hemodynamic characteristics and baroreceptor reflex function ([Pacher et al., 2005d](#)). As pointed out by a recent editorial ([Awumey et al., 2005](#)), these observations indicate a lack of involvement of endogenous cannabinoids in cardiovascular regulation under normal conditions.

In contrast, a number of observations indicate that endocannabinoids *are* involved in cardiovascular regulation in hypertension. Both THC ([Kosersky, 1978](#)) and anandamide ([Lake et al., 1997b](#), [Bátkai et al., 2004b](#)) induce larger and longer lasting hypotension in anesthetized SHR compared with normotensive controls, and the hypotensive effect of anandamide is preserved in conscious SHR ([Lake et al., 1997b](#)). Interestingly, inhalation of THC also resulted in a greater and longer lasting decrease of arterial blood pressure in hypertensive compared with normotensive individuals ([Crawford and Merritt, 1979](#)). By using a sophisticated pressure-volume analysis system, the hemodynamic effects of cannabinoid agonists and antagonists were evaluated in three different models of experimental hypertension ([Bátkai et al., 2004b](#)). In anesthetized SHR, the CB<sub>1</sub> antagonists AM251 and SR141716 both caused marked and sustained further increases in blood pressure and cardiac contractility ([Fig. 5](#)). Conversely, preventing the degradation or uptake of endogenous anandamide by treatment with the FAAH inhibitor URB597 or the transport inhibitor OMDM2 reduced blood pressure, cardiac contractility, and vascular resistance to levels observed in normotensive controls, and these effects were prevented by pretreatment with a CB<sub>1</sub> antagonist. Similar effects were seen in Dahl salt-sensitive rats and rats with angiotensin II-induced hypertension, whereas in the respective normotensive controls the same parameters remained unaffected by any of these treatments ([Bátkai et al., 2004b](#)) ([Fig. 5](#)). Anandamide and HU-210 induced more pronounced and longer lasting hypotension in SHR than in WKY rats. Unexpectedly, decreased cardiac contractility rather than a reduction in peripheral resistance was primarily responsible for the antihypertensive effect of anandamide, which was fully prevented by CB<sub>1</sub> antagonists, but was unaffected by the TRPV<sub>1</sub> antagonist capsazepine. In the same study, the expression of CB<sub>1</sub> receptors was found to be increased in the myocardium and the aortic endothelium of SHR compared with WKY rats.



[Fig. 5](#)

Effects of anandamide, URB597, SR141716, and AM251 on left ventricular (LV) function in normotensive and spontaneously hypertensive rats. Representative left ventricular pressure-volume (PV) loops from WKY rats (a, d, and g) and SHR (b, c, e, f, h, and i) before (black) and after (red) treatment with indicated agents or their combinations. A leftward shift of PV loops and an increase in amplitude (pressure) indicate increased LV contractility, whereas a rightward shift and decrease in amplitude indicate decreased LV function. Experiments were repeated in three more animals in each treatment group with similar results. AEA, anandamide. Reproduced with permission from [Bátkai et al. \(2004\) \*Circulation\* 110:1996–2002](#); © Lippincott Williams and Wilkins.

These findings point to the existence of an endocannabinoid tone in hypertension that limits the elevation of blood pressure and cardiac contractile performance through tonic activation of cardiac and probably vascular CB<sub>1</sub>. A possible underlying mechanism is the observed up-regulation of cardiac and

vascular CB<sub>1</sub> in SHR compared with their normotensive controls, although increased coupling of these CB<sub>1</sub> receptors may also contribute to the augmented sensitivity to the cardiovascular effects of anandamide (Bátkai et al., 2004b). A proposed alternative mechanism would involve up-regulation of vascular TRPV<sub>1</sub> receptors in hypertension, based on the reported ability of capsaizepine to partially inhibit the hypotensive effect of anandamide and *R*-methanandamide in hypertensive but not in normotensive rats (Li et al., 2003; Wang et al., 2005). However, capsaizepine is known to have nonspecific effects even at low concentrations (Ray et al., 2003), and up-regulation of TRPV<sub>1</sub> cannot account either for the increased hypotensive potency of HU-210 (Bátkai et al., 2004b), which is not a ligand for TRPV<sub>1</sub> receptors (Zygmunt et al., 1999), or for the dominant cardiac component in the hypotensive effect of exogenous or endogenous anandamide (Bátkai et al., 2004b). Also, physiological concentrations of endogenous anandamide are at least an order of magnitude lower than the micromolar concentrations required to activate TRPV<sub>1</sub> receptors.

A practical implication of these findings is that enhancing endocannabinoid tone by blocking the enzymatic degradation or cellular uptake of anandamide could be a novel therapeutic approach in the treatment of hypertension. Such a strategy has a number of desirable features: 1) unlike the generalized activation of CB<sub>1</sub> receptors by direct acting agonists, inhibition of FAAH causes a more restricted profile of cannabinoid-like effects with no indication of psychoactivity (Kathuria et al., 2003; Gobbi et al., 2005), probably related to the discrete distribution of FAAH in the brain; 2) FAAH or transport inhibitors have no hemodynamic effects under normotensive conditions, which predicts the absence of postural hypotension or other side effects; and 3) having a major effect on the inappropriately increased cardiac contractility, such treatment may be effective in reversing the cardiac hypertrophy that usually accompanies chronic hypertension.

**2. Circulatory Shock** The profound hypotension that can be elicited through pharmacological activation of CB<sub>1</sub> receptors (Lake et al., 1997a) triggered numerous studies to investigate the role of the endocannabinoid system in the hypotension associated with various forms of shock, including hemorrhagic (Wagner et al., 1997; Cainazzo et al., 2002), endotoxic (Varga et al., 1998; Wang et al., 2001; Liu et al., 2003a; Bátkai et al., 2004a; Gardiner et al., 2005; Kadoi et al., 2005), and cardiogenic shock (Wagner et al., 2001a, 2003), and the shock associated with necrotizing pancreatitis (Matsuda et al., 2005). Initial studies demonstrated that the putative CB<sub>1</sub> receptor antagonist SR141716 prevented or reversed the hypotension associated with hemorrhagic, endotoxic, and cardiogenic shock (Wagner et al., 1997, 2001a,b; Varga et al., 1998). Likewise, SR141716 reversed the hypotension associated with advanced liver cirrhosis (Bátkai et al., 2001; Ros et al., 2002), which is possibly secondary to the endotoxemia frequently found in patients with late-stage cirrhosis (Lumsden et al., 1988). Observations that circulating macrophages and platelets from endotoxemic or cirrhotic animals or humans had elevated levels of endocannabinoids and, when isolated and injected into normal rats, these cells elicited SR141716-sensitive hypotension also pointed toward the involvement of CB<sub>1</sub> receptors in many of these conditions (Wagner et al., 1997; Varga et al., 1998; Bátkai et al., 2001; Maccarrone et al., 2001, 2002; Ros et al., 2002; Liu et al., 2003a).

Several recent reports demonstrated that anandamide and some atypical cannabinoids can cause both cardiodepressant and vasodilatory effects via as-yet-undefined receptors sensitive to inhibition by SR141716 but not by AM251 (Járai et al., 1999; Ford et al., 2002; Ho and Hiley, 2003; O'Sullivan et al., 2004b), a selective CB<sub>1</sub> antagonist equipotent with SR141716 (Gatley et al., 1997). A recent study compared the effects of SR141716 and AM251 in rats on the acute hypotensive effect of bacterial endotoxin (LPS) administered as an intravenous bolus. Hypotension in this model is fully attributable to the decreased cardiac contractility, whereas peripheral vascular resistance is increased, indicating vasoconstriction (Biber et al., 1988; Cheng et al., 2003). Using this model, the cardiodepressant and

hypotensive effects of LPS were inhibited by SR141716 but not by AM251. Furthermore, LPS induced SR141716-sensitive hypotension in wild-type mice and in mice deficient in CB<sub>1</sub> or both CB<sub>1</sub> and CB<sub>2</sub> receptors, suggesting that receptors distinct from CB<sub>1</sub> or CB<sub>2</sub> are primarily responsible for the observed hypotension ([Bátkai et al., 2004a](#)). Interestingly, another recent study has demonstrated that the CB<sub>1</sub>-selective cannabinoid antagonist AM281 prevented the hemodynamic changes induced by acute LPS injection in rats ([Kadoi et al., 2005a](#)). Other results indicate that endocannabinoids may also contribute to endotoxin-induced hypotension indirectly, through CB<sub>1</sub>-mediated prejunctional inhibition of sympathoexcitation ([Godlewski et al., 2004](#)). In a different shock model in which continuous infusion of LPS in conscious rats causes marked peripheral vasodilatation and increased cardiac output, AM251 attenuated the tachycardic and hind quarter vasodilator effects of LPS. This result was attributed to modulation of  $\beta$ -adrenergic vasodilation, rather than suppression of a direct vasodilator effect by endocannabinoids ([Gardiner et al., 2005](#)). Interestingly, in a recent study, [Matsuda et al. \(2005\)](#) demonstrated that AM251 improved mean arterial pressure and survival rate in models of severe acute necrotizing pancreatitis without affecting inflammatory changes, which suggests the involvement of cardiac or vascular CB<sub>1</sub> receptors in the hypotension associated with this condition.

