altona DIAGNOSTICS

Instructions for Use

RealStar®
Clostridium difficile PCR Kit 2.0

03/2019 EN

RealStar®

RealStar®

Clostridium difficile PCR Kit 2.0

For use with

Mx 3005P™ QPCR System (Stratagene)

VERSANT® kPCR Molecular System AD (Siemens Healthcare Diagnostics)

ABI Prism® 7500 SDS (Applied Biosystems)

ABI Prism® 7500 Fast SDS (Applied Biosystems)

LightCycler® 480 Instrument II (Roche)

Rotor-Gene® 6000 (Corbett Research)

Rotor-Gene® Q5/6 plex Platform (QIAGEN)

CFX96™ Real-Time PCR Detection System (Bio-Rad)

CFX96™ Deep Well Real-Time PCR Detection System (Bio-Rad)

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Content

1. Intended Use

The RealStar® Clostridium difficile PCR Kit 2.0 is an *in vitro* diagnostic test, based on real-time PCR technology, for the qualitative detection and differentiation of *toxin A (tcdA)* and *toxin B (tcdB)* specific DNA of *Clostridium difficile*.

2. Kit Components

Lid Color	Component	Number of Vials	Volume [µl/Vial]
Blue	Master A	8	60
Purple	Master B	8	180
Green	Internal Control	1	1000
Red	Positive Control	1	250
White	Water (PCR grade)	1	500

3. Storage

- The RealStar® *Clostridium difficile* PCR Kit 2.0 is shipped on dry ice. The components of the kit should arrive frozen. If one or more components are not frozen upon receipt, or if tubes have been compromised during shipment, contact altona Diagnostics GmbH for assistance.
- All components should be stored between -25°C and -15°C upon arrival.
- Repeated thawing and freezing of Master reagents (more than twice) should be avoided, as this might affect the performance of the assay. The reagents should be frozen in aliquots, if they are to be used intermittently.
- Storage between +2°C and +8°C should not exceed a period of two hours.
- Protect Master A and Master B from light.

4. Material and Devices required but not provided

- Appropriate real-time PCR instrument (see chapter 6.1 Real-Time PCR Instruments)
- Appropriate nucleic acid extraction system or kit (see chapter 8.1 Sample Preparation)
- Desktop centrifuge with a rotor for 2 ml reaction tubes
- Centrifuge with a rotor for microtiter plates, if using 96 well reaction plates
- Vortex mixer
- Appropriate 96 well reaction plates or reaction tubes with corresponding (optical) closing material
- Pipettes (adjustable)
- Pipette tips with filters (disposable)
- Powder-free gloves (disposable)

NOTE

Please ensure that all instruments used have been installed, calibrated, checked and maintained according to the manufacturer´s instructions and recommendations.

NOTE

It is highly recommended to use the 72-well rotor with the appropriate 0.1 ml reaction tubes, if using the Rotor-Gene® 6000 (Corbett Research) or the Rotor-Gene® Q 5/6 plex (QIAGEN).

5. Background Information

Clostridium difficile is a spore forming, anaerobic bacterium in the genus *Clostridium*. The species comprises a diverse population structure with hundreds of different strain types. The bacteria are gram positive with a large circular genome of about 4.3 Mb. [1]

The bacterium is transmitted via the fecal-oral route often during hospitalization, which is due to improper isolation of infected patients and poor hygienic routine. But not all infected patients show symptoms. *Clostridium difficile* cannot grow in the normal, healthy gastrointestinal flora since other bacteria impair its proliferation. After eradication of the normal flora, due to administration of antibiotics, an overgrowth and full colonization of the colon is possible. [2]

After colonization of the colon the bacteria can produce either one or two toxins, toxin A and B, which cause the pathogenic effect. This toxin production is triggered by *quorum sensing*. The produced toxins disrupt mucosal cell adherence (*tcdA*) and induce apoptosis (*tcdB*), which can lead to various pathologies ranging from mild diarrhea to life threatening inflammatory complications like pseudomembranous colitis or toxic megacolon. In 1.5 % of all hospitalized cases with *Clostridium difficile* diarrhea, the infection is fatal, with the highest risk for elderly patients. [2, 3]

