# COVID-19 Antigen Nasal Test Kit



# 1. INTENDED USE

The COVID-19 Antigen Nasal Test Kit is an in vitro immunoassay. The assay is for the direct and qualitative detection of SARS-CoV-2 viral nucleocapsid protein from nasal secretions collected from symptomatic individuals suspected of COVID-19. The kit is in aid of diagnosis of COVID-19 and is to be used within 7 days of onset of symptoms.

The COVID-19 Antigen Nasal Test Kit is intended for use by trained healthcare professionals. For laboratory and point of care use. This assay is not intended for home testing (or self-testing).

Results are for the identification of SARS-CoV-2 viral nucleoprotein antigen. Antigen is generally detectable in nasal samples during the acute phase of infection. Positive results indicate the presence of viral antigens, but clinical correlation with patient history and other diagnostic information is necessary to determine infection status. Positive results do not rule out bacterial infection or co-infection with other viruses. The agent detected may not be the definite cause of disease. Laboratories are required to report all positive results to the appropriate public health authority.

Negative results do not rule out SARS-CoV-2 infection and should not be used as the sole basis for treatment or patient management decisions, including infection control decisions. Negative results should be considered in the context of a patient's recent exposures, history and the presence of clinical signs and symptoms consistent with COVID-19, and confirmed with a molecular assay, if necessary for patient management.

# 2. PRINCIPLE

The COVID-19 Antigen Nasal Test Kit detects SARS-CoV-2 viral antigens through visual interpretation of color development. Anti-SARS-CoV-2 antibodies are immobilized at the test region of the nitrocellulose membrane. Anti-SARS-CoV-2 antibodies conjugated to colored particles are immobilized on the conjugated pad.

The nasal secretion collected by the intended user, is supposed to be mixed with the extraction buffer, which is individually packed in the kit.

During testing, target antigens, if present in the nasal secretions, will be released into the extraction buffer. As the specimen migrