iCatcher® Stool DNA 4000 Kit

Cat. No.	Rxn
AD22400-36	36

Kit Content

	36rxn	
Syringe	36	set
Elution Tube	36	pcs
AD22400 Cartridge	36	set
AD22400 Column Set	36	set
AD22400 Tip Set	36	set
EtOH Tube	36	pcs
Sample Tube	36	pcs
3mm Beads Tube	36	tube
Buffer ST1	175x2	ml
Buffer ST2	115	ml
Proteinase K	152	mg
Proteinase K Solvent	10	ml

Kit Storage

Upon arrival,

1. Please store **Proteinase K** at **-20 ℃** for long term storage.

Buffer, solvent and consumables, please store at 15-25 ℃.

If a precipitate has formed in Buffer ST1, dissolve by incubating at 60˚C for 10 min.

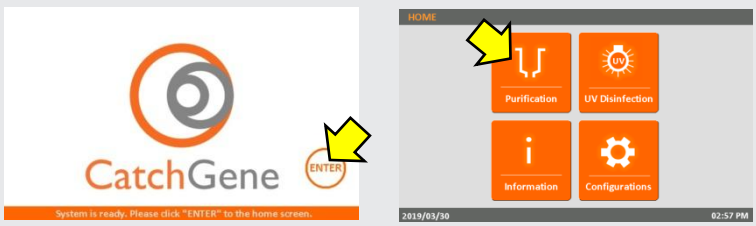
Kit Preparation

Prepare 20 mg/ml Proteinase K

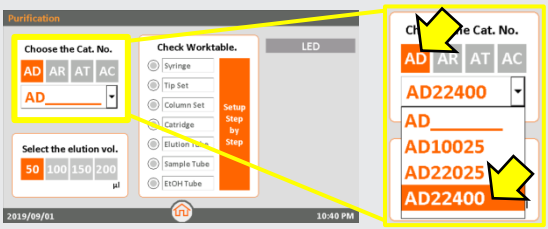
For 11 mg Proteinase K, please add 550 µl Proteinase K Solvent into tube and vortex thoroughly for dissolving.
For 130 mg Proteinase K, please add 6.5 ml Proteinase K Solvent into tube and vortex thoroughly for dissolving.
After dissolving into solvent, plase store in 4°C for 6 month or -20°C for 1 year.

Step by Step to start a AD22400 Purification Run

1. On the **Start** screen: Click "ENTER" button to enter the HOME screen.
2. On the **HOME** screen: Click "Purification" icon to start a purification run.

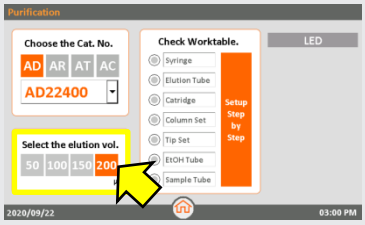


3. Please choose **Cat. No.**



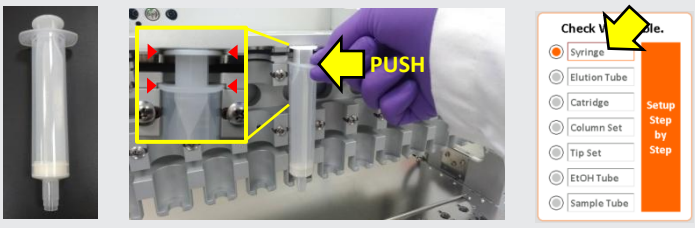
Please click "**AD**"
Then choose "**AD22400**"
For **iCatcher® Stool DNA 4000 Kit**

4. Choose **Elution Vol.**



We suggest to elute in **200µl** for stool DNA.

5. Insert the **Syringe** into the groove of Syringe Seat and push it to the end.



Check the **Syringe**.