Affected patients receive antibiotic treatment and supportive measures to counteract dehydration and electrolyte loss. But the spore form of *Clostridium difficile* is resistant to the antibiotics which aggravates the treatment and can lead to the reoccurrence of symptoms after withdrawal of the antibiotic treatment. [2] *Clostridium difficile* is a leading cause of antibiotic-associated diarrhea and healthcare-associated infections in developed countries with a considerable high economic impact. As the number of severe infections has continued to increase over the last decades, the need for rapid and accurate detection and treatment rises to lower mortality, medical costs and for infection control. [4]

[1] Knight DR, Elliot B, Chang BJ, Perkins TT, Riley TV (2015) Diversity and Evolution in the Genome of *Clostridium difficile*. Clin Microbiol Rev. 28: 721-741.

[2] Tonna I and Welsby PD (2005). Pathogenesis and treatment of Clostridium difficile infection. Postgrad Med J. 81: 367-369.

[3] Kirk JA, Banerji O, Fagan RP (2017). Characteristics of the Clostridium difficile cell envelope and its importance in therapeutics. Microb Bioltechnol. 10: 76-90.

[4] Peng Z, Ling L, Stratton CW, Li C, Polage CR, Wu B, Tang Y-W (2018). Advances in the diagnosis and treatment of Clostridium difficile infections. Emerg Microbes Infect. Doi: 10.1038/s41426-017-0019-4.

6. Product Description

The RealStar® Clostridium difficile PCR Kit 2.0 is an *in vitro* diagnostic test, based on real-time PCR technology, for the qualitative detection and differentiation of *toxin A (tcdA)* and *toxin B (tcdB)* specific DNA of *Clostridium difficile*.

The assay includes a heterologous amplification system (Internal Control) to identify possible PCR inhibition and to confirm the integrity of the reagents of the kit.

Real-time PCR technology utilizes polymerase chain reaction (PCR) for the amplification of specific target sequences and target specific probes for the detection of the amplified DNA. The probes are labelled with fluorescent reporter and quencher dyes.

Probes specific for *tcdA* DNA are labelled with the fluorophore Cy®5 whereas the probes specific for *tcdB* DNA are labelled with the fluorophore FAM™. The probe specific for Internal Control (IC) is labelled with the fluorophore JOE™.

Using probes linked to distinguishable dyes enables the parallel detection of *tcdA* and *tcdB* specific DNA as well as the detection of the Internal Control in corresponding detector channels of the real-time PCR instrument.

The test consists of two processes in a single tube assay:

- PCR amplification of target DNA and Internal Control
- Simultaneous detection of PCR amplicons by fluorescent dye labelled probes

The RealStar® Clostridium difficile PCR Kit 2.0 consists of:

- Master A
- Master B
- Internal Control
- Positive Control
- Water (PCR grade)

Master A and Master B contain all components (PCR buffer, DNA polymerase, magnesium salt, primers and probes) to allow PCR mediated amplification and detection of *tcdA* specific DNA, *tcdB* specific DNA and Internal Control in one reaction setup.

6.1 Real-Time PCR Instruments

The RealStar® Clostridium difficile PCR Kit 2.0 was developed and validated to be used with the following real-time PCR instruments:

- Mx 3005P™ QPCR System (Stratagene)
- VERSANT® kPCR Molecular System AD (Siemens Healthcare Diagnostics)
- ABI Prism® 7500 SDS (Applied Biosystems)
- ABI Prism® 7500 Fast SDS (Applied Biosystems)
- LightCycler® 480 Instrument II (Roche)
- Rotor-Gene® 6000 (Corbett Research)
- Rotor-Gene® Q5/6 plex Platform (QIAGEN)
- CFX96™ Real-Time PCR Detection System (Bio-Rad)
- CFX96™ Deep Well Real-Time PCR Detection System (Bio-Rad)

7. Warnings and Precautions

Read the Instructions for Use carefully before using the product.

