Article Addendum Cannabinoids act as necrosis-inducing factors in *Cannabis sativa*

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Cannabis sativa is well known to produce unique secondary metabolites called cannabinoids. We recently discovered that Cannabis leaves induce cell death by secreting tetrahydrocannabinolic acid (THCA) into leaf tissues. Examinations using isolated Cannabis mitochondria demonstrated that THCA causes mitochondrial permeability transition (MPT) though opening of MPT pores, resulting in mitochondrial dysfunction (the important feature of necrosis). Although Ca²⁺ is known to cause opening of animal MPT pores, THCA directly opened Cannabis MPT pores in the absence of Ca²⁺. Based on these results, we conclude that THCA has the ability to induce necrosis though MPT in Cannabis leaves, independently of Ca²⁺. We confirmed that other cannabinoids (cannabidiolic acid and cannabigerolic acid) also have MPT-inducing activity similar to that of THCA. Moreover, mitochondria of plants which do not produce cannabinoids were shown to induce MPT by THCA treatment, thus suggesting that many higher plants may have systems to cause THCA-dependent necrosis.

Cannabis sativa produces unique secondary metabolites consisting of alkylresorcinol and monoterpene groups.¹ These metabolites called cannabinoids are well known to show a variety of interesting pharmacological activities including psychoactive effect and analgesic effect. Therefore, cannabinoids have attracted a great deal of attention, whereas why *C. sativa* produces such metabolites has long remained unclear. However, we have recently obtained evidences indicating the physiological function of THCA in Cannabis leaves.²

We discovered that THCA is stored in capitate-sessile glands on Cannabis leaves and that secretion of this cannabinoid into leaf tissues causes cell death. When the properties of THCA were examined using cultured Cannabis cells, this cannabinoid induced plasmamembrane shrinkage and DNA degradation. These responses

Previously published online as a *Plant Signaling & Behavior* E-publication: http://www.landesbioscience.com/journals/psb/article/7011 are regarded as the features of apoptotic cells, but were not suppressed by apoptosis inhibitors. In contrast, the necrosis inhibitor cyclosporine A significantly inhibited both plasmamembrane shrinkage and DNA degradation in Cannabis cells. Therefore, we assumed that THCA induces necrotic cell death in Cannabis cells and leaves.

Necrosis in plants and animals is usually triggered by MPT though opening of MPT pores.^{3,4} MPT is known to cause mitochondrial dysfunction by mitochondrial swelling and loss of mitochondrial membrane potential ($\Delta \Psi m$),^{5,6} and we also confirmed that THCA induces mitochondrial swelling and $\Delta \Psi m$ reduction in mitochondria isolated from Cannabis cells and that pretreatment with cyclosporine A inhibits both responses. Based on these evidences, we concluded that THCA has the activity to induce MPT-dependent necrosis.

As described above, MPT pores play an important role in necrosis induction, whereas the mechanism of their opening in higher plants has not been fully understood. However, binding of cyclophilin D (a protein present in mitochondrial matrix) to MPT pores is shown to be essential for their opening in plants as well as animal.⁷⁻⁹ In animal mitochondria, Ca²⁺ mediates this binding reaction, leading to opening of MPT pores. Wheat mitochondria are also shown to undergo swelling through opening of MPT pores in response to Ca²⁺,⁹ whereas MPT pores of oats,¹⁰ Arabidopsis thaliana¹¹ and C. sativa² do not open by Ca²⁺ treatment. In contrast, THCA catalyzed opening of Cannabis MPT pores in the absence of Ca²⁺, suggesting that THCA directly mediates binding of cyclophilin D to MPT pores (Fig. 1). In addition, we have now confirmed that THCA causes dysfunction though MPT in mitochondria of plants (rice, soybean, A. thaliana and Scutellaria baicalensis) lacking cannabinoid-producing ability (data not shown). Therefore, many higher plants may have the systems to induce THCA-dependent necrosis.

Furthermore, we investigated whether other cannabinoids and their related compounds can mediate MPT in Cannabis mitochondria. When the MPT-inducing activity of each sample was measured by monitoring both $\Delta\Psi$ m reduction (Fig. 2) and mitochondrial swelling (data not shown), we confirmed that cannabinoids tested here (cannabidiolic acid and cannabigerolic acid) possess the activities similar to those of THCA. On the other hand, olivetolic acid (the akylresorcinol moiety of cannabinoid) and geraniol (the monoterpene moiety of cannabigerolic acid) showed neither $\Delta\Psi$ m reduction nor mitochondrial swelling (Fig. 2). These results suggested that the structures (cannabinoid skeleton) where monoterpene and olivetolic acid are coupled to each other seem essential for opening of MPT pores. Therefore, we assumed that plant cyclophilin D and MPT pores have the cannabinoid-binding site.

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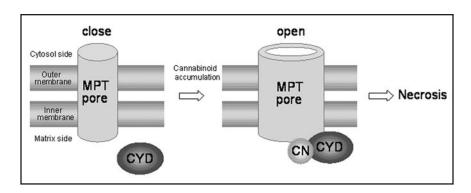


Figure 1. A model depicting the opening mechanism of MPT pores in mitochondria. CYD, cyclophilin D; CN, cannabinoid.

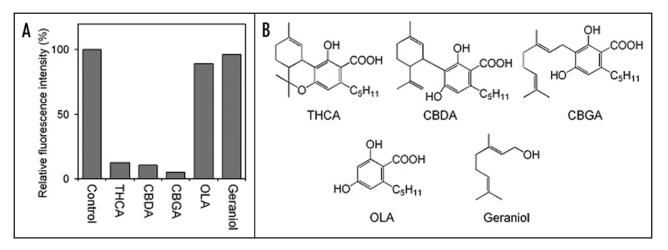


Figure 2. Change of $\Delta \Psi m$ by treatment with various compounds (A) and their chemical structures (B). The isolated mitochondria were stained with the $\Delta \Psi m$ -indicating reagent (tetramethylrhodamine methylester, TMRM) and then incubated with 200 μ M of each compound for 60 min. The intensity of TMRM fluorescence was measured using a fluorescence microplate reader. A decrease of the fluorescence intensity indicates $\Delta \Psi m$ reduction. CBDA, cannabidiolic acid; CBGA, cannabigerolic acid; OLA, olivetolic acid.

Plant cell death is shown to participate in important physiological responses such as leaf senescence, somatic embryogenesis and defense against microbial pathogens.^{12,13} Based on its induction mechanism, plant cell death is largely classified into apoptosis and necrosis. Although the molecular mechanism of apoptosis has been extensively investigated, there is little precise information on plant necrosis. However, our study would provide important insight into necrosis-inducing mechanisms in higher plants.

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