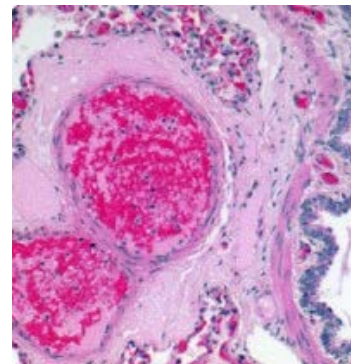


Hematoxylin and Eosin Stain Kit

Description

The Hematoxylin and Eosin Stain Kit is for histology and cytology applications. Included in this kit is a newly formulated Eosin that provides the benefits of a traditional alcoholic formula with significant improvements in usability. Advantages include lower evaporation rate, better color patterns, reduced tendency to spill over container, hands, and countertops, and improved surface tension to remain on tissue section. Our Hematoxylin produces crisp, intense blue nuclei providing optimal contrast to the Eosin-stained cytoplasm.



| | |
|---------------------|------------|
| Cytoplasm | Light Pink |
| Collagen | Pink |
| Muscle | Pink/Rose |
| Erythrocytes | Pink/Red |
| Nuclei | Blue |

Uses/Limitations

Not to be taken internally.
 For in vitro diagnostic use only.
 Histological application
 Do not use if reagents become cloudy. Do not use past expiration date.
 Use caution when handling reagents. Non-Sterile.

Control Tissue

Any well fixed paraffin embedded or frozen tissue section.
 Cell smear.

Kit Contents

| Catalog | Product | Volume | Storage |
|---------|--|--------|---------|
| HHMM500 | Hematoxylin, Mayer's (Lillie's Modification) | 500 ml | 18-25°C |
| HBRT500 | Bluing Reagent | 500 ml | 18-25°C |
| HEYB500 | Eosin Y Solution (Modified Alcoholic) | 500 ml | 18-25°C |

Precautions

Avoid contact with skin and eyes.
 Harmful if swallowed.
 Follow all Federal, State, and local regulations regarding disposal.

Procedure

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Apply adequate Hematoxylin, Mayer's (Lillie's Modification) to completely cover tissue section and incubate for 5 minutes.
3. Rinse slide in 2 changes of distilled water to remove excess stain.
4. Apply adequate Bluing Reagent to completely cover tissue section and incubate for 10-15 seconds.
5. Rinse slide in 2 changes of distilled water.
6. Dip slide in absolute alcohol and blot excess off.
7. Apply adequate Eosin Y Solution (Modified Alcoholic) to completely cover tissue section to excess and incubate for 2-3 minutes.
8. Rinse slide using absolute alcohol.
9. Dehydrate slide in 3 changes of absolute alcohol.
10. Clear slide and mount in synthetic resin.

References

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