In hemorrhagic, cardiogenic, and endotoxic shock, the cannabinoid agonists HU-210, WIN 55,212-2, and THC improved endothelial function and/or survival ([Wagner et al., 1997, 2001a, 2003](#); [Varga et al., 1998](#); [Smith et al., 2000, 2001](#)). Surprisingly, the use of cannabinoid receptor antagonists, including SR141716, AM281, AM251, and SR144528, also leads to survival benefits in endotoxic and septic shock or necrotizing pancreatitis ([Varga et al., 1998](#); [Smith et al., 2000, 2001](#); [Cainazzo et al., 2002](#); [Kadoi et al., 2005a,b](#); [Matsuda et al., 2005](#)). In contrast, CB<sub>1</sub> receptor blockade increased mortality in hemorrhagic ([Wagner et al., 1997](#)) and cardiogenic shock ([Wagner et al., 2001a, 2003](#)), despite the increase in blood pressure. In these latter conditions, endocannabinoid-mediated vasodilation may have survival value through improving tissue oxygenation by counteracting the excessive sympathetic vasoconstriction triggered by hemorrhage or myocardial infarction, and this would be removed by CB<sub>1</sub> blockade. In contrast, CB<sub>1</sub> blockade may improve survival in endotoxic shock by preventing the primary hypotensive response to LPS (reviewed in [Kunós et al., 2000](#); [Hiley and Ford, 2003, 2004](#); [Pacher et al., 2005a,c](#)).

It should also be kept in mind that in most of the above conditions, hemodynamic changes are triggered by overwhelming inflammatory reaction, increased oxidative stress, and activation of downstream effector pathways, eventually leading to cardiovascular dysfunction and failure (reviewed in [Evgenov and Liaudet, 2005](#); [Pacher et al., 2005e](#)). Therefore, the well known immune-modulatory, anti-inflammatory, and antioxidant effects of cannabinoids should not be overlooked in these conditions. Indeed, endocannabinoids and synthetic cannabinoid agonists decrease inflammatory cytokine release in endotoxin-stimulated cells and in endotoxin-challenged animals (reviewed in [Walter and Stella, 2004](#); [Klein et al., 2005](#)). Surprisingly, SR141716 and the CB<sub>2</sub> antagonist SR144528 were also reported to have anti-inflammatory effects ([Smith et al., 2000, 2001](#)), which may be attributed to their inverse agonist properties or to CB<sub>1/2</sub> receptor-independent mechanisms (reviewed in [Begg et al., 2005](#); [Pertwee, 2005b,c](#)).

Collectively, it appears that both cannabinoids and antagonists of cannabinoid receptors may exert some beneficial effects in various rodent shock models. Further studies should establish the specificity of these effects and the relevance to various forms of circulatory shock in humans.

**3. Myocardial Reperfusion Injury** The endocannabinoid system has been implicated in endotoxin-induced preconditioning against myocardial I/R injury ([Lagneux and Lamontagne, 2001](#)). In this study, the effects of 90 min of low-flow ischemia followed by 60 min of reperfusion at normal flow were compared in isolated hearts from rats pretreated with LPS or saline. Endotoxin pretreatment enhanced

functional recovery on reperfusion and reduced infarct size compared with controls, and pretreatment with the CB<sub>2</sub> antagonist SR144528 but not the CB<sub>1</sub> antagonist SR141716 abolished the beneficial effects of preconditioning ([Lagneux and Lamontagne, 2001](#)). In a follow-up study, SR144528 but not SR141716 also abolished the infarct size-reducing effect of preconditioning induced by heat stress ([Joyeux et al., 2002](#)). These initial studies have suggested that the protection was mediated by endocannabinoids acting on CB<sub>2</sub> receptors. In preconditioning induced by a brief period of ischemia (5 min), either CB<sub>2</sub> or CB<sub>1</sub> receptor blockade abolished the protection, and both CB<sub>1</sub> and CB<sub>2</sub> receptors were implicated in the preservation of endothelium-dependent, 5-HT-induced vasodilation by ischemic preconditioning ([Bouchard et al., 2003](#)). Perfusion of isolated rat hearts with PEA or 2-AG but not anandamide afforded protection against ischemia by improving myocardial recovery and decreasing myocardial damage and infarct size ([Lepicier et al., 2003](#)). The cardioprotective effect of both PEA and 2-AG were completely blocked by SR144528, whereas SR141716 partially inhibited the effect of 2-AG only ([Lepicier et al., 2003](#)). Likewise, the selective CB<sub>1</sub> agonist ACEA and the selective CB<sub>2</sub> agonist JWH-015 both reduced infarct size in this model, and the CB<sub>2</sub> receptor-mediated cardioprotection by PEA involved activation of p38/extracellular signal-regulated kinases 1 and 2 and protein kinase C ([Lepicier et al., 2003](#)). In another study using isolated perfused rat hearts subjected to ischemia and reperfusion, reduction of the infarct size by anandamide could be equally well antagonized by CB<sub>1</sub> or CB<sub>2</sub> antagonists but could not be mimicked by selective CB<sub>1</sub> or CB<sub>2</sub> agonists, suggesting the involvement of a site distinct from CB<sub>1</sub> or CB<sub>2</sub> receptors ([Underdown et al., 2005](#)).

Others have used whole animal models of I/R injury induced by coronary occlusion/reocclusion in anesthetized rats. In this model, anandamide and HU-210 both decreased the incidence of ventricular arrhythmias and reduced infarct size through activation of CB<sub>2</sub> but not CB<sub>1</sub> receptors ([Krylatov et al., 2001, 2002a,b,c](#); [Ugdyzhkova et al., 2001, 2002](#)). The moderately CB<sub>2</sub>-selective agonist WIN 55,212-2 also reduced the extent of leukocyte-dependent myocardial damage in a more recent mouse study of myocardial I/R in vivo. This effect was abolished by the selective CB<sub>2</sub> receptor antagonist AM630 but was unaffected by AM251 ([Di Filippo et al., 2004](#)). In summary, evidence to date indicates that endocannabinoids protect against myocardial ischemic injury models predominantly via CB<sub>2</sub> receptors.

**4. Atherosclerosis** Chronic inflammation and the associated oxidative-nitrosative stress are key players in atherosclerosis and cardiovascular aging, and pharmacological modulation of these processes could be of therapeutic benefit (reviewed in [Csiszar et al., 2005](#); [Libby and Theroux, 2005](#)). Using the apolipoprotein E knockout mouse model of atherosclerosis, [Steffens et al. \(2005\)](#) reported that orally administered THC significantly inhibited disease progression. Furthermore, CB<sub>2</sub> receptor expressing immune cells were present both in human and mouse atherosclerotic plaques, lymphoid cells isolated from THC-treated mice had diminished proliferation capacity and decreased interferon- $\gamma$  production, and THC inhibited macrophage chemotaxis in vitro. Most importantly, all of these effects were completely blocked by a selective CB<sub>2</sub> receptor antagonist, suggesting that targeting CB<sub>2</sub> receptors may offer a new approach in the treatment of atherosclerosis ([Roth, 2005](#); [Steffens et al., 2005](#)).

**5. Asthma** The effect of marijuana on airway functions was among the first to be explored for potential therapeutic benefit (reviewed in [Lemberger, 1980](#); [Tashkin et al., 2002](#)). Smoking marijuana and ingesting THC were both found to increase airway conductance in normal, healthy subjects ([Tashkin et al., 1973](#); [Vachon et al., 1973](#)), and these effects lasted longer than the bronchodilator effect of the  $\beta$ -adrenergic agonist isoproterenol. Bronchodilation induced by smoked marijuana and oral THC was also documented in subjects with mild to moderate asthma and in asthmatic patients with methacholine- or exercise-induced bronchoconstriction ([Tashkin et al., 1974, 1975](#)). Bronchodilation without side effects was observed in asthmatic patients after a low dose (0.2 mg) of nebulized THC ([Williams et al., 1976](#); [Hartley et al., 1978](#)). In contrast, aerosols containing larger doses of THC (5–20 mg) caused

paradoxical bronchoconstriction attributed to local irritation ([Tashkin et al., 1977](#)). In another study of normal and asthmatic subjects, orally administered THC elicited only minimal and inconsistent bronchodilation associated with significant CNS side effects ([Abboud and Sanders, 1976](#)). Nevertheless, most of these initial observations had suggested some therapeutic benefit of using cannabinoids in asthma.

As for the mechanisms underlying THC-induced bronchodilation, the potential involvement of  $\beta$ -adrenergic and muscarinic receptors on airway smooth muscle could be excluded ([Kelly and Butcher, 1973](#); [Shapiro et al., 1977](#); [Lemberger, 1980](#)). This conclusion was supported by the inability of THC to relax isolated rings of resting or precontracted human bronchioles ([Orzelek-O'Neil et al., 1980a,b](#)), suggesting a more proximal site of action in the lung ([Cavero et al., 1972](#)) or a central mechanism.

More recently, [Calignano et al. \(2000\)](#) reported that CB<sub>1</sub> receptors are present on axon terminals innervating airway smooth muscle, and anandamide inhibited capsaicin-induced bronchospasm and cough in guinea pigs in an SR141716-sensitive manner. They also documented calcium-induced biosynthesis of anandamide in lung tissue, suggesting that locally generated anandamide participates in the intrinsic control of airway responsiveness by inhibiting prejunctional acetylcholine release. Indeed, SR141716 treatment was found to enhance capsaicin-evoked bronchospasm and cough. Interestingly, when airway smooth muscle was completely relaxed by vagotomy and atropine treatment, anandamide caused dose-dependent bronchoconstriction, which could be also prevented by CB<sub>1</sub> blockade. This effect was tentatively attributed to direct stimulation of putative cannabinoid receptors on the airway smooth muscle or a CB<sub>1</sub>-mediated corelease of bronchoconstrictor neurotransmitters from nerve endings in the lung. In a follow-up study, presynaptic CB<sub>1</sub> receptors in the guinea pig lung were only found on noradrenergic terminals where their stimulation by WIN 55,212-2 inhibited norepinephrine release ([Vizi et al., 2001](#)), consistent with the lack of a mediated CB<sub>1</sub>-mediated effect on acetylcholine release in guinea pig trachea ([Spicuzza et al., 2000](#)). In contrast to the findings of [Calignano et al. \(2000\)](#), [Stengel et al. \(1998\)](#) reported that anandamide given either intravenously or in aerosol did not affect airway resistance in guinea pigs, but possessed modest anti-inflammatory properties. It should be noted, however, that in this study bronchoconstriction was induced by a calcium ionophore rather than capsaicin. In an in vitro study of guinea pig airway smooth muscle ([Yoshihara et al., 2005](#)), anandamide and palmitoylethanolamide inhibited contractions elicited by electrical field stimulation but not by neurokinin A, and also blocked capsaicin-induced release of substance P-like immunoreactivity. These effects were selectively inhibited by a CB<sub>2</sub> but not a CB<sub>1</sub> antagonist, or by maxi-K<sup>+</sup> channel blockers, suggesting that CB<sub>2</sub> agonists may have therapeutic value in asthma ([Yoshihara et al., 2005](#)). In a recent study, inhibition of anandamide transport potently suppressed capsaicin-induced cough in mice, suggesting that the anandamide transporter may be a target for peripherally acting antitussive medications ([Kamei et al., 2006](#)). Diverse effects of endocannabinoids and synthetic agonist have also been reported on respiratory function and pulmonary circulation both in vivo and in vitro ([Schmid et al., 2003](#); [Wahn et al., 2005](#)).

