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Cat#: CR120C (96 tests) For Order and Inquiries, please contact Calbiotech Inc., 1935 Cordell Ct., El Cajon, CA 92020 Tel (619) 660-6162, Fax (619) 660-6970, www.calbiotech.com



# HIGH SENSITIVITY C-REACTIVE PROTEIN (CRP) ELISA

Catalog No.: CR120C (96 tests)

INTENDED USE

For Research Use Only. Not for use in diagnostic procedures.

	MATERIALS PROVIDED	96 tests
1.	Microwells coated with CRP MAb	12x8x1
2.	CRP Standard: 6 vials (ready to use)	0.7ml
3.	CRP Enzyme Conjugate: 1 bottle (ready to use)	12 ml
4.	TMB Substrate: 1 bottle (ready to use)	12ml
5.	Stop Solution: 1 bottle (ready to use)	12ml
6.	Sample Diluent	50 ml
7.	20X Wash concentrate: 1 bottle	25ml

## MATERIALS NOT PROVIDED

- 1. Distilled or deionized water
- 2. Precision pipettes
- 3. Disposable pipette tips
- 4. ELISA reader capable of reading absorbance at 450nm
- 5. Absorbance paper or paper towel
- 6. Graph paper

## STORAGE AND STABILITY

- 1. Store the kit at 2 8° C.
- 2. Keep microwells sealed in a dry bag with desiccants.
- 3. The reagents are stable until expiration of the kit.
- 4. Do not expose test reagents to heat, sun, or strong light.

#### CRP rev.01

#### WARNINGS AND PRECAUTIONS

1. Potential biohazardous materials:

The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent. These reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories" 1984.

- 2. This test kit is designed for research use only.
- 3. Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.
- 5. The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.
- 6. It is recommended that standards, control and serum samples be run in duplicate.
- 7. Optimal results will be obtained by strict adherence to this protocol. Accurate and precise pipetting, as well as following the exact time and temperature requirements prescribed are essential. Any deviation from this may yield invalid data.

#### SPECIMEN COLLECTION HANDLING

- 1. Collect blood specimens and separate the serum immediately.
- 2. Typically, specimens may be stored refrigerated at (2-8°C) for 5 days. If storage time exceeds 5 days, store frozen at (-20°C) for up to one month.
- 3. Avoid multiple freeze-thaw cycles.
- 4. Prior to assay, frozen sera should be completely thawed and mixed well.
- 5. Do not use grossly lipemic specimens.

#### **REAGENTS PREPARATION**

1X Wash Buffer: Prepare 1X Wash buffer by adding the contents of the bottle (25 ml, 20X) to 475 ml of distilled or deionized water. Store at room temperature (20-25°C).

## ASSAY PROCEDURE

Prior to assay, allow reagents to stand at room temperature (20-25°C). Gently mix all reagents before use.

- 1. Place the desired number of coated strips into the holder
- Dilute patient samples and controls 1:100 by adding 5 µl of samples to 495 µl of sample Diluent (STANDARDS ARE READY TO USE).
- 3. Dispense 10 µL of standard, diluted samples and controls into the appropriate wells
- 4. Add 100  $\mu l$  of enzyme conjugate to all wells. Tap the holder to remove air bubbles from the liquid and mix well.
- 5. Incubate for 60 minutes at room temperature (20-25°C).
- 6. Remove liquid from all wells. Wash wells three times with 300  $\mu$ l of 1X wash buffer. Blot on absorbent paper towels.
- 7. Add 100 µl of TMB substrate to all wells.
- 8. Incubate for 15 minutes at room temperature.

- 9. Add 50 µl of stop solution to all wells. Shake the plate gently to mix the solution.
- 10. Read absorbance on ELISA Reader at 450 nm within 15 minutes after adding the stopping solution.

### **CALCULATION OF RESULTS**

The standard curve is constructed as follows:

- 1. Check CRP standard value on each standard vial. This value might vary from lot to lot. Make sure you check the value on every kit. See example of the standard attached.
- 2. To construct the standard curve, plot the absorbance for the CRP standards (vertical axis) versus the CRP standard concentrations (horizontal axis) on a linear graph paper. Draw the best curve through the points.
- 3. Read the absorbance for controls and each unknown sample from the curve. Record the value for each control or unknown sample.
- 4. The obtained values of the patient samples and control sera should be multiplied by the dilution factor of 100 to obtain CRP results in mg/l.
- 5. Patient samples with CRP concentrations greater than 10 mg/l should be further diluted 10-fold after the initial 100-fold dilution (total dilution 1:1,000), and the final CRP values should be multiplied by 1,000 to obtain CRP results in mg/l.

#### Example of a Standard Curve

	OD 450 nm	Conc. mg/L
Std 1	0.02	0
Std 2	0.23	0.005
Std 3	0.49	0.01
Std 4	1.01	0.025
Std 5	1.66	0.05
Std 6	2.40	0.1