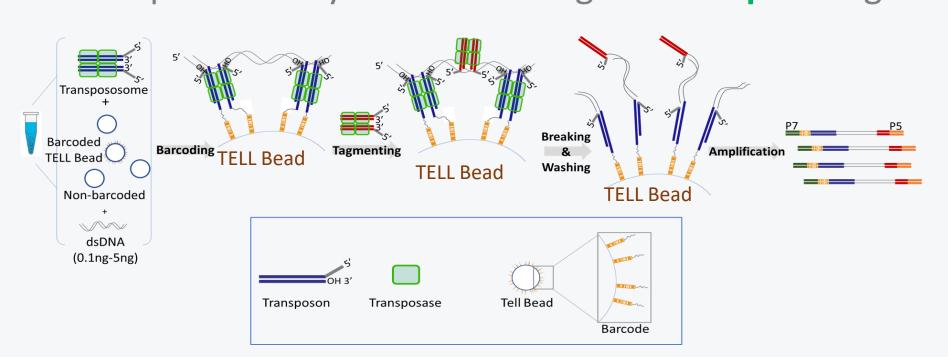
Tell-TaxContigs: Microbial Metagenome Assembly, Taxonomy, and Abundance Estimation Using UST TELL-seq Long-range Information

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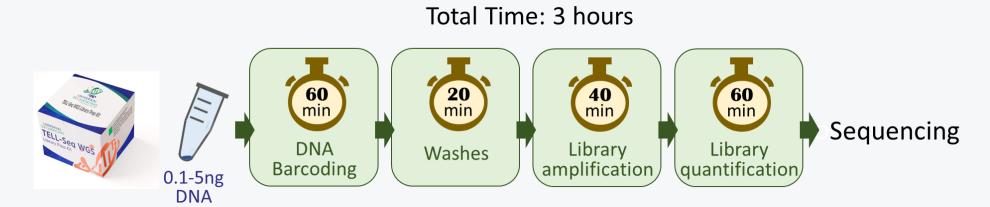
Abstract. Deconvolving diversity of a metagenome is critical for understanding the role of a given microbial community in human health and disease, small molecule biosynthesis, and other complex ecosystems where more reductive analysis proves to be elusive. Metagenomic assembly is one common method for characterizing a metagenome, especially for identifying novel gene content or novel organisms. However, analyzing sequencing data from microbial mixtures with a high dynamic range of relative abundance of strains, close relatedness, and repetitive genomic content amongst members can vastly complicate the genome assembly process of individual microorganisms and strains. Furthermore, efficient assembly requires high fidelity sequencing reads to avoid ambiguities. We previously developed a method that captures long-range molecular origin information from kilobase-long genomic fragments by a process of DNA barcoding that we called transposase enzyme-linked long read sequencing (TELL-seq) developed by Universal Sequencing Technologies (UST)¹. TELL-seq barcoded fragments can be sequenced with instruments that process short reads (i.e., high-fidelity sequencing). Here, we show that integration of TELL-seq data with a computational pipeline that combines de novo genome assembly (Tell-Link) with taxonomic classification and abundance estimation (Tell-TaxContigs) provides highly accurate metagenomic analyses. We show how the application of Tell-Link and Tell-TaxContigs on sequencing data generated from commercially available microbial mixture standards results in genome assemblies with contiguities larger than 1Mbp (N50), and highly accurate classification and relative abundance estimation for organisms at 0.18% or greater relative abundance, respectively. Therefore, Tell-Link, in combination with genome binning software (e.g. metabat2²) provides highly contiguous and high-fidelity genome assemblies of abundant organisms in a metagenomic sample. Tell-TaxContigs classifies contigs and unassembled reads with BLASTn and using a deep learning approach resolves ambiguities, rules out false positive classifications, and accurately estimates relative abundances of classified species. This approach has an average margin of error of lower than 1% in enumerating relative abundance for the microbial mixture standards tested.

How Does UST TELL-seq Work?