Allergic asthma is currently viewed as a complex inflammatory disorder characterized by recruitment of eosinophils into the lung, mucus hypersecretion by goblet cells, elevated serum IgE, and airway hyperresponsiveness (reviewed in [Wills-Karp, 1999](#)). Given the well known anti-inflammatory effects of cannabinoids, these effects could also be of therapeutic value. Indeed, in a murine model of allergic airway disease induced by ovalbumin sensitization, pretreatment with cannabidiol or THC blunted the increase in IL-2, IL-4, IL-5, and IL-13 mRNA expression and decreased mucus overproduction and serum IgE levels ([Jan et al., 2003](#)). Anti-inflammatory effects of WIN 55,212-2, THC, anandamide, and palmitoylethanolamide were also reported in a mouse model of LPS-induced pulmonary inflammation ([Berdyshev et al., 1998](#)).



In conclusion, the effects of cannabinoids on respiratory function are rather complex, and evidence for their therapeutic potential in asthma is equivocal. The possibility remains that novel, nonpsychoactive cannabinoid analogs with long-lasting anti-inflammatory activity turn out to be useful adjuncts in the treatment of allergic asthma.

### E. Eye Disorders (Glaucoma and Retinopathy)

Glaucoma, the leading cause of irreversible blindness in the United States, is characterized by an increase in intraocular pressure and consequent damage to the optic nerve. Despite the multitude of effective medications that can be used to decrease ocular hypertension (e.g., cholinergic agonists,  $\beta$ - and  $\alpha_2$ -adrenoceptor agonists, dopaminergic agonists, prostaglandins, and carbonic anhydrase inhibitors), some patients remain refractory to these drugs and may eventually become blind (reviewed in [Alward, 1998](#); [Crowston and Weinreb, 2005](#)).

A decrease in intraocular pressure in a small number of healthy marijuana smokers was a serendipitous finding ([Hepler and Frank, 1971](#)), subsequently confirmed in a placebo-controlled, double-blind study of healthy volunteers who smoked either natural marijuana of known THC content or ingested synthetic THC ([Hepler et al., 1972](#)). THC or marijuana decreased intraocular pressure whether administered orally, topically, or intravenously, with no major tolerance to their effect reported ([Shapiro, 1974](#); [Purnell and Gregg, 1975](#); [Cuendet et al., 1976](#); [Hepler et al., 1976](#); [Brown et al., 1977](#); [Merritt et al., 1980, 1981a,b](#)). Most of these studies also reported various systemic side effects, such as hypotension, tachycardia, euphoria, and dysphoria, as well as other ocular effects, such as changes in pupil size, decreased tear production, and conjunctival hyperemia. Endocannabinoids and synthetic cannabinoid ligands have also been reported to reduce intraocular pressure when given topically or systemically, both in animals and humans ([Shapiro, 1974](#); [ElSohly et al., 1981, 1984](#); [Colasanti et al., 1984a,b,c](#); [Pate et al., 1995](#); [Porcella et al., 1998](#); [Buchwald et al., 2002](#); [Laine et al., 2002a,b](#); reviewed in [Jarvinen et al., 2002](#); [Chien et al., 2003](#); [Tomida et al., 2004](#)).

Early investigations into the mechanisms of the intraocular pressure-lowering effect of marijuana and THC implicated the sympathetic and central nervous systems in this effect ([Green and Pederson, 1973](#); [Green and Podos, 1974](#); [Green et al., 1977a,b](#)). However, in subsequent studies, the effect of a unilateral topical application of cannabinoids was limited to the treated eye, pointing toward a local site of action ([Colasanti et al., 1984a,b,c](#)). Indeed, a CNS site of action could be ruled out by the lack of change in intraocular pressure upon intracerebroventricular or ventriculocisternal application of THC in rabbits ([Liu and Dacus, 1987](#)).

Multiple lines of evidence suggest that endocannabinoids and cannabinoid receptors, in particular CB<sub>1</sub>, play an important role in the regulation of intraocular pressure, and topically applied cannabinoids and cannabinoid ligands may be of significant benefit in the treatment of glaucoma (reviewed in [Jarvinen et al., 2002](#); [Tomida et al., 2004](#)). First, CB<sub>1</sub> receptors are expressed in the rat ciliary body ([Porcella et al., 1998](#)), in human ciliary epithelium, ciliary muscle, ciliary body vessels, trabecular meshwork, Schlemm's canal, and retina ([Straiker et al., 1999a](#); [Porcella et al., 2000](#); [Stamer et al., 2001](#)), and the retina of a variety of species ([Straiker et al., 1999b](#); [Yazulla et al., 1999, 2000](#)). Second, ocular CB<sub>1</sub> receptors are functionally active, as CB<sub>1</sub> receptor agonists (CP55,940 and WIN 55,212-2) applied topically lower intraocular pressure both in animals and humans, and their effect can be antagonized by SR141716 ([Pate et al., 1998](#); [Song and Slowey, 2000](#); [Porcella et al., 2001](#); [Chien et al., 2003](#); [Stumpff et al., 2005](#); reviewed in [Jarvinen et al., 2002](#)). The CB<sub>2</sub> receptor agonist JWH-133 did not modify the intraocular pressure, suggesting that CB<sub>2</sub> receptors may play only a minor, if any, role ([Laine et al., 2003](#)). CB<sub>1</sub> receptor signaling is also operational in the ocular trabecular meshwork ([Stumpff et al., 2005](#)), and ciliary muscle ([Lograno and Romano, 2004](#)). Third, endocannabinoids are detectable in

ocular tissues including the retina, ciliary body, and choroids plexus ([Bisogno et al., 1999b](#); [Straiker et al., 1999a,b](#); [Stamer et al., 2001](#); [Chen et al., 2005](#)), and the levels of anandamide and especially 2-AG are significantly decreased in patients with glaucoma ([Chen et al., 2005](#)).

The cellular/molecular mechanisms responsible for the intraocular pressure-reducing effect of cannabinoids are not completely understood but may involve direct effects on ciliary processes such as vasodilation and reduction of capillary pressure and secretion and do not seem to be related to systemic reduction of arterial blood pressure ([Green and Pederson, 1973](#); [Korzczyn, 1980](#)). Cannabinoids may also inhibit calcium influx through presynaptic ion channels, thereby reducing norepinephrine release in the ciliary body, resulting in decreased aqueous humor production ([Sugrue, 1997](#)). In addition, cannabinoids may improve the uveoscleral outflow by dilating blood vessels of the anterior uvea ([Porcella et al., 1998](#)), most likely by induction of several outflow-facilitating mediators ([Rosch et al., 2006](#)). Some evidence implicates prostanoids in the intraocular pressure-reducing effect of certain cannabinoids and anandamide ([Pate et al., 1996](#); [Green et al., 2001](#); [Rosch et al., 2006](#)).

Endocannabinoids as well as functional CB<sub>1</sub> receptors are present in the retina ([Bisogno et al., 1999b](#); [Straiker et al., 1999a,b](#); [Fan and Yazulla, 2003](#); [Savinainen and Laitinen, 2004](#)). Cannabinoids exert neuroprotective effects against retinal neurotoxicity ([El-Remessy et al., 2003](#)), and cannabidiol helps to preserve the blood-retinal barrier in experimental diabetes ([El-Remessy et al., 2006](#)). These effects could suggest their usefulness in various retinopathies. Unlike CB<sub>1</sub> receptors, CB<sub>2</sub> receptors were undetectable in human retina, although they were found in the rat retina ([Lu et al., 2000](#); [Porcella et al., 2000](#)).

Taken together, these findings indicate that cannabinoids may have great potential in the treatment of glaucoma, if the difficulty in formulating a stable and effective topical preparation and the problem of systemic side effects are conquered. Because of their well known neuroprotective, anti-inflammatory, and antiangiogenic effects, cannabinoid analogs may also be considered for the treatment of inflammatory and degenerative eye disorders and diabetic retinopathy.

## F. Cancer

The palliative effects of cannabinoids in cancer patients are well known and may include appetite stimulation, inhibition of nausea and emesis associated with chemo- or radiotherapy, pain relief, mood elevation, and relief from insomnia (reviewed in [Walsh et al., 2003](#); [Hall et al., 2005](#)) ([Table 1](#)).  $\Delta^9$ -THC (dronabinol, Marinol) and its synthetic derivative nabilone have been approved by the U.S. Food and Drug Administration to control nausea in cancer patients undergoing chemotherapy and to stimulate appetite in patients with AIDS ([Walsh et al., 2003](#); [Hall et al., 2005](#)).

