Effects of menthol-flavored substances at the cellular level on oral mucosal sites

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Abstract

Background:
The purpose of this study was to determine the effects of menthol-flavored substances at the cellular level in different mucosal sites of the oral cavity and to compare the cellular changes between individuals without the habit of chewing menthol-flavored substances and individuals with the habit.

Materials and Methods:
This was an experimental cytology study including a total of 500 individuals belonging to the age group of 18–45 years based on the inclusion or exclusion criteria. The selected participants were divided into two groups of 250 participants each, based on participants not having the habit of chewing menthol-flavored substances (Group I) and participants having the habit of chewing menthol-flavored substances (Group II). Cytological smears were taken by gently scraping the mucosal surfaces in different sites of the oral mucosa using a wooden spatula and stained with Papanicolaou, analyzed under microscope for any cellular changes. The results were tabulated and statistically analyzed using Chi-square test and Fisher's exact test. P < 0.05 was considered statistically significant.

Results:
Micronuclei seen in all the participants belonging to group with the habit of chewing menthol-flavored substances with a P < 0.001 which was considered highly significant. Alteration in the nuclear-cytoplasmic ratio was also seen P = 0.001, which showed significant at 1% significance level.
Conclusion:

Participants with habit of chewing menthol-flavored substances showed the presence of micronuclei and slight alteration in the nuclear-cytoplasmic ratio, which could be directly related to genotoxicity and cell damage.

Keywords: Cell death, chewing, menthol

INTRODUCTION

The name “spear” mint derives from the pointed leaf tips of the plant. Mint descends from the Latin word mentha, which is rooted in the Greek word minthe, personified in Greek mythology as Minthe. There are about 26 species of mint. All of them considered as the most fragrant plants with a pleasant taste and many healing properties. The main active substances contained in the essential oil of peppermint are menthol (alcohol), the menthone (ketone), and tannins. Its leaves contain Vitamins A and C, niacin (Vitamin B3), magnesium, and iron. Menthol is one of the monocyclic terpenes that occur naturally in more than 100 essential oils, which includes spearmint and peppermint. The characteristic minty aroma and cooling qualities of synthetic d-1 menthol and the natural l-menthol isomer have resulted in its use in a variety of commercial food and pharmaceutical products. The chewing of nonfood items for pleasure has a long history. Tree resins were chewed by the ancient Egyptians, the Mayan Indians, and the early American Indians. The first commercial chewing gum, State of Maine Pure Spruce Gum, appeared in 1848. During chewing, the oral cavity functions such as a bellow, forcing volatile flavor compounds into the exhaling air to the nasal compartment. Accordingly, that flavor release from chewing gum is predominantly governed by chewing frequency, although other oral functions, such as masseter muscle activity, chewing force, and saliva flow rate, may also play a role. Menthol appears to alter cell membranes, and the findings of animal studies have suggested, that there are changes in cell membrane integrity and it does appear, however, that menthol alters cell permeability too. Menthol acts as a coolant and a mild local anesthetic. One of the more immediate cellular effects of menthol on cell membranes is that of cell death (cytotoxicity). Menthol has been shown to be toxic in vitro biologic model systems in normal tissue, it inhibits fatty acid-induced (receptor-mediated) cell respiration in brown adipose tissue and increased cellular respiration rate and osmotic swelling suggesting deterioration of biologic membranes in mitochondria. Growing evidence has shown that menthol can induce mitochondrial membrane depolarization through the transient receptor potential melastatin family member 8 (TRPM8) channel in cells of the human bladder cancer cell line T24 resulting in cell death. Since the 1920s, menthol has been added to cigarettes and used as a characterizing flavor. However, its use in chewing gums came after 1950s. Among other effects, menthol vapor can modulate sensitivity to chemical irritation in the upper airways in humans. Inhaled menthol also exerts complex olfactory and sensory effects by interacting with olfactory and somatosensory neurons and respiratory tissues. It is puzzling, in the light of increased awareness, that more studies such as ours have not been undertaken. However, few studies have been done in English literature so far to evaluate toxicity and cellular effects of menthol. Gaworski et al. exposed rats to menthol cigarette smoke through nose inhalation for 1 h a day, 5 days/week for 13 weeks. In their study, they concluded that exposure to menthol cigarette smoke produced reduced body weights and histopathological changes including epithelial hyperplasia and/or squamous metaplasia in the nasal passages, trachea, larynx, lungs, and bronchi. Olfactory epithelial degeneration was also observed in these cases. As a parallel to the findings by Gaworski et al., Alakayak and Knall found that the gap junctions between the cells were “loosened” up and integrity was lost as a result of effect of tobacco smoke effects on transepithelial electrical resistance the tight gap junctions between the human bronchial epithelial
To the best of our knowledge, no study in English literature has been done so far to study the effect of chewing menthol-flavored substances. Therefore, this study was undertaken to see the effect of the use of menthol-flavored substances at the cellular level of the mucosal sites of oral cavity.