- Before first use check the product and its components for:
  - Integrity
  - Completeness with respect to number, type and filling (see chapter 2. Kit Components)
  - Correct labelling
  - Frozenness upon arrival
- Use of this product is limited to personnel specially instructed and trained in the techniques of real-time PCR and *in vitro* diagnostic procedures.
- Specimen should always be treated as infectious and/or biohazardous in accordance with safe laboratory procedures.
- Wear protective disposable powder-free gloves, a laboratory coat and eye protection when handling specimens.
- Avoid microbial and nuclease (DNase/RNase) contamination of the specimens and the components of the kit.
- Always use DNase/RNase-free disposable pipette tips with aerosol barriers.
- Always wear protective disposable powder-free gloves when handling kit components.
- Use separated and segregated working areas for (i) sample preparation, (ii) reaction setup and (iii) amplification/detection activities. The workflow in the laboratory should proceed in unidirectional manner. Always wear disposable gloves in each area and change them before entering a different area.
- Dedicate supplies and equipment to the separate working areas and do not move them from one area to another.
- Store positive and/or potentially positive material separated from all other components of the kit.
- Do not open the reaction tubes/plates post amplification, to avoid contamination with amplicons.

- Additional controls may be tested according to guidelines or requirements of local, state and/or federal regulations or accrediting organizations.
- Do not autoclave reaction tubes after the PCR, since this will not degrade the amplified nucleic acid and will bear the risk to contaminate the laboratory area.
- Do not use components of the kit that have passed their expiration date.
- Discard sample and assay waste according to your local safety regulations.

8. Procedure

8.1 Sample Preparation

Extracted DNA is the starting material for the RealStar® Clostridium difficile PCR Kit 2.0.

The quality of the extracted DNA has a profound impact on the performance of the entire test system. It is recommended to ensure that the system used for nucleic acid extraction is compatible with real-time PCR technology. The following kits and systems are suitable for nucleic acid extraction:

- QIAamp® DNA Mini Kit (QIAGEN)
- QIAsymphony® (QIAGEN)
- NucliSENS® easyMag® (bioMérieux)
- MagNA Pure 96 System (Roche)
- m2000sp (Abbott)
- Maxwell® 16 IVD Instrument (Promega)
- VERSANT® kPCR Molecular System SP (Siemens Healthcare)

Alternative nucleic acid extraction systems and kits might also be appropriate. The suitability of the nucleic acid extraction procedure for use with RealStar® Clostridium difficile PCR Kit 2.0 has to be validated by the user.

If using a spin column based sample preparation procedure including washing buffers containing ethanol, it is highly recommended to perform an additional centrifugation step for 10 min at approximately 17000 x g (~ 13000 rpm), using a new collection tube, prior to the elution of the nucleic acid.

CAUTION

If your sample preparation system is using washing buffers containing ethanol, make sure to eliminate any traces of ethanol prior to elution of the nucleic acid. Ethanol is a strong inhibitor of real-time PCR.

CAUTION

The use of carrier RNA is crucial for extraction efficiency and stability of the extracted nucleic acid.

For additional information and technical support regarding pre-treatment and sample preparation please contact our Technical Support (see chapter 14. Technical Assistance).

8.2 Master Mix Setup

All reagents and samples should be thawed completely, mixed (by pipetting or gentle vortexing) and centrifuged briefly before use.

The RealStar® Clostridium difficile PCR Kit 2.0 contains a heterologous Internal Control (IC), which can either be used as a PCR inhibition control or as a control of the sample preparation procedure (nucleic acid extraction) and as a PCR inhibition control.

► If the IC is used as a PCR inhibition control, but not as a control for the sample preparation procedure, set up the Master Mix according to the following pipetting scheme:

Number of Reactions (rxns)	1	12
Master A	5 µl	60 µl
Master B	15 µl	180 µl
Internal Control	1 µl	12 µl
Volume Master Mix	**21 µl**	**252 µl**

- If the IC is used as a control for the sample preparation procedure and as a PCR inhibition control, add the IC during the nucleic acid extraction procedure.
- No matter which method/system is used for nucleic acid extraction, the IC **must not** be added directly to the specimen. The IC should always be added to the specimen/lysis buffer mixture. The volume of the IC which has to be added, always and only depends on the elution volume. It represents 10% of the elution volume. For instance, if the nucleic acid is going to be eluted in 60 µl of elution buffer or water, 6 µl of IC per sample must be added into the specimen/lysis buffer mixture.
- If the IC was added during the sample preparation procedure, set up the Master Mix according to the following pipetting scheme:

Number of Reactions (rxns)	1	12
Master A	5 µl	60 µl
Master B	15 µl	180 µl
Volume Master Mix	**20 µl**	**240 µl**

CAUTION

If the IC (Internal Control) was added during the sample preparation procedure, at least the negative control must include the IC.