Numerous recent studies have suggested that cannabinoids might directly inhibit cancer growth (reviewed in [Parolaro et al., 2002](#); [Guzmán et al., 2002](#); [Guzmán, 2003](#); [Jones and Howl, 2003](#); [Velasco et al., 2004](#); [Patsos et al., 2005](#)). The proposed mechanisms are complex and may involve induction of apoptosis in tumor cells, antiproliferative action, and an antimetastatic effect through inhibition of angiogenesis and tumor cell migration (reviewed in [Bifulco and Di Marzo, 2002](#); [Parolaro et al., 2002](#); [Guzmán et al., 2002](#); [Guzmán, 2003](#); [Jones and Howl, 2003](#); [Velasco et al., 2004](#); [Patsos et al., 2005](#)).

Various cannabinoids, including cannabidiol, anandamide, and 2-AG, and endocannabinoid transport inhibitors have been shown to induce apoptotic cell death and to inhibit proliferation and migration in numerous murine and human tumor cell lines including glioma (C6, U87, U373, and H4), oligodendroglioma (Gos3), glioblastoma multiforme, astrocytoma (U373-MG, U87MG, and human grade IV astrocytoma), neuroblastoma (N18 TG2 and CHP100), pheochromocytoma (PC12), breast cancer (MCF-7, EFM-19, T47D, TSA-E1, and MDA-MB-231), prostate cancer (LNCaP, DU145, and

PC3), colon carcinoma (SW 480), uterine cervix carcinoma (CxCa), thyroid cancer (KiMoI), leukemia (CEM, HEL-92, HL60, and Jurkat cell lines), and lymphoid tumors (EL-4 and P815) tumor cells via CB<sub>1</sub>/CB<sub>2</sub>- and VR<sub>1</sub> receptor-dependent or independent (e.g., cyclooxygenase) mechanisms ([De Petrocellis et al., 1998](#); [Sánchez et al., 1998, 2003](#); [Jacobsson et al., 2000](#); [Maccarrone et al., 2000b](#); [Sarker et al., 2000](#); [McKallip et al., 2002a,b](#); [Fowler et al., 2003](#); [Jonsson et al., 2003](#); [Mimeault et al., 2003](#); [Bifulco et al., 2004](#); [Contassot et al., 2004a,b](#); [Hinz et al., 2004](#); [Joseph et al., 2004](#); [Kogan et al., 2004](#); [Massi et al., 2004](#); [Nithipatikom et al., 2004](#); [Allister et al., 2005](#); [Ellert-Miklaszewska et al., 2005](#); [Herrera et al., 2005, 2006](#); [Lombard et al., 2005](#); [Powles et al., 2005](#); [Sarfaraz et al., 2005](#); [Vaccani et al., 2005](#); [Carracedo et al., 2006](#); [Grimaldi et al., 2006](#); [Ligresti et al., 2006b](#)). More importantly, systemic or local treatment with cannabinoids inhibited the growth of various types of tumor or tumor cell xenografts in vivo, including lung carcinoma ([Munson et al., 1975](#)), glioma ([Galve-Roperh et al., 2000](#); [Sánchez et al., 2001a](#); [Massi et al., 2004](#)), thyroid epithelioma ([Bifulco et al., 2001](#)), lymphoma ([McKallip et al., 2002a](#)), and skin carcinoma ([Casanova et al., 2003](#)) in mice.

The proapoptotic effect of cannabinoids in tumor cells is complex and may involve increased synthesis of the proapoptotic sphingolipid ceramide ([Galve-Roperh et al., 2000](#); [Gómez del Pulgar et al., 2002a,b](#)), ceramide-dependent up-regulation of the stress protein p8 and several downstream stress-related genes expressed in the endoplasmic reticulum (ATF-4, CHOP, and TRB3; [Carracedo et al., 2006](#)), prolonged activation of the Raf-1/mitogen-activated protein kinase/extracellular signal-regulated kinase signaling cascade ([Galve-Roperh et al., 2000](#)), and inhibition of Akt ([Gómez del Pulgar et al., 2000](#); [Ellert-Miklaszewska et al., 2005](#)), c-Jun NH<sub>2</sub>-terminal kinase and p38 mitogen-activated protein kinase ([Galve-Roperh et al., 2000](#); [Sarker et al., 2003](#); [Hinz et al., 2004](#); [Powles et al., 2005](#)). As mentioned above, cannabinoids also inhibit the proliferation of various tumor cells, possibly through inhibition of adenylyl cyclase and the cAMP/protein kinase A pathway ([Melck et al., 1999](#)), induction of the cyclin-dependent kinase inhibitor p27<sup>kip1</sup> ([Portella et al., 2003](#)), a decrease in epidermal growth factor receptor expression and/or the attenuation of epidermal growth factor receptor tyrosine kinase activity ([Casanova et al., 2003](#); [Mimeault et al., 2003](#)), and a decrease in the activity and/or expression of nerve growth factor or vascular endothelial growth factor tyrosine kinase receptors and prolactin ([De Petrocellis et al., 1998](#); [Melck et al., 2000](#); [Portella et al., 2003](#)). In addition to their proapoptotic and antiproliferative effects in tumor cells, cannabinoids also inhibit the expression of proangiogenic mediators or their receptors (e.g., vascular endothelial growth factor) and reduce vascular hyperplasia and cell migration, which play crucial roles in tumor growth and metastasis formation ([Blázquez 2004; et al., 2003, 2004](#); [Casanova et al., 2003](#); [Portella et al., 2003](#)).

In sharp contrast to the above, [Hart et al. \(2004\)](#) have demonstrated that treatment of lung cancer (NCI-H292), squamous cell carcinoma (SCC-9), bladder carcinoma (5637), glioblastoma (U373-MG), astrocytoma (1321N1), and kidney cancer (A498) cells with nanomolar concentrations of cannabinoids such as THC, anandamide, HU-210, and WIN 55,212-2 leads to rapid epidermal growth factor receptor- and metalloprotease-dependent cancer cell proliferation. However, the same study also documented that at micromolar concentrations cannabinoids induced cancer cell apoptosis, in agreement with previous reports ([Hart et al., 2004](#)). These results highlight the bimodal action of cannabinoids on cancer cell growth, with low concentrations being proliferative and high concentrations having antiproliferative effects.

The key role of the immune system in controlling the development of cancers is supported by findings that immunosuppressed individuals are at increased risk for developing cancer. For example, there is increased incidence of non-Hodgkin's lymphoma, Burkitt's lymphoma, Kaposi's sarcoma, and cervical cancer in AIDS patients and increased susceptibility to various lymphomas and solid tumors after organ transplantation ([Bhatia et al., 2001](#); [Scadden, 2003](#); [Abu-Elmagd et al., 2004](#); [Oruc et al., 2004](#)). This concept is particularly important, because cannabinoids have well-known immunosuppressant effects

(reviewed in [Klein, 2005](#)), which may compromise antitumor immune responses. Indeed, THC enhances breast and lung cancer growth and metastasis by suppressing CB<sub>2</sub> receptor-mediated antitumor immune responses ([Zhu et al., 2000](#); [McKallip et al., 2005](#)) and can also lead to increased susceptibility to infections with various pathogens such as herpes simplex virus, *Legionella pneumophila*, and Fried leukemia virus ([Morahan et al., 1979](#); [Cabral et al., 1986](#); [Specter et al., 1991](#); [Klein et al., 2000b](#)).

Epidemiological studies investigating the relationship of cannabis smoking and various forms of cancer have yielded inconsistent results, thus failing to resolve the conflicting findings in animal models of cancer or in cancer cell lines ([Taylor, 1988](#); [Caplan and Brigham, 1990](#); [Kuijten et al., 1992](#); [Grufferman et al., 1993](#); [Sidney et al., 1997](#); [Barsky et al., 1998](#); [Zhang et al., 1999](#); [Efird et al., 2004](#); [Llewellyn et al., 2004](#); [Rosenblatt et al., 2004](#); reviewed in [Hall et al., 2005](#)). The variability of the effects of cannabinoids in different tumor models may be related to the differential expression of CB<sub>1</sub> and CB<sub>2</sub> receptors. Thus, cannabinoids may be effective in killing tumors that abundantly express cannabinoid receptors, such as gliomas, but may increase the growth and metastasis of other types of tumors, such as breast cancer, with no or low expression of cannabinoid receptors, due to the suppression of the antitumor immune response ([McKallip et al., 2005](#)). Nevertheless, the majority of the findings to date are encouraging and suggest that cannabinoids may be useful not only as palliative therapy but also because of their ability to inhibit tumor growth and metastasis.

## G. Gastrointestinal and Liver Disorders

Cannabis has been used empirically for centuries to stimulate appetite and decrease emesis and diarrhea. Recent evidence indicates that the endocannabinoid system plays an important role in the control of gastrointestinal motility and secretion both under physiological conditions and in various gastrointestinal disorders (reviewed in [Pertwee, 2001](#); [Pinto et al., 2002a,b](#); [Di Carlo and Izzo, 2003](#); [Coutts and Izzo, 2004](#); [Duncan et al., 2005](#); [Massa et al., 2005](#)). Unexpectedly, recent data also implicate endocannabinoids and their receptors in several aspects of acute and chronic liver disease, including hemodynamic changes, modulation of inflammatory processes, fibrosis, and altered brain function (reviewed in [Gabbay et al., 2005](#); [Jimenez, 2005](#)).