**MATERIALS AND METHODS**

This comparative experimental cytology study was conducted with approval from Institutional and Ethical committee (SRGCDS/2015/502) in the department of oral and maxillofacial pathology in our institution from February 2016 to April 2016. Informed consent was obtained from all individuals before the initiation of the study. This study has two groups. The control group (Group I) consists of individuals not having the habit of chewing menthol-flavored substances, and the study group (Group II) consists of individuals having the habit of chewing menthol-flavored substances. A total of 500 participants were evaluated, out of which 250 participants have the habit of chewing only menthol-flavored substances for more than 5 years while the remaining 250 participants do not have the habit of chewing menthol-flavored substances. Participants in the age group between 18 and 45 years in each group were evaluated. Inclusion criteria included participants with the habit of chewing 5 or more than 5 menthol-flavored substances per day for more than 5 years. For this, the participants reporting to our outpatient department were asked randomly whether they had this habit and took their consent. Once confirmed they were asked to sign consent letter whereas the exclusion criteria included participants having the habit of chewing other than menthol-flavored substances. Participants having the habit of chewing less than 5 menthol-flavored substances and participants having any systemic diseases, smoking habits, and other oral manifestations such as proliferative, traumatic, or immune-mediated epithelial lesions (e.g., papilloma, aphthous ulcer, lichen planus, and traumatic ulcer) were excluded from our study. Cytological smears were taken by gently scraping the mucosal surfaces in different sites of the oral mucosa, using a wooden spatula supplied from Asian hobby crafts, New Delhi, India. For the preparation of smears, clean, fresh, dry glass slides (Blue star micro slides, supplied from Polar industrial corporation Mumbai, India) were used. The material from the wooden spatula was spread on the middle third of clean dried glass slides. The smears were spread over a large area, preventing the clumping of cells. The prepared slides were immediately sprayed with Biofix spray microanatomy fixative supplied from Biolab Diagnostics Pvt., Ltd., Tarapur, Maharashtra, India, to ensure proper fixation. The smears were stained by using the rapid Papanicolaou (PAP) stain supplied from Biolab Diagnostics Pvt., Ltd., Tarapur, Maharashtra, India.

**Papanicolaou stain procedure**

1. Smears were hydrated by pouring few drops of distilled water on the slide for 1–3 min
2. Excess water was blot out from the slide, and nuclear stain was poured on the slide for 45–60 s
3. On the same slide, 3 drops of buffer solution (Scottie's buffer solution) were added and kept for 30–40 s
4. After draining the buffer solution, dehydrant was poured with two changes each for 30 s
5. After discarding dehydrant, working solution (2A + 2B) was poured and kept for 45 s
6. After draining the above solution, dehydrant was poured with two changes each for 30 s followed by xylene for 30 s and mounted with coverslip using dibutyl phthalate xylene.

Stained slides from both the groups were then analyzed and verified by five different oral pathologists who were blinded to eliminate bias under the microscope (Lawrence and Mayo, London) who were reporting on the presence of micronuclei and alteration in the nucleus-cytoplasmic ratio to determine
the effects of menthol-flavored substances at the cellular level in different mucosal sites of the oral cavity, and the observatory findings were subjected to appropriate statistical analysis. Interobserver reliability for all 5 observers was tested. The Cronbach's alpha test proved that there was good interobserver reliability (>0.7) among all the five different oral pathologists.

**Statistical analysis**

All the obtained data were entered in an Excel Spreadsheet. A comparative study was done using Chi-square test and Fisher's exact test between the participants not having the habit of chewing menthol-flavored substances and participants having the habit of chewing menthol flavored substances. The values were tabulated for comparison purpose. $P < 0.05$ in Chi-square test and Fisher's exact test was accepted as indicating statistical significance.

