

Anti-Peripherin Immunocytofluorescence Protocol

Catalog #: 1630-PER

Species: mouse

Tissue/Cells: Mixed fibroblasts and PC12 cells

Fixation: 3.7% Neutral buffered formalin for 1 minute

Antibody incubation: Primary Antibody- 4C, overnight

Secondary Antibody- RT, 3 hours or 4C, overnight

Materials Required

- ✓ **Fixative:** 3.7% Neutral buffered formalin (10mls of 37% Fisher formalin, 90mls of 1xPBS)
 - ✓ **methanol:** chilled to -20C in a well-sealed container
 - ✓ **1X PBS:** 137 mM NaCl, 28 mM Na₂HPO₄, 5.4 mM KCl, 2.9 mM KH₂PO₄, pH 7.6
 - ✓ **Blocking/Incubation buffer:** 1X PBS with 1% goat serum
 - ✓ **Secondary Antibody:** example used is Goat-Anti-Mouse Alexa 647 from ThermoFisher
 - ✓ **Mounting media:** ThermoFisher Histomount [Cat #: 008030](#)
 - ✓ **Glass cover slip**
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Before you begin

This protocol is intended for adherent cells grown in 6 well plates or in 35mm dishes using a glass coverslip placed on top of the cells. These work well with most immunofluorescence microscopes using lenses up to 100X magnification.

Protocol

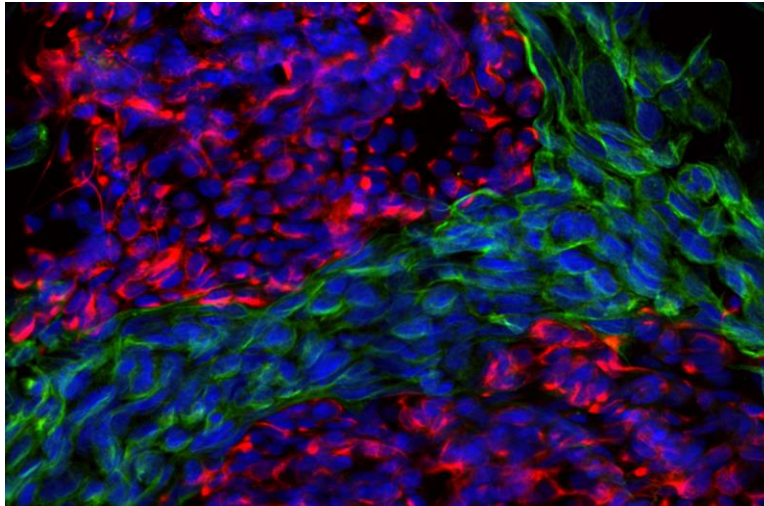
1. Draw off culture medium with aspirator and add 1 ml of 3.7 % formalin in PBS solution to the dish. Incubate at room temperature for 1 minute.
Tech Tip:
 - a. To reduce background you can add 0.1% Tween 20 to PBS here and all subsequent steps. Use with caution as it may extract your antigen or wash cells off the dish.
2. Remove the neutral buffered formalin and add 1 ml of cold methanol. Incubate for 1 minute or less.
3. Remove methanol and add 1 ml of PBS, not letting the specimen dry out.
4. Remove PBS and add blocking buffer. Incubate for 30 minutes at room temperature.
5. Rinse cells with PBS 3 times, in 10 minute intervals.
6. Dilute Anti-Peripherin (Cat. # 1630-PER) to 1:500 in incubation buffer. Incubate cells for 1 hour at room temperature or overnight at 4C.
7. Remove primary antibody and wash with 1 ml PBS 3 times, in 10 minute intervals.
8. Dilute secondary antibody in incubation buffer per manufacturer's recommendation. Incubate cells for 3 hours at room temperature or overnight at 4C.

Tech Tip:

- a. Alexa Fluor 647 dye diluted 1:2000 was used to produce image below.
9. Remove secondary antibody and wash with PBS 3 times, in 5-10 minute intervals.
10. Apply mounting medium onto dish and gently place glass cover slip before viewing under the microscope.

Tech Tip:

- a. Any mounting media can be used, for this protocol ThermoFisher Histomount medium was used. [Cat #: 008030](#).



Immunofluorescence of mixed fibroblasts and PC12 cells labeled with anti-peripherin (cat # 1630-PER, red, 1:500) and anti-vimentin (cat # 2105-Vim, green, 1:10,000). The blue is Hoechst staining nuclear DNA. Peripherin is labeled in only the PC12 cells cytoplasmic filaments and vimentin is only labeled in the fibroblasts intermediate filaments where peripherin is not present.