Numerous studies using autoradiography, immunohistochemistry, and/or reverse transcription-polymerase chain reaction demonstrated colocalization of CB<sub>1</sub> receptors with cholinergic neurons across the enteric nervous system, including sensory and interneuronal as well as motoneuronal cell bodies of the myenteric plexus, in mice ([Mascolo et al., 2002](#); [Pinto et al., 2002a,b](#); [Casu et al., 2003](#); [Izzo et al., 2003](#); [Storr et al., 2004](#)), rats ([Adami et al., 2002](#); [Coutts et al., 2002](#); [Storr et al., 2002](#); [Burdyga et al., 2004](#)), guinea-pigs ([Coutts et al., 2002](#); [MacNaughton et al., 2004](#)), and pigs ([Kulkarni-Narla and Brown, 2000](#)). CB<sub>1</sub> receptors are also colocalized with neuropeptide Y and vasoactive intestinal peptide in a small population of submucous plexus neurons ([Kulkarni-Narla and Brown, 2000](#); [Coutts et al., 2002](#)). CB<sub>1</sub> receptor immunoreactivity was evident in normal human colonic epithelium, smooth muscle, and the sub-mucosal myenteric plexus ([Wright et al., 2005](#)). Both CB<sub>1</sub> and CB<sub>2</sub> receptors were found on plasma cells in the lamina propria, whereas only CB<sub>2</sub> were detectable on macrophages ([Wright et al., 2005](#)). Endocannabinoids are also present in the gastrointestinal tract. Indeed, 2-AG was originally isolated from gut tissue ([Mechoulam et al., 1995](#)), and the intestinal content of anandamide was found to be regulated by feeding status ([Gomez et al., 2002](#)).

Although in earlier studies CB<sub>1</sub> receptor expression was undetectable in the liver relative to the brain ([Porcella et al., 2002](#)), several recent studies revealed the presence of low levels of both CB<sub>1</sub> mRNA ([Bátkai et al., 2001](#); [Michalopoulos et al., 2003](#); [Biecker et al., 2004](#); [Engeli et al., 2005](#); [Osei-Hyiaman et al., 2005b](#); [Teixeira-Clerc et al., 2006](#)) and CB<sub>1</sub> immunoreactivity ([Osei-Hyiaman et al., 2005b](#)) in whole liver or in various types of cells present in the liver, including hepatocytes ([Michalopoulos et al.,](#)

2003; Osei-Hyiaman et al., 2005b), stellate cells (Siegmund et al., 2005; Teixeira-Clerc et al., 2006), and vascular endothelial cells (Bátkai et al., 2001). CB<sub>2</sub> receptor mRNA was also detected in cirrhotic but not in normal liver tissue (Julien et al., 2005). Endocannabinoids are detectable in the liver or liver cells both in animals and humans at levels similar to those in the brain and play an important role under various physiological and pathophysiological conditions (Cravatt et al., 2004; Kurabayashi et al., 2005; Osei-Hyiaman et al., 2005b) (see also section III.A.3.).

A functional role for endocannabinoids and CB<sub>1</sub> receptors in the gastrointestinal tract is supported by pharmacological studies demonstrating that anandamide and various CB<sub>1</sub> agonists (WIN 55,212-2, CP55,940, and ACEA) but not the CB<sub>2</sub>-selective agonists JWH-133 inhibit gastrointestinal motility in rodents in vivo and in isolated ileum and colon from both experimental animals and humans (Shook and Burks, 1989; Pertwee et al., 1995, 1996; Coutts and Pertwee, 1997; McCallum et al., 1999; Mancinelli et al., 2001; Mang et al., 2001; Landi et al., 2002; Manara et al., 2002; Hinds et al., 2006). A similar role for endogenous substrates of FAAH is suggested by recent in vivo findings in mice, documenting inhibition of intestinal motility by the FAAH inhibitors *N*-arachidonoylserotonin and palmitoylisopropylamide and by the FAAH substrates palmitoylethanolamide, oleamide, and oleoylethanolamide in wild-type but not in FAAH knockout mice (Capasso et al., 2005). Furthermore, the effect of *N*-arachidonoylserotonin was reduced either by the CB<sub>1</sub> receptor antagonist SR141716 or by CB<sub>1</sub> deficiency, but not by the TRPV<sub>1</sub> receptor antagonist 5'-iodoresiniferatoxin (Capasso et al., 2005). Interestingly, in clinical trials using rimonabant for nicotine cessation or for the treatment of obesity, diarrhea was 2 to 2.4 times more frequent among subjects treated with the drug than with placebo, suggesting accelerated transit and/or enhanced secretion caused by CB<sub>1</sub> blockade (Fernandez and Allison, 2004; Van Gaal et al., 2005). This and some of the above experimental reports suggest the existence of an inhibitory endocannabinoid tone in the gastrointestinal tract. Multiple mechanisms, including reduction of acetylcholine release from enteric nerves, inhibition of nonadrenergic/noncholinergic excitatory transmission, activation of apamin-sensitive K<sup>+</sup> channels, and modulation of adenosine release have been proposed to explain the CB<sub>1</sub>-mediated reduction in enteric contractility and peristalsis (reviewed in Coutts and Izzo, 2004).

Activation of both CB<sub>1</sub> and CB<sub>2</sub> receptors may decrease the pathologically increased intestinal motility elicited by an inflammatory stimulus. In a mouse model of croton oil-induced intestinal inflammation, the increased efficacy of cannabinoids in inhibiting intestinal motility was attributed to up-regulation of intestinal CB<sub>1</sub> receptors (Izzo et al., 2001a,b). Conversely, the accelerated gastrointestinal transit induced by bacterial endotoxin in rats could be inhibited by CB<sub>2</sub> but not CB<sub>1</sub> receptor agonists (Mathison et al., 2004). Interestingly, intestinal hypomotility in a mouse model of paralytic ileus has been linked, at least in part, to the enhancement of anandamide levels and CB<sub>1</sub> expression in the gut, and it could be attenuated by CB<sub>1</sub> receptor antagonists (Mascolo et al., 2002). Additionally, there is evidence that CB<sub>1</sub> receptors are involved in the regulation of the lower esophageal sphincter, and CB<sub>1</sub> activation might be beneficial in gastroesophageal reflux disease (reviewed in Coutts and Izzo, 2004; Massa et al., 2005).

The endocannabinoid system has also been implicated in the regulation of gastric acid and intestinal secretions. At high doses, THC decreased histamine-induced gastric acid secretion in isolated stomach preparations (Rivas-V and Garcia, 1980) and in pylorus-ligated rats (Sofia et al., 1978). Pentagastrin-induced gastric acid secretion was also inhibited by HU-210 and WIN 55,212-2, an effect that could be prevented by CB<sub>1</sub> blockade (Coruzzi et al., 1999; Adami et al., 2002). These studies suggest a role for CB<sub>1</sub> receptors located on preganglionic and postganglionic cholinergic pathways in the regulation of gastric acid secretion. The therapeutic relevance of this regulatory mechanism was highlighted by the CB<sub>1</sub> receptor-mediated antiulcer activity of ACEA or WIN 55,212-2 treatment in a rat model of aspirin-

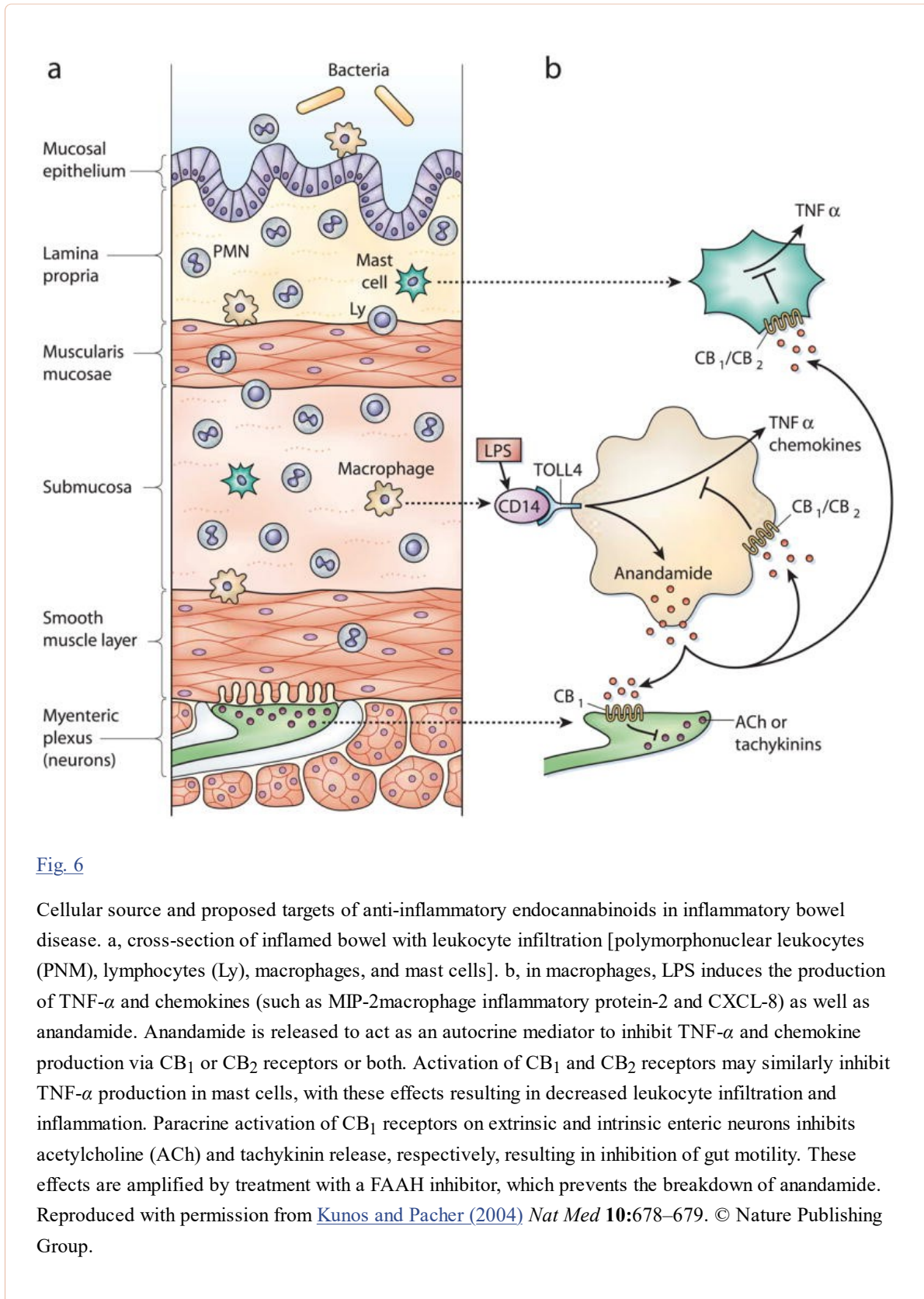
and cold/restraint stress-induced gastric ulcers ([Germano et al., 2001](#); [Rutkowska and Fereniec-Goltbiewska, 2006](#)). WIN 55,212-2 also reduced intestinal secretions evoked by electrical field stimulation or capsaicin ([MacNaughton et al., 2004](#)). Anandamide, the anandamide transport inhibitor VDM11, and the CB<sub>1</sub> agonist ACEA all inhibited intestinal secretion and fluid accumulation in mice treated with cholera toxin, whereas SR141716 exerted opposite effects ([Izzo et al., 2003](#)). The ability of cannabinoids to inhibit gastrointestinal motility and secretion coupled with their anti-inflammatory properties strongly suggests that the modulation of this system could offer significant benefits in the treatment of various gastrointestinal pathological conditions, including inflammatory bowel disease (see below).