**RESULTS**

On observing the cytological smears, micronuclei was seen in all the 250 participants belonging to study group and in none of the participants in the control group [Figure 1a and b and Table 1] with a $P < 0.001$ which was considered highly significant. Alteration in the nuclear-cytoplasmic ratio was also seen in 90 participants out of 250 participants in study group with a $P = 0.001$, which showed significant at 1% significance level [Figure 1c and Table 2].
Figure 1

(a) Photomicrograph showing the presence of micronuclei in the study group. (Papanicolaou ×10, Papanicolaou ×40). (b) Exfoliated cells in control group (Papanicolaou ×10, Papanicolaou ×40). (c) Altered nuclear-cytoplasmic ratio in the study group (Papanicolaou ×10, Papanicolaou ×40).
DISCUSSION

This study examined the consequences of using menthol-flavored substances at the cellular level of the mucosal sites of oral cavity and hence the potential for increased cytotoxicity. Menthol is one of the monocyclic terpenes that occur naturally in over 100 essential oils, which includes spearmint and peppermint. The characteristic minty aroma, cooling qualities, and mild local anesthetic properties have resulted in its use in a variety of commercial food and pharmaceutical products. The world market for chewing gum is estimated to be around 560,000 tons/year, and around 374 billion pieces of chewing gum are sold worldwide every year, representing 187 billion of gum-chewing if each piece of gum is chewed for 30 min. Chewing gum can thus be expected to have an influence on oral health. One of the more immediate cellular effects of menthol on cell membranes is that of cell death (cytotoxicity). In 1997, Gaworski et al. exposed rats to menthol or nonmenthol cigarette smoke through nose-inhalation for 1 h a day, 5 days/week for 13 weeks and concluded that smoke produced reduced body weights and histopathological changes including epithelial hyperplasia and/or squamous metaplasia in the nasal
passages, trachea, larynx, lungs, and bronchi olfactory epithelial degeneration was also observed.[2] In 2006, Azzi et al. conducted a study on permeation and reservoir formation of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone and benzo[a] pyrene across porcine esophageal tissue in the presence of ethanol and menthol and concluded that menthol alters cell permeability.[8] Alakayak and Knall 2008 conducted a study evaluating tobacco smoke effects on transepithelial electrical resistance and concluded that both menthol and nonmenthol smoke reduce transepithelial electrical resistance.[15] Wise et al. 2011 stated that menthol vapor can modulate sensitivity to chemical irritation in the upper airways in humans.[17] Similarly, many studies carried out over the past 20 years have shown that menthol is capable of increasing both the transdermal and transbuccal penetration of dideoxycytidine, propofol, propranolol, and ofloxacin. Menthol’s characteristic cooling sensation is due, in part, to the activation of sensory neurons generally termed transient receptor potential channels, in particular, TRPM8 and transient receptor potential subfamily A, member 1. Menthol acts on TRPM8 receptors by rapidly increasing intracellular calcium and mobilizing calcium flux through the channels to induce cold response signals at the application site. Aside from its cold-inducing sensation capabilities, menthol exhibits cytotoxic effects in cancer cells, induces reduction in malignant cell growth, and engages in synergistic excitation of gamma-aminobutyric receptors and sodium ion channels resulting in analgesia. [19,20] Menthol, likely due to its effects on cold-sensing peripheral sensory neurons, is known to inhibit the sensation of irritation elicited by respiratory irritants.[21] Some studies demonstrate that menthol attenuates signaling through human α3 β4 nAChRs. Menthol also acts as a competitive inhibitor on the specific binding of [3H] PN 200-110 and [3H] nitrendipine, dihydropyridine class antagonists of L-type Ca2+ channels in cardiac and smooth muscles and neuronal tissue.[22] A recent study by Pezzoli et al. 2014[23] in mouse cortical neurons reported that menthol (250 μM) dampens the generation of action potentials in a time- and voltage-dependent manner in TRPM8 knock-out mice and in the presence of a TRPM8 blocker. On reviewing English literature, we found that so far no study has been done to evaluate the effect of chewing menthol-flavored substances on the mucosal sites of the oral cavity. The parameters described in our study shows a higher frequency of micronuclei associated with participants who are in the habit of chewing menthol-flavored substances as compared to participants who do not have the habit of chewing menthol-flavored substances. A slight alteration in the nucleus-cytoplasmic ratio was also observed in participants having the habit of chewing menthol-flavored substances which may indicate short-term cytotoxicity and cell damage.

CONCLUSION

To conclude, participants with habit of chewing menthol-flavored substances (Group II) showed the presence of micronuclei and slight alteration in the nuclear-cytoplasmic ratio, which could be directly related to genotoxicity and cell damage. Although we have significant positive results in our study, further research in this area is expected with larger sample size and multiple observers where the cytological smears can be subjected to all stains, to consider menthol-flavored substances as a cytotoxic agent.

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Conflicts of interest

The authors of this manuscript declare that they have no conflicts of interest, real or perceived, financial or nonfinancial in this article.

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