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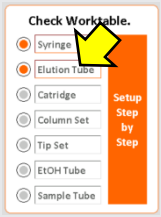
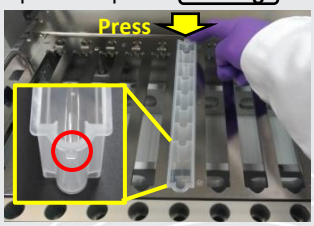
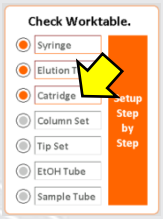
6. Labeling , then open the lid and place the **Elution Tube** on the Elution Tube position.

Check the **Elution Tube**.

7. Insert the front protrude part of **Cartridge** into Cartridge position and press the bottom down. Then remove the foil.





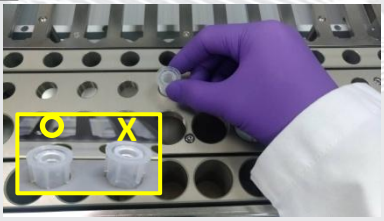
Check the **Cartridge**.

Important! Please must **remove the foil** before running a protocol.

8. Insert **Column Set** into Column Set position and press into bottom.

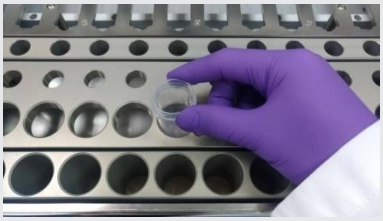
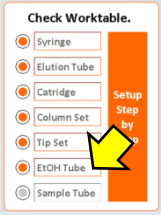

Check the **Column Set**.

9. Place **Tip Set** on Tip Set position and press into bottom.

Check the **Tip Set**.

10. Add **24 ml** 100% EtOH into **EtOH Tube** and place on the EtOH Tube position.

Check the **EtOH Tube**.

Add **24 ml** 100% EtOH into **EtOH Tube**

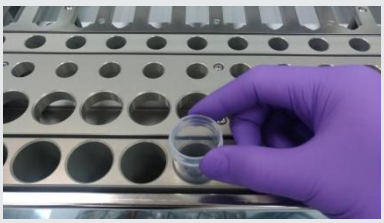
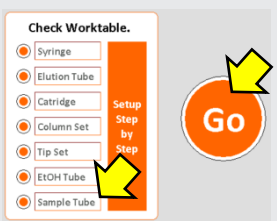
11. Prepare sample as below,

For raw stool sample without preservation buffer

a. Weigh up to 500 mg stool sample into the 50 ml tube and add one tube of 3 mm Beads Tube in.
b. Add 9 ml Buffer ST1 to tube. Vortex continuously for 5 min or until the stool sample is thoroughly homogenized.
c. Proceed to step "e" in below.

For stool sample in preservation buffer tube

a. Add one tube of 3 mm Beads into preservation tube (with maximum 2 g of stool sample inside).
b. Vortex continuously for 5 min or until the stool sample is thoroughly homogenized.
c. Centrifuge at 3,000 x g for 10 min to pellet stool debris, then transfer up to 4 ml supernatant into a new 15/50 ml tube.
d. Add 5 ml Buffer ST1 to the tube, vortex 10 sec to let buffer and sample mix properly.
(If the supernatant from step "c" is less then 4ml, increase the volume of Buffer ST1 to let final volume reach to 9 ml.)
e. Incubate at 95°C for 15 min, then cool down to room temperature.
f. Add 3 ml Buffer ST2, vortex for 15 s and incubate on ice for 15 min.
g. Centrifuge sample at 12,000 x g for 15 min to pellet stool debris.
h. Transfer 4 ml of the supernatant into the 30 ml Sample Tube (Avoid to aspirate any gel like precipitate or stool debris.)
i. Add 200 µl Proteinase K . (no need to mix or pipette it)
j. (Optional) Add 4 µl of 10 mg/ml RNase A (not included) (no need to mix or pipette it)
k. Load the **Sample Tube** into the Sample Tube position of iCatcher (no need to mix or pipette it).

Check the **Sample Tube**.
Click "**Go**" to start purification.

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