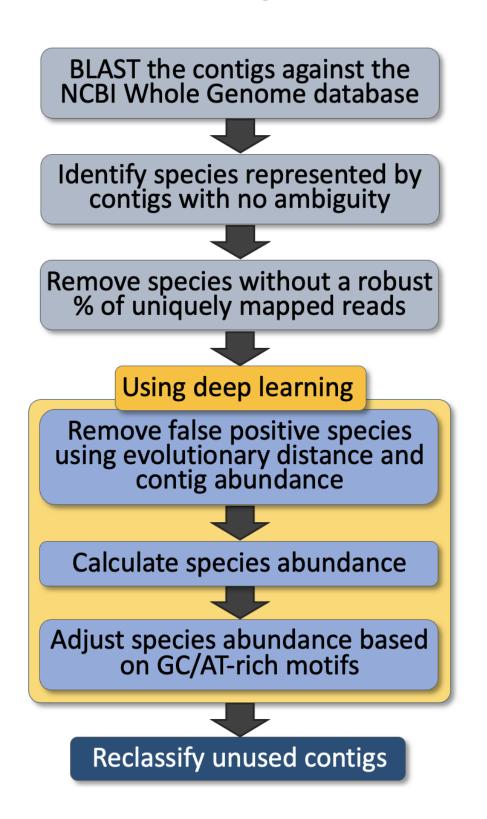
Transposase Enzyme Linked Long-Read Sequencing



TELL-seq Library Workflow is Simple & Scalable



Tell-TaxContigs Overview



1. Metagenomic Assembly & Genome Binning

Detailed metagenomic assembly and genome binning results

	Sample (color-coded)			Even	Staggered / Stagg	rered-deep		
	Species Name	Contigs	Length (Mbp)	N50 (Mbp)	Length/N50	Completeness	Contamination	Quality draft
	Staphylococcus epidermidis	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA
y	Streptococcus mutans	NA 6 NA	NA 1.87 NA	NA 1.33 NA	NA 1.40 NA	NA 92.77 NA	NA 0.00 NA	NA Yes NA
	Porphyromonas gingivalis	10 15 10	2.12 2.07 2.13	1.23 2.02 1.17	1.72 1.02 1.82	98.82 98.79 99.29	0.00 0.00 0.00	Yes Yes Yes
,	Escherichia coli	3 11 4	4.51 4.41 4.52	4.50 4.38 4.50	1.00 1.01 1.00	99.87 98.32 99.97	0.04 0.04 0.04	Yes Yes Yes
	Rhodobacter sphaeroides	23 11 5	4.37 4.39 4.41	3.03 3.11 3.10	1.44 1.41 1.42	94.11 97.36 99.24	0.15 0.15 0.15	Yes Yes Yes
	Staphylococcus aureus	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA Na NA	NA NA NA
	Streptococcus agalactiae	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA	NA NA NA
	Bacillus cereus	7 173 NA	5.33 1.19 NA	2.72 2.07 NA	1.96 5.75 NA	97.61 95.57 NA	0.33 193.23 NA	Yes No NA
	Clostridium beijerinckii	42 110 NA	5.92 5.68 NA	2.64 1.37 NA	2.24 4.16 NA	99.19 96.77 NA	2.42 2.42 NA	Yes Yes NA
ic	Pseudomonas aeruginosa	6 262 3	6.29 5.53 6.29	6.27 1.26 6.28	1.00 4.39 1.00	98.40 86.96 99.68	0.45 0.92 0.45	Yes No Yes
v:	Lactobacillus gasseri	2 NA NA	1.80 NA NA	1.79 NA NA	1.00 NA NA	98.36 NA NA	0.00 NA NA	Yes NA NA
,	Helicobacter pylori	1 NA 14	1.59 NA 1.59	1.59 NA 1.55	1.00 NA 1.03	99.09 NA 98.20	0.00 NA 0.00	Yes NA Yes
oly	Acinetobacter baumannii	15 NA 5	7.93 NA 3.83	2.05 NA 3.81	3.86 NA 1.00	100.00 NA 99.45	175.51 NA 0.27	No NA Yes
re.	Neisseria meningitidis	8 NA 48	2.02 NA 1.84	1.99 NA 1.69	1.01 NA 1.09	98.23 NA 92.14	0.19 NA 0.19	Yes NA Yes
ım	Cutibacterium acnes	1 NA 75	2.48 NA 2.25	2.48 NA 2.01	1.00 NA 1.12	98.90 NA 79.07	0.00 NA 0.00	Yes NA No
to	Enterococcus faecalis	3 NA NA	2.68 NA NA	2.67 NA NA	1.00 NA NA	98.89 NA NA	0.00 NA NA	Yes NA NA
ım	Bacteroides vulgatus	7 NA NA	4.82 NA NA	3.12 NA NA	1.55 NA NA	98.45 NA NA	0.19 NA NA	Yes NA NA
SS.	Deinococcus radiodurans	5 NA NA	3.03 NA NA	2.61 NA NA	1.16 NA NA	99.04 NA NA	0.21 NA NA	Yes NA NA
	Actinomyces odontolyticus	2 NA NA	2.36 NA NA	2.36 NA NA	1.00 NA NA	97.17 NA NA	0.47 NA NA	Yes NA NA
	Bifidobacterium adolescentis	1 NA NA	1.99 NA NA	1.99 NA NA	1.00 NA NA	97.15 NA NA	0.00 NA NA	Yes NA NA

