

Kit Content

	36rxn	
Syringe	36	set
Elution Tube	36	pcs
AR21100 Cartridge	36	set
AR21100 Column Set	36	set
AR21100 Tip Set	36	set
EtOH Tube	36	pcs
Sample Tube	36	pcs
Buffer DWX	25	ml
Buffer RFTL	45	ml
Proteinase K	60	mg
Proteinase K Solvent	5	ml

Kit Storage

Upon arrival,

1. Please store **AR21100 Column Set** at **4°C** for long term storage.
2. Please store **Proteinase K** at **-20 °C** for long term storage.

Buffer, solvent and consumables, please store at 15-25 °C .

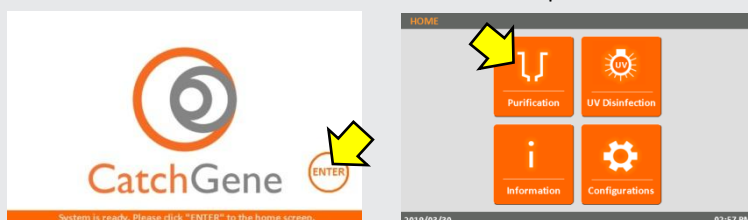
If a precipitate has formed in Buffer RFTL, dissolve by incubating at 60°C for 10 min.

Kit Preparation

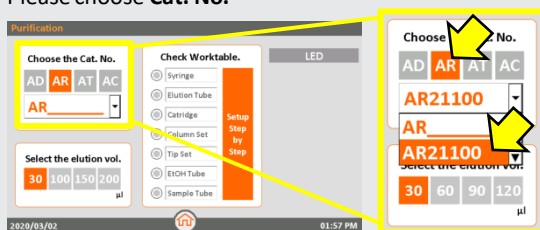
1. **Prepare 20 mg/ml Proteinase K**
For 60 mg Proteinase K, please add 3 ml Proteinase K Solvent into tube and vortex thoroughly for dissolving. After dissolving into solvent, please store in 4°C for 6 month or -20°C for 1 year.

Step by Step to start a AR21100 Purification Run

1. On the **Start** screen: Click “ENTER” button to enter the HOME screen.
2. On the **HOME** screen: Click “Purification” icon to start a purification run.

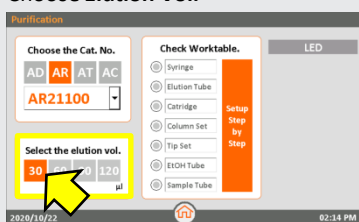


3. Please choose **Cat. No.**



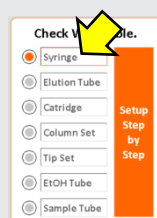
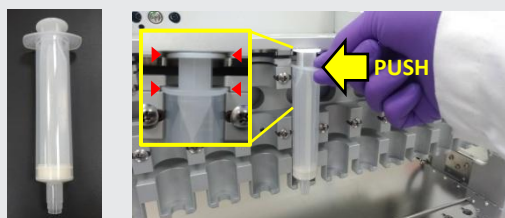
Please click “AR”
Then choose “AR21100”
For iCatcher® FFPE Tissue RNA Kit

4. Choose **Elution Vol.**



We suggest to elute in **30 µl** for FFPE Tissue RNA.

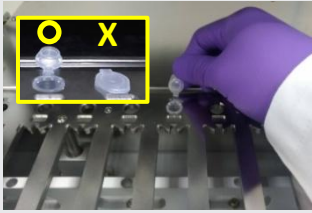
5. Insert the **Syringe** into the groove of Syringe Seat and push it to the end.



Check the **Syringe**.

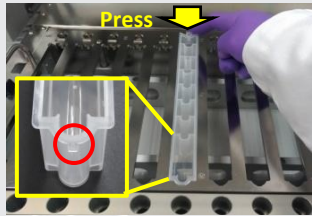
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6. Labeling, then open the lid and place the **Elution Tube** on the Elution Tube position.



Check the **Elution Tube**.

7. Insert the front protrude part of **Cartridge** into Cartridge position and press the bottom down. Then remove the foil.



Check the **Cartridge**.

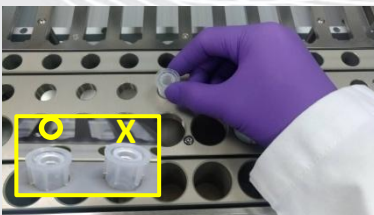
Important! Please must remove the foil before running a protocol.

8. Insert **Column Set** into Column Set position and press into bottom.



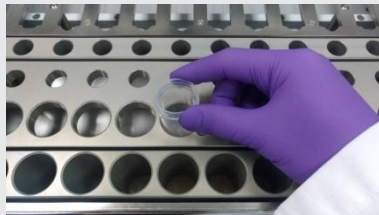
Check the **Column Set**.

9. Place **Tip Set** on Tip Set position and press into bottom.



Check the **Tip Set**.

10. Add **18 ml** 100% EtOH into **EtOH Tube** and place on the EtOH Tube position.



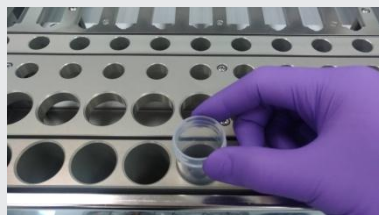
Check the **EtOH Tube**.

Add **18 ml** 100% EtOH into **EtOH Tube**

11. Prepare sample as below,

- Place 5-10 μm sections (up to 4 sections) in a Sample Tube. Add 0.6 ml DWX buffer, vortex vigorously for 30 sec. Spin down to collect sample in the bottom.
- Incubate at 60°C for 20 min.
- Add 1 ml RFTL Buffer (Please add 1% β - mercaptoethanol freshly) and mix thoroughly by vortex 10 sec.
- Centrifuge at 11,000 x g for 1 min. (After centrifugation, sample will separate into two layers. Upper layer is in yellow color which mainly Buffer DWX. Lower layer is colorless which mainly Buffer RFTL and tissue debris.)
- Add 80 μl PK (20 mg/ml) to the lower clear phase. Mix gently by pipetting.
- Incubate at 60°C for 30 min. Brief spin down.
- Incubate at 80°C for 15min. (Please avoid leaving samples in the incubator and heating it from 60°C to 80°C . Please place take samples out from incubator and place them back while the incubator reach 80°C . Otherwise it will affect the result of de-cross-linking.)
- Centrifuge at 11,000 x g for 2 min.
- Transfer 1000 μl lower clear phase lysate (avoid to aspirate any debris) into a Sample Tube.

12. Load the Sample Tube into the **Sample Tube** position of iCatcher.



Check the **Sample Tube**.
Click "Go" to start purification.

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