**1. Inflammatory Bowel Disease** Idiopathic inflammatory bowel disease (IBD) includes ulcerative colitis and Crohn's disease, and is characterized by intestinal inflammation presumably of autoimmune origin and a chronic relapsing course associated with local and systemic complications and affects >1 million people in the United States ([Loftus, 2004](#)). Although the etiology of IBD remains unclear, it may involve complex genetic, environmental, and immunological interactions. The most common symptoms of IBD are abdominal pain and diarrhea, which eventually lead to malabsorption and malnutrition, and in approximately half of patients surgery is eventually required to remove the affected bowel segment. Despite recent therapeutic advances, patients with IBD are often unresponsive to available treatment options.

As discussed above, the endocannabinoid system plays an important role in the control of gastrointestinal motility and secretion. Studies using animal models of IBD have suggested that targeting the endocannabinoid system may offer significant benefits in the treatment of IBD. Several studies have indicated that chemically induced intestinal inflammation is associated with the up-regulation of intestinal CB<sub>1</sub> receptors, which may represent a compensatory, protective mechanism. For example, in croton oil-treated mice, the ability of CB<sub>1</sub> agonists to inhibit intestinal motility is increased compared with that in control animals ([Izzo et al., 2001a](#)). More importantly, the anandamide transport inhibitor VDM11 was also shown to inhibit gastrointestinal motility and secretions in cholera toxin-treated mice, which implicates endocannabinoids in this action and holds out the promise of a nonpsychoactive form of treatment ([Izzo et al., 2003](#)). In a mouse model of colitis induced by 2,4-dinitrobenzene sulfonic acid and dextrane sulfate, [Massa et al. \(2004\)](#) have confirmed the up-regulation of CB<sub>1</sub> receptors in experimental colitis. Furthermore, they demonstrated that the inflammation was more severe in mice deficient in CB<sub>1</sub> receptors than in wild-type mice, whereas genetic ablation of FAAH resulted in protection against this chemically induced colitis ([Massa et al., 2004](#)). In a recent study, the anandamide reuptake inhibitor VDM11 afforded protection against colitis in mice, and elevated anandamide levels have been measured in biopsy material from patients with ulcerative colitis ([D'Argenio et al., 2006](#)). These findings strongly support the natural protective role of the endocannabinoid system in this form of experimental IBD. In contrast, [Croci et al. \(2003\)](#) have reported a CB<sub>1</sub> receptor-independent protective effect of SR141716 against indomethacin-induced inflammation and ulcer formation in the small intestine of rats. Elevated levels of anandamide and desensitization of the presynaptic neural CB<sub>1</sub> receptor found in colonic longitudinal muscle strips from patients undergoing surgery for complicated diverticulitis suggest that the endocannabinoid system may be also involved in the pathophysiology of this frequent complication of colitis and/or colon cancer ([Guagnini et al., 2006](#)).

Taken together, most of the above studies suggest that the endocannabinoid system in the gut is activated during inflammation, and endogenous anandamide may counteract inflammation ([Kunos and Pacher, 2004](#)) (Fig. 6). The findings of [Massa et al. \(2004\)](#) and [D'Argenio et al. \(2006\)](#) also suggest that inhibitors of FAAH or anandamide reuptake may amplify the natural protective action of endogenous anandamide, which warrants further studies to test such inhibitors in the treatment of experimental and, ultimately, human IBD ([Kunos and Pacher, 2004](#)). Future studies should further explore the mechanisms

of the anti-inflammatory effects of cannabinoids and the potential role of CB<sub>2</sub> receptors as therapeutic targets (Mathison et al., 2004; Wright et al., 2005).



**Fig. 6**

Cellular source and proposed targets of anti-inflammatory endocannabinoids in inflammatory bowel disease. **a**, cross-section of inflamed bowel with leukocyte infiltration [polymorphonuclear leukocytes (PNM), lymphocytes (Ly), macrophages, and mast cells]. **b**, in macrophages, LPS induces the production of TNF- $\alpha$  and chemokines (such as MIP-2macrophage inflammatory protein-2 and CXCL-8) as well as anandamide. Anandamide is released to act as an autocrine mediator to inhibit TNF- $\alpha$  and chemokine production via CB<sub>1</sub> or CB<sub>2</sub> receptors or both. Activation of CB<sub>1</sub> and CB<sub>2</sub> receptors may similarly inhibit TNF- $\alpha$  production in mast cells, with these effects resulting in decreased leukocyte infiltration and inflammation. Paracrine activation of CB<sub>1</sub> receptors on extrinsic and intrinsic enteric neurons inhibits acetylcholine (ACh) and tachykinin release, respectively, resulting in inhibition of gut motility. These effects are amplified by treatment with a FAAH inhibitor, which prevents the breakdown of anandamide. Reproduced with permission from [Kunos and Pacher \(2004\)](#) *Nat Med* **10**:678–679. © Nature Publishing Group.

**2. Acute and Chronic Liver Disease (Hepatitis and Liver Cirrhosis)** Endocannabinoids and CB<sub>1</sub> receptors have been implicated in the systemic and portal vasodilation and hypotension associated with

chronic liver cirrhosis ([Bátkai et al., 2001](#); [Garcia et al., 2001](#); [Ros et al., 2002](#)). These studies demonstrated that CB<sub>1</sub> receptor blockade with SR141716 reversed the hypotension and low peripheral resistance and decreased the elevated mesenteric blood flow and portal pressure in rats with biliary and carbon tetrachloride-induced cirrhosis, whereas these hemodynamic parameters were unaffected by SR141716 in noncirrhotic control subjects ([Bátkai et al., 2001](#); [Ros et al., 2002](#)). These findings suggested an increased endocannabinoid tone in cirrhosis, which could be attributed to both an up-regulation of CB<sub>1</sub> receptors in hepatic vascular endothelial cells and an increased production of anandamide by circulating monocytes ([Bátkai et al., 2001](#)). Increased expression of CB<sub>1</sub> receptors was also reported in whole liver from bile duct-ligated mice ([Biecker et al., 2004](#)). This increase was greater when bile duct ligation was performed in NO synthase-3 knockout compared with wild-type mice, which may account for the similar level of portal hypertension in the two strains despite the much higher systemic blood pressure in the knockout mice ([Biecker et al., 2004](#)). Increased anandamide-induced vasorelaxation mediated by CB<sub>1</sub> and TRPV<sub>1</sub> receptors was also reported in mesenteric arteries isolated from cirrhotic compared with control rats ([Domenicali et al., 2005](#)). The increase in anandamide in monocytes from cirrhotic rats or humans is functionally important, as these cells elicit SR141716-sensitive hypotension when injected into normal recipient rats ([Bátkai et al., 2001](#); [Ros et al., 2002](#)). Plasma endotoxin levels progressively increase as liver function deteriorates in cirrhosis ([Lumsden et al., 1988](#); [Chan et al., 1997](#)), and this effect is probably responsible for the elevated endocannabinoid production in plasma monocytes and platelets of cirrhotic animals and patients ([Bátkai et al., 2001](#); [Ros et al., 2002](#); [Liu et al., 2003](#); [Fernandez-Rodriguez et al., 2004](#)). There is also recent experimental evidence implicating increased signaling through myocardial CB<sub>1</sub> receptors in the pathogenesis of cirrhotic cardiomyopathy ([Gaskari et al., 2005](#); [Pacher et al., 2005c](#)).

Beyond the vasculopathy of end-stage cirrhosis, the endocannabinoid system may also be involved in the pathogenesis of liver fibrosis. [Siegmund et al. \(2005\)](#) have recently reported that anandamide exerts antifibrogenic effects in vitro by inhibiting activated hepatic stellate cells at low micromolar concentrations and by inducing their necrosis at higher concentrations, via CB<sub>1/2</sub>- and TRPV<sub>1</sub>-independent mechanism(s). In a study by [Julien et al. \(2005\)](#), the liver fibrosis induced by carbon tetrachloride was more severe in CB<sub>2</sub> knockout mice compared with their wild-type littermates. Also, the expression of CB<sub>2</sub> receptors was found to be strongly induced in liver biopsy specimens from patients with active cirrhosis of various etiologies, particularly in non-parenchymal cells located within and at the edge of fibrous septa ([Julien et al., 2005](#)). Furthermore, CB<sub>2</sub> receptor activation triggered growth inhibition and apoptosis in myofibroblasts and in activated hepatic stellate cells, highlighting the antifibrogenic role of CB<sub>2</sub> receptors during chronic liver injury ([Julien et al., 2005](#)). However, chronic marijuana use has been associated with hepatotoxicity rather than hepatoprotection as expected from the above results ([Borini et al., 2004](#)), and results of a recent epidemiological study indicate that daily marijuana smoking is a risk factor for progression of fibrosis among people with chronic hepatitis C infection ([Hezode et al., 2005](#)). This finding has triggered an investigation into the possible profibrogenic role of CB<sub>1</sub> receptor activation, which is supported by the results of a preliminary study showing that the progression of experimental liver fibrosis induced by carbon tetrachloride is slower in mice with genetic ablation of CB<sub>1</sub> receptors or treated with CB<sub>1</sub> receptor antagonist SR141716 ([Teixeira-Clerc et al., 2006](#)). These latter findings suggest a broader role of CB<sub>1</sub> receptors in the pathogenesis of cirrhosis and forecast additional potential benefits from the therapeutic use of a CB<sub>1</sub> antagonist in chronic liver disease.