Metagenomic assembly summary

Microbial Mix	Even	Staggered	Staggered-deep
Sequencing depth	59M	98M	700M
True positives (out of 20)	16	7	8
Unclassified	1	0	3
Quality draft	15	5	7
Avg. contiguity	1.52	2.73	1.64

Notes on metagenomic assembly:

Contiguity is defined as assembly length divided by N50 score.

Quality draft denotes medium quality draft assembly according to Bowers et al. (2017)³ minimum information of MAGs.

2. Taxonomic Classification & Abundance Estimation

Detailed classification results

Bacterial strains	True Abundance		Tell-TaxContigs Estimated Abundance			
Species Name	Even	Staggered	Even	Staggered	Staggered-deep	
Staphylococcus epidermidis	5%	18.00%	5.13%	17.28%	15.63%	
Streptococcus mutans	5%	18.00%	6.23%	14.45%	14.68%	
Porphyromonas gingivalis	5%	18.00%	5.44%	16.26%	15.82%	
Escherichia coli	5%	18.00%	4.47%	20.67%	21.79%	
Rhodobacter sphaeroides	5%	18.00%	4.91%	18.18%	16.12%	
Staphylococcus aureus	5%	1.80%	5.62%	1.57%	1.49%	
Streptococcus agalactiae	5%	1.80%	5.68%	1.56%	2.14%	
Bacillus cereus	5%	1.80%	4.33%	2.17%	2.18%	
Clostridium beijerinckii	5%	1.80%	5.67%	1.57%	1.33%	
Pseudomonas aeruginosa	5%	1.80%	5.49%	1.61%	0.91%	
Lactobacillus gasseri	5%	0.18%	4.15%	0.23%	0.11%	
Helicobacter pylori	5%	0.18%	4.08%	0.24%	0.10%	
Acinetobacter baumannii	5%	0.18%	4.46%	0.21%	0.15%	
Neisseria meningitidis	5%	0.18%	3.09%	0.65%	0.10%	
Cutibacterium acnes	5%	0.18%	3.67%	0.31%	0.10%	
Enterococcus faecalis	5%	0.02%	6.08%	NA	0.02%	
Bacteroides vulgatus	5%	0.02%	6.79%	NA	0.05%	
Deinococcus radiodurans	5%	0.02%	4.55%	NA	0.04%	
Actinomyces odontolyticus	5%	0.02%	3.68%	NA	NA	
Bifidobacterium adolescentis	5%	0.02%	5.87%	NA	NA	

Classification summary

Microbial Mix	Even	Staggered	Staggered-deep
Total sequencing reads	59M	98M	700M
True positives (out of 20)	20	15	18
False positives	0	0	2
Avg. RA error	0.83%	0.72%	0.91%
Max. RA error	1.91%	3.55%	3.79%

References

Chen, Z et al. 2020. Genome Research 30: 898-909.
 Dongwan et al. 2019. PeerJ, 7:e7359
 Bowers et al. 2017. Nat. Biotechnol. 35:725–731
 Wick et al. 2017. PLoS Comput Biol 13(6): e1005595.

Conclusions. UST TELL-seq allows for long-range information to be integrated into short-read sequencing projects. This offers several advantages to standard whole metagenome shotgun sequencing projects. We have shown that de novo assembly of TELL-seq barcoded Illumina reads results in highly contiguous, high fidelity binned genome assemblies with N50 scores in the Mbp range. Achieving this level of contiguity and fidelity normally requires the hybridization of high-fidelity short reads with error-prone long-read technologies, ideally from the same DNA extraction⁴. Using UST TELL-seq is a simpler process that can achieve similar results, with only a single sequencing source. As a result, the highly contiguous and accurate contigs generated with TELL-seq can be used for various metagenomics downstream analysis steps, such as taxonomic classification and relative abundance estimation, with relatively low margins of error, as we have demonstrated with Tell-TaxContigs. Tell-TaxContigs will be released publicly later this year.