CAUTION

No matter which method/system is used for nucleic acid extraction, never add the IC directly to the specimen.

8.3 Reaction Setup

- ► Pipette 20 µl of the Master Mix into each required well of an appropriate optical 96-well reaction plate or an appropriate optical reaction tube.
- ► Add 10 µl of the sample (eluate from the nucleic acid extraction) or 10 µl of the controls (Positive or Negative Control).

Reaction Setup	
Master Mix	20 µl
Sample or Control	10 µl
Total Volume	**30 µl**

- ► Make sure that at least one Positive Control and at least one Negative Control is used per Master Mix and run.
- ► Thoroughly mix the samples and controls with the Master Mix by pipetting up and down.
- ► Close the 96-well reaction plate with appropriate lids or optical adhesive film and the reaction tubes with appropriate lids.
- ► Centrifuge the 96-well reaction plate in a centrifuge with a microtiter plate rotor for 30 seconds at approximately 1000 x g (~ 3000 rpm).

9. Programming the Real-Time PCR Instrument

For basic information regarding the setup and programming of the different real-time PCR instruments, please refer to the user manual of the respective instrument.

For detailed programming instructions regarding the use of the RealStar® Clostridium difficile PCR Kit 2.0 on specific real-time PCR instruments please contact our Technical Support (see chapter 14. Technical Assistance).

9.1 Settings

► Define the following settings:

Settings	
Reaction Volume	30 µl
Ramp Rate	Default
Passive Reference	None

9.2 Fluorescence Detectors (Dyes)

► Define the fluorescence detectors (dyes):

Target	Detector Name	Reporter	Quencher
tcdA specific DNA	*tcdA*	Cy®5	(None)
tcdB specific DNA	*tcdB*	FAM™	(None)
Internal Control	IC	JOE™	(None)

9.3 Temperature Profile and Dye Acquisition

► Define the temperature profile and dye acquisition:

	Stage	Cycle Repeats	Acquisition	Temperature [°C]	Time [min:sec]
Denaturation	Hold	1	-	95	02:00
Amplification	Cycling	45	-	95	00:15
			yes	58	00:45
			-	72	00:15

10. Data Analysis

For basic information regarding data analysis on specific real-time PCR instruments, please refer to the user manual of the respective instrument.

For detailed instructions regarding the analysis of the data generated with the RealStar® Clostridium difficile PCR Kit 2.0 on different real-time PCR instruments please contact our Technical Support (see chapter 14. Technical Assistance).

10.1 Validity of Diagnostic Test Runs

10.1.1 Valid Diagnostic Test Run (qualitative)

A **qualitative** diagnostic test run is **valid**, if the following control conditions are met:

Control ID	Detection Channel		
	Cy®5	FAM™	JOE™
Positive Control [*tcdA* and *tcdB*]	+	+	+/-*
Negative Control	-	-	+

* The presence or absence of a signal in the JOE™ channel is not relevant for the validity of the test run.

10.1.2 Invalid Diagnostic Test Run (qualitative)

A **qualitative** diagnostic test run is **invalid**, (i) if the run has not been completed or (ii) if any of the control conditions for a **valid** diagnostic test run are not met.

In case of an **invalid** diagnostic test run, repeat testing by using the remaining purified nucleic acids or start from the original samples again.

10.2 Interpretation of Results

10.2.1 Qualitative Analysis

Detection Channel			Result Interpretation
Cy®5	FAM™	JOE™	
+	+	+*	*tcdA* and *tcdB* specific DNA detected.
+	-	+*	*tcdA* specific DNA detected.
-	+	+*	*tcdB* specific DNA detected.
-	-	+	Neither *tcdA* nor *tcdB* specific DNA detected. The sample does not contain detectable amounts of *tcdA* or *tcdB* specific DNA.
-	-	-	PCR inhibition or reagent failure. Repeat testing from original sample or collect and test a new sample.

* Detection of the Internal Control in the JOE™ detection channel is not required for positive results either in the Cy®5 detection channel or in the FAM™ detection channel. A high *tcdA* and/or *tcdB* DNA load in the sample can lead to reduced or absent Internal Control signals.

11. Performance Evaluation

Performance evaluation of the RealStar® Clostridium difficile PCR Kit 2.0 was done using quantified DNA of *Clostridium difficile* strain ATCC®BAA-1804™ from the American Type Culture containing both targets (toxin A (*tcdA*) and toxin B (*tcdB*)).

11.1 Analytical Sensitivity

The analytical sensitivity of the RealStar® Clostridium difficile PCR Kit 2.0 is defined as the concentration (copies/µl of the eluate) of *tcdA* or *tcdB* specific DNA molecules that can be detected with a positivity rate of 95%. The analytical sensitivity was determined by analysis of dilution series of quantified *tcdA* DNA and *tcdB* DNA.

Table 1: PCR results used for the calculation of the analytical sensitivity with respect to the detection of *tcdA* specific DNA

Input Conc. [copies/µl]	Number of Replicates	Number of Positives	Hit Rate [%]
100.000	24	24	100.000
31.620	24	24	100.000
10.000	24	24	100.000
3.162	24	24	100.000
1.000	24	24	100.000
0.316	24	23	95.833
0.200	24	21	87.500
0.100	24	14	58.333
0.032	24	7	29.167
0.010	24	4	16.667
0.0032	24	1	4.167

Table 2: PCR results used for the calculation of the analytical sensitivity with respect to the detection of *tcdB* specific DNA

Input Conc. [copies/µl]	Number of Replicates	Number of Positives	Hit Rate [%]
100.00	24	24	100.000
31.62	24	24	100.000
10.00	24	24	100.000
3.162	24	24	100.000
1.000	24	24	100.000
0.316	24	24	100.000
0.200	24	21	100.000
0.100	24	10	41.667
0.032	24	5	20.833

Input Conc. [copies/µl]	Number of Replicates	Number of Positives	Hit Rate [%]
0.010	24	2	8.333
0.0032	24	1	4.167

The analytical sensitivity of the RealStar® Clostridium difficile PCR Kit 2.0 was determined by Probit analysis:

- For the detection of *tcdA* specific DNA, the analytical sensitivity is 0.46 copies/µl [95% confidence interval (CI): 0.28 - 0.96 copies/µl]
- For the detection of *tcdB* specific DNA, the analytical sensitivity is 0.47 copies/µl [95% confidence interval (CI): 0.30 - 0.93 copies/µl]

11.2 Analytical Specificity

Reactivity

The analytical specificity with respect to the reactivity of the RealStar® Clostridium difficile PCR Kit 2.0 was evaluated by a panel of genomic DNA extracted from *C. difficile* strains producing different toxins.

The RealStar® Clostridium difficile PCR Kit 2.0 is able to detect and differentiate DNA of following *C. difficile* strains producing different toxins:

- ATCC® BAA-1875™ *Clostridium difficile* (presence of tcdB genes confirmed by PCR)
- ATCC® BAA-1875™ *Clostridium difficile* (presence of tcdA and tcdB genes confirmed by PCR)
- ATCC® BAA-1801™ *Clostridium difficile* (absence of tcdA and tcdB genes confirmed by PCR)

Specificity

The analytical specificity of the RealStar® Clostridium difficile PCR Kit 2.0 was evaluated by testing a panel of genomic RNA/DNA extracted from different gastrointestinal pathogens and commensal flora found in the intestine and stool.

The RealStar® Clostridium difficile PCR Kit 2.0 did not cross-react with any of the following pathogens:

- Astrovirus
- *Campylobacter coli*
- *Campylobacter jejuni*
- *Campylobacter lari*
- *Clostridium sordellii*
- *Cryptococcus neoformans*
- *Entamoeba histolytica*
- *Enterococcus faecalis*
- *Enterohemorrhagic Escherichia coli (EHEC)*
- *Escherichia coli*
- *Giardia lamblia*
- *Norovirus GI*
- *Norovirus GII*
- *Rotavirus*
- *Proteus mirabilis*
- *Proteus vulgaris*
- *Salmonella enterica*
- *Sapovirus*
- *Shigella flexneri*
- *Yersinia enterocolitica*

11.3 Precision

Precision of the RealStar® Clostridium difficile PCR Kit 2.0 was determined as intra-assay variability (variability within one experiment), inter-assay variability (variability between different experiments) and inter-lot variability (variability between different production lots). Total variability was calculated by combining the three analyses.

The variability data are expressed in terms of standard deviation and coefficient of variation based on threshold cycle (C_t) - values. At least 6 replicates per sample were analysed for intra-assay variability, inter-assay and inter-lot variability.

Table 3: Precision data for the detection of *tcdA* and *tcdB* specific DNA

tcdA and *tcdB*		Average Threshold Cycle (C_t)	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	*tcdA*	30.91	0.15	0.49
	tcdB	30.59	0.15	0.47
Inter-Assay Variability	*tcdA*	30.77	0.18	0.58
	tcdB	30.82	0.20	0.64
Inter-Lot Variability	*tcdA*	30.57	0.13	0.41
	tcdB	30.63	0.12	0.39
Total Variability	*tcdA*	30.68	0.12	0.39
	tcdB	30.75	0.21	0.68

Table 4: Precision data for the detection of the Internal Control

Internal Control	Average Threshold Cycle (C_t)	Standard Deviation	Coefficient of Variation [%]
Intra-Assay Variability	26.37	0.08	0.30
Inter-Assay Variability	26.28	0.12	0.47
Inter-Lot Variability	26.17	0.06	0.23
Total Variability	26.24	0.12	0.45

12. Limitations

- Strict compliance with the Instructions for Use is required for optimal results.
- Use of this product is limited to personnel specially instructed and trained in the techniques of real-time PCR and *in vitro* diagnostic procedures.
- Good laboratory practice is essential for proper performance of this assay. Extreme care should be taken to preserve the purity of the components of the

kit and reaction setups. All reagents should be closely monitored for impurity and contamination. Any suspicious reagents should be discarded.

- Appropriate specimen collection, transport, storage and processing procedures are required for the optimal performance of this test.
- This assay must not be used on the specimen directly. Appropriate nucleic acid extraction methods have to be conducted prior to using this assay.
- The presence of PCR inhibitors (e.g. heparin) may cause false negative or invalid results.
- Potential mutations within the target regions of the *tcdA* and *tcdB* genome covered by the primers and/or probes used in the kit may result in failure to detect the presence of the pathogens.
- As with any diagnostic test, results of the RealStar® Clostridium difficile PCR Kit 2.0 need to be interpreted in consideration of all clinical and laboratory findings.

13. Quality Control

In accordance with the altona Diagnostics GmbH ISO EN 13485-certified Quality Management System, each lot of RealStar® Clostridium difficile PCR Kit 2.0 is tested against predetermined specifications to ensure consistent product quality.

14. Technical Assistance

For customer support, please contact our Technical Support:

e-mail: **support@altona-diagnostics.com**
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15. Literature

Versalovic, James, Carroll, Karen C.,Funke, Guido, Jorgensen, James H., Landry, Marie Louise and David W. Warnock (ed). Manual of Clinical Microbiology. 10th Edition. ASM Press, 2011.

Cohen, Jonathan, Powderly, William G, and Steven M Opal. Infectious Diseases, Third Edition. Mosby, 2010.

16. Trademarks and Disclaimers

RealStar® (altona Diagnostics); ABI Prism® (Applied Biosystems); ATCC® (American Type Culture Collection); CFX96™ (Bio-Rad); Cy® (GE Healthcare); FAM™, JOE™, ROX™ (Life Technologies); LightCycler® (Roche); SmartCycler® (Cepheid); Maxwell® (Promega); Mx 3005P™ (Stratagene); NucliSENS®, easyMag® (bioMérieux); Rotor-Gene®, QIAamp®, MinElute®, QIAsymphony® (QIAGEN); VERSANT® (Siemens Healthcare).

Registered names, trademarks, etc. used in this document, even if not specifically marked as such, are not to be considered unprotected by law.

The RealStar® Clostridium difficile PCR Kit 2.0 is a CE-marked diagnostic kit according to the European *in vitro* diagnostic directive 98/79/EC.

Product not licensed with Health Canada and not FDA cleared or approved.

Not available in all countries.

17. Explanation of Symbols

Symbol	Explanation
IVD	*In vitro* diagnostic medical device
LOT	Batch code
CAP	Cap color
REF	Catalogue number
CONT	Content
NUM	Number
COMP	Component
GTIN	Global trade item number
	Consult instructions for use
	Contains sufficient for “n” tests/reactions (rxns)
	Temperature limit
	Use-by date
	Manufacturer
	Caution
	Note
	Version

Notes:

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always a drop ahead.

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