In contrast to the hepatotoxicity associated with chronic marijuana use, a synthetic, nonpsychotropic cannabinoid derivative (PRS-211,092) was reported to inhibit acute hepatitis induced by concanavalin A via negative cytokine regulation in mice ([Lavon et al., 2003](#)). Interestingly, in animal models of acute hepatic failure-induced encephalopathy, both 2-AG and SR141716 have been reported to exert



beneficial effects on neurological and cognitive function ([Gabbay et al., 2005](#); [Avraham et al., 2006](#)). Cannabinoids may also be beneficial in intractable cholestatic pruritus ([Neff et al., 2002](#)), which is associated with severe forms of liver disease, presumably by increasing the nociceptive threshold ([Gingold and Bergasa, 2003](#)).

Collectively, the studies discussed in this section highlight the potential regulatory role of the endocannabinoid system in a variety of gastrointestinal and liver disorders, opening new avenues for their pharmacotherapy. It appears that CB<sub>1</sub> agonists and perhaps FAAH antagonists might be beneficial in reducing increased gastrointestinal motility, bowel inflammation, and associated diarrhea, whereas CB<sub>1</sub> antagonists could be used in the treatment of constipation. In chronic liver cirrhosis, CB<sub>1</sub> antagonists may not only attenuate or reverse the adverse hemodynamic consequences of cirrhosis, thus extending life until a suitable liver becomes available for transplantation, but also could have additional benefits by slowing the progression of fibrosis and the neurological decline associated with hepatic encephalopathy. Selective CB<sub>2</sub> receptor agonists might also be expected to protect against progression of liver fibrosis and perhaps against the chronic inflammation associated with IBD.

## H. Musculoskeletal Disorders

**1. Arthritis** Rheumatoid arthritis (RA) is a chronic, systemic, inflammatory autoimmune disease affecting ~0.8% of adults worldwide. RA is more common in women, and it leads to joint destruction, deformity, loss of function, chronic pain, and reduced quality of life. When unchecked, it leads to substantial disability and premature death ([O'Dell, 2004](#)). Current medications used to treat rheumatoid arthritis are divided into three main classes: nonsteroidal anti-inflammatory drugs, corticosteroids, and disease-modifying antirheumatic drugs such as methotrexate ([O'Dell, 2004](#)). A better understanding of the cytokine networks that are responsible for the ongoing inflammatory response in RA has led to the successful use of novel therapies that target TNF- $\alpha$  and IL-1.

The immunosuppressant and anti-inflammatory properties of cannabinoids are highly relevant for RA and other autoimmune disorders (e.g., systemic lupus erythematosus, autoimmune vasculitis, Sjögren's syndrome, and ankylosing spondylitis). Indeed, ajulemic acid (THC-11-oic acid, CT-3, IP-751), a potent analog of the acid metabolites of THC ([Burstein, 2000, 2005](#)) and cannabidiol have been shown to have analgesic, anti-inflammatory, and immunosuppressive effects in animal models of arthritis ([Zurier et al., 1998](#); [Dajani et al., 1999](#); [Malfait et al., 2000](#)). Chronic administration of ajulemic acid attenuated joint inflammation in a murine model of adjuvant-induced arthritis and suppressed prostaglandin production in vitro to a greater extent than the potent nonsteroidal anti-inflammatory drug, indomethacin ([Zurier et al., 1998](#)). In another study, ajulemic acid caused less gastrointestinal ulcerations and was more effective in reducing adjuvant-induced arthritis than common nonsteroidal anti-inflammatory agents ([Dajani et al., 1999](#)). As discussed earlier in this review, ajulemic acid is a high-affinity agonist for human cannabinoid receptors and has CB<sub>1</sub>-mediated, potent antihyperalgesic activity in models of chronic neuropathic and inflammatory pain in the rat ([Dyson et al., 2005](#)). Ajulemic acid also induces apoptosis in human T lymphocytes ([Bidinger et al., 2003](#)) and suppresses IL-1 $\beta$  production in human monocytes ([Zurier et al., 2003](#)), which could contribute to its therapeutic effects in RA and other inflammatory disorders. Treatment with cannabidiol or its more potent dimethylheptyl derivative (HU-320) reduced an LPS-induced increase in serum TNF- $\alpha$  and immune function and effectively blocked the progression of collagen-induced arthritis in mice ([Malfait et al., 2000](#); [Sumariwalla et al., 2004](#)). Other studies described the antinociceptive effects of anandamide and THC in rats with arthritis ([Sofia et al., 1973](#); [Smith et al., 1998](#); [Cox and Welch, 2004](#)). [Mbvundula et al. \(2005, 2006\)](#) have recently reported that WIN 55,212-2 and HU-210 inhibited IL-1-stimulated NO production in bovine articular chondrocytes, in contrast to AM281 and AM630, which elicited an opposite effect. Anandamide, WIN 55212-2, and HU-210 also inhibited the release of sulfated glycosaminoglycans in bovine cartilage explants and IL-1 $\alpha$

stimulated proteoglycan and collagen degradation ([Mbvundula et al., 2005, 2006](#)).

In a survey of 2969 people using cannabis for medicinal purposes, ~25% of subjects mentioned relief of arthritis symptoms as the main reason for cannabis smoking, which was surpassed only by chronic pain, MS, and depression ([Ware et al., 2003](#)). Studies using cannabinoid-based extracts are also underway in patients with RA ([Russo, 2006](#)). The potential benefit of cannabinoids in fibromyalgia, a syndrome of widespread musculoskeletal pain, nonrestorative sleep, disturbed mood, and fatigue of unknown etiology, has also been reviewed ([Russo, 2004](#)).

**2. Osteoporosis** Osteoporosis is a skeletal disorder characterized by low bone mass and microarchitectural deterioration of bone, leading to increased susceptibility to bone fractures. The associated fractures and the subsequent morbidity and mortality make osteoporosis an enormous public health concern. Osteoporosis is no longer considered an age-related disease, as it is increasingly recognized in children. Osteoporosis is thought to be a polygenic disorder, with vulnerability determined by multiple genes and environmental risk factors. It currently affects up to one in three women and 1 in 12 men worldwide ([Keen, 2003](#)). Treatment options include general measures on lifestyle, calcium and vitamin D supplements, hormone therapy, raloxifene, and bisphosphonates.

Cannabinoid receptors were first implicated in the regulation of bone mass by [Karsak et al. \(2004\)](#), who found that CB<sub>2</sub> knockout mice had markedly accelerated age-related trabecular bone loss and cortical expansion accompanied by increased activity of trabecular osteoblasts, increased numbers of osteoclasts, and decreased numbers of diaphyseal osteoblast precursors ([Ofek et al., 2006](#)). CB<sub>2</sub> receptors were expressed in osteoblasts, osteocytes, and osteoclasts. The selective CB<sub>2</sub> agonist HU-308, but not the CB<sub>1</sub> agonist noladine ether, attenuated ovariectomy-induced bone loss and markedly stimulated cortical thickness through the suppression of osteoclast number and stimulation of endocortical bone formation ([Ofek et al., 2006](#)). Furthermore, HU-308 dose dependently increased the number and activity of endocortical osteoblasts and restrained trabecular osteoclastogenesis by inhibiting proliferation of osteoclast precursors ([Ofek et al., 2006](#)). These results, coupled with CB<sub>2</sub> but not CB<sub>1</sub> receptor mRNA expression during osteoblastic differentiation, suggested a role for CB<sub>2</sub> receptors in bone remodeling. Such a role of CB<sub>2</sub> but not CB<sub>1</sub> receptors is also supported by a recent genetic association study in human samples of postmenopausal osteoporosis patients and matched female control subjects ([Karsak et al., 2005](#)).

In contrast, [Idris et al. \(2005\)](#) have recently reported that CB<sub>1</sub> receptor knockout mice or mice treated with antagonists of either CB<sub>1</sub> or CB<sub>2</sub> receptors were protected from ovariectomy-induced bone loss. Furthermore, cannabinoid antagonists promoted osteoclast apoptosis, inhibited osteoclast activity, and decreased the production of several osteoclast survival factors in vitro, suggesting that cannabinoid antagonists may be beneficial in the treatment of osteoporosis. Although the reason for the discrepancy between the above studies is not clear; they suggest a role for the endocannabinoid system in the regulation of bone mass.

## I. Endocannabinoids and Reproductive Functions

There is abundant evidence that the endocannabinoid system is involved in reproductive functions in both males and females and in both animals and humans, as discussed in more detail in recent reviews ([Fride, 2004](#); [Park et al., 2004](#); [Schuel and Burkman, 2005](#); [Tranguch et al., 2005](#); [Wang et al., 2006](#)). In males, marijuana, synthetic cannabinoids, and anandamide adversely affect the fertilizing capacity of sperm, which express functional CB<sub>1</sub> receptors ([Rossato et al., 2005](#); [Schuel and Burkman, 2005](#); [Whan et al., 2006](#)). On the other hand, there is preclinical evidence to suggest that blockade of CB<sub>1</sub> may be useful in the treatment of erectile dysfunction ([Melis et al., 2004b, 2006](#)).

