

T25 flask

*Coating T25 flasks. Add 2 ml AlphaBioCoat (AC001) into 3- T25 flasks and ensure entire interior surface is coated with the solution. After 30 minutes, dispose of AlphaBioCoat (AC001) by aspiration. Gently rinse and aspirate the flask with Phosphate Buffer Solution (1XPBS-001). The flask is now ready for use (no need for overnight incubation when coated with AC001). Add fresh media to flask, if color changes from pink to yellow, discard the media, and add fresh media to each flask.

1. Inspect to make sure Flask is at 90% confluence, if not remove transport media, and add 5ml of fresh media to the flask. Place flask in 37°C incubator until cells are at 90% confluence. Change media every 2 days.
2. If flask is at 90% confluence, aspirate transport media from flask.
3. Rinse T25 flask containing cells with 5 ml 1XPBS (1XPBS-001).
4. Gently aspirate out the PBS after rinsing, and discard.
5. Add 2ml of RT trypsin/ EDTA to T25 flask containing cells (ensure entire interior surface is covered).
6. Place T25 flask containing cells into 37°C incubator for 1 or 2 minutes (cells will normally come off of the surface within 1 or 2 minutes).
7. Suspend the cells with 15ml of ENDO-Growth medium (EGM-2102) and transfer equally into 3 pre-coated T25 flasks (the cells are now at a subculture ratio of 1:3).
8. There is no need to spin cells during subculture.
9. Proliferating cell culture: ENDO-Growth medium (EGM-2102) should be changed every 2 days. The cells normally become confluent within 7 days (when split at a 1:3 ratio)
10. Use ENDO- Basal media (EBM-002) containing 0.5% FBS to induce quiescent cells (after 18-24 hours).