High levels of functional CB<sub>1</sub> receptor, anandamide, and FAAH are present in the preimplantation embryo and/or in the uterus (Das et al., 1995; Paria et al., 1995, 2001; Schmid et al., 1997; Park et al., 2003; Guo et al., 2005). Anandamide synthesized in the uterus exerts dose- and stage-specific effects on embryo development and implantation. A temporary reduction of anandamide levels is essential for embryo implantation, and higher anandamide levels are associated with uterine nonreceptivity and impairment of blastocyst formation, zona hatching, and trophoblast outgrowth via CB<sub>1</sub> receptors (Das et al., 1995; Paria et al., 1995, 2001, 2002; Schmid et al., 1997; Wang et al., 1999; Guo et al., 2005). Consequently, cannabinoids may retard the development of embryos, eventually leading to fetal loss and pregnancy failure (Bloch et al., 1978; Smith and Asch, 1987; Park et al., 2004). Anandamide levels in the uterus are regulated by FAAH activity (Paria et al., 1995, 1999; Schmid et al., 1997). Accordingly, pregnant women with low FAAH activity in lymphocytes were found to have an increased incidence of miscarriage (Maccarrone et al., 2000c), and low FAAH activity also correlated with failure to maintain pregnancy after in vitro fertilization (Maccarrone et al., 2002b). Finally, cannabinoids may also affect the levels of various hormones crucial for normal fertility and reproduction (Brown and Dobs, 2002; Park et al., 2004; Scorticati et al., 2004; Gammon et al., 2005). Although such findings may suggest the potential usefulness of CB<sub>1</sub> antagonists in the treatment of infertility problems, a note of caution is warranted because CB<sub>1</sub> knockout mice were reported to have impaired oviductal transport of embryos, leading to embryo retention. This suggests that treatment with CB<sub>1</sub> antagonists may facilitate ectopic pregnancy (Wang et al., 2004).

#### IV. Future Directions

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The length of this review, necessitated by the steady growth in the number of indications for the potential therapeutic use of cannabinoid-related medications, is a clear sign of the emerging importance of this field. This is further underlined by the quantity of articles in the public database dealing with the biology of cannabinoids, which numbered ~200 to 300/year throughout the 1970s to reach an astonishing 5900 in 2004. The growing interest in the underlying science has been matched by a growth in the number of cannabinoid drugs in pharmaceutical development from two in 1995 to 27 in 2004, with the most actively pursued therapeutic targets being pain, obesity, and multiple sclerosis (Hensen, 2005). As in any rapidly growing area of research, not all the leads will turn out to be useful or even valid. Nevertheless, it is safe to predict that new therapeutic agents that affect the activity of the endocannabinoid system will emerge and become members of our therapeutic armamentarium. The plant-derived cannabinoid preparation Sativex has already gained regulatory approval in Canada for the treatment of spasticity and pain associated with multiple sclerosis, and the CB<sub>1</sub> receptor antagonist rimonabant has been approved in Europe and is awaiting Food and Drug Administration approval in the United States for the treatment of the metabolic syndrome. Undoubtedly, these will be followed by new and improved compounds aimed at the same or additional targets in the endocannabinoid system. However, it may be only after the widespread therapeutic use of such compounds that some important side effects will emerge. Although this occurrence would be undesirable from a health care perspective, such side effects may shed further light on the biological functions of endocannabinoids in health and disease.

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## Footnotes

<sup>1</sup>Abbreviations: THC or  $\Delta^9$ -THC,  $\Delta^9$ -tetrahydrocannabinol; CP-55,940, (1*R*,3*R*,4*R*)-3-[2-hydroxy-4-(1,1-dimethylheptyl)phenyl]-4-(3-hydroxypropyl)cyclohexan-1-ol; GPCR, G protein-coupled receptor; CB<sub>1</sub> or CB<sub>2</sub>, cannabinoid 1 or 2; CBD, cannabidiol; SR141716, *N*-(piperidin-1-yl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1*H*-pyrazole-3-carboximide hydrochloride (rimonabant); AM251, *N*-(piperin-1-yl)-5-(4-iodophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1*H*-pyrazole-3-carboxamide; TRPV<sub>1</sub> or VR<sub>1</sub>, transient receptor potential vanilloid 1 or vanilloid 1; WIN 55,212-2, *R*-(+)-[2,3-dihydro-5-methyl-3-[(morpholinyl)methyl]pyrrolo-[1,2,3-*de*]-1,4-benzoxazinyl]- (1-naphthalenyl)methanone mesylate; GTP $\gamma$ S, guanosine 5'-*O*-(3-thio)triphosphate; HU-210,  $\Delta^8$ -tetrahydrocannabinol dimethyl heptyl; DARPP-32, dopamine- and cAMP-regulated phosphoprotein of 32 kDa; 2-AG, 2-arachidonoylglycerol; NAPE, *N*-arachidonoyl phosphatidylethanolamide; PE, phosphatidylethanolamine; PL, phospholipase; DAG, diacylglycerol; FAAH, fatty acid amide hydrolase; UCM707, *N*-(3-furanylmethyl)-5*Z*,8*Z*,11*Z*,14*Z*-eicosatetraenamide; LY2318912, 5-(4-azido-3-iodo-benzoylaminoethyl)-tetrazole-1-carboxylic acid dimethylamide; MGL, monoacylglyceride lipase; DSI, depolarization-induced suppression of inhibition; SR144528, *N*-((1*S*)-endo-1,3,3-trimethyl bicyclo heptan-2-yl)-5-(4chloro-3-methylphenyl)-1-(4-methylbenzyl)-pyrazole-3-carboxamide); NPY, neuropeptide Y; MCH, melanin concentrating hormone;  $\alpha$ -MSH,  $\alpha$ -melanocyte-stimulating hormone; CRH, corticotropin-releasing hormone; CART, cocaine- and amphetamine-related transcript; AMPK, AMP-activated protein kinase; ACC1, acetyl CoA carboxylase-1; SREBP1c, sterol response element binding protein 1c; HDL, high-density lipoprotein; LDL, low-density lipoprotein; CNS, central nervous system; HIV, human immunodeficiency virus; LPS, lipopolysaccharide or endotoxin; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; IL, interleukin; CXCL, CXC chemokine ligand; NMDA receptor, *N*-methyl-D-aspartate receptor; HU-211, dexanabinol; TBI, traumatic brain injury; BAY 38-7271, (-)-*R*-3-(2-hydroxymethylindanyl-4-oxy)phenyl-4,4,4-trifluoro-1-sulfonate; MCAo, middle cerebral artery occlusion; GABA, gamma-aminobutyric acid; GPe or GPi, external or internal globus pallidus; HD, Huntington's disease; HPA axis, hypothalamic-pituitary-adrenal axis; HU-211, dexanabinol; ICAM-1, intercellular adhesion molecule-1; IL, interleukin; I/R, ischemia reperfusion; KA, kainic acid; LID, levodopa-induced dyskinesia; methyl-D-aspartate receptor; NO, nitric oxide; PD, Parkinson's disease; [LY320135](#), [6-methoxy-2-(4-methoxyphenyl)benzo[*b*]-thien-3-yl][4-cyanophenyl] methanone; MS, multiple sclerosis; SCI, spinal cord injury; EAE, experimental autoimmune encephalomyelitis; JWH-133, 1,1-dimethylbutyl-1-deoxy- $\Delta^9$ -tetrahydrocannabinol; PEA, palmitoylethanolamide; ACEA, arachidonyl-2'-chloroethylamide/(all *Z*)-*N*-(2-cycloethyl)-5,8,11,14-eicosatetraenamide; JWH-015, (2-methyl-1-propyl-1*H*-indol-3-yl)-1-naphthalenylmethanone; OM-DM1, (*R*)-*N*-oleoyl-(1'-hydroxybenzyl)-2'-ethanolamine; OMDM2, (*S*)-*N*-oleoyl-(1'-hydroxybenzyl)-2'-ethanolamine; SNr, substantia nigra pars reticulata; LID, levodopa-induced dyskinesia; GPe or GPi, external or internal globus pallidus; HD, Huntington's disease; ALS, amyotrophic lateral sclerosis; AM404, *N*-(4-hydroxyphenyl)-eicosa-5,8,11,14-tetraenamide; VDM11, *N*-(4-hydroxy-2-methylphenyl) arachidonoyl amide; AM374, palmitylsulfonyl fluoride; TS, Gilles de la Tourette's syndrome; AD, Alzheimer's disease; A $\beta$ ,  $\beta$ -amyloid; HPA, hypothalamic-pituitary-adrenal; URB597, cyclohexyl carbamic acid 3'-carbamoyle-biphenyl-3-yl ester; 5-HT, 5-hydroxytryptamine (serotonin); VTA, ventral tegmental area; nAc, nucleus accumbens; CPP, conditioned place preference; MDMA, 3,4-methylenedioxyamphetamine (Ecstasy); SHR, spontaneously hypertensive rat(s); WKY, Wistar-Kyoto; AM281, *N*-(morpholin-4-yl)-1-(2,4-dichlorophenyl)-5-(4-iodophenyl)-4-methyl-1*H*-pyrazole-3-carboxamide; AM630, 6-iodo-2-methyl-1-[2-(4-morpholinyl)ethyl]-1-*H*-indol-3-yl(4-methoxyphenyl)-methanone; IBD, inflammatory bowel disease; PRS-211,092, [(+)-(6*aS*,10*aS*)-6,6-dimethyl-3-(1,1-dimethylheptyl)-1-hydroxy-9-(1*H*-imidazol-2-ylsulfanylmethyl)-6*a*-,7,10,10*a*-tetrahydro-6*H*-dibenzo[*b,d*]pyran; RA, rheumatoid arthritis; HU-320, cannabidiol-dimethylheptyl-7-oic acid; HU-308, (+)-(1-*aH*,3*H*,5*aH*)-4-[2,6-dimethoxy-4-(1,1-dimethylheptyl)phenyl]-6,6-dimethylbicyclo[3.1.1]hept-2-ene-2-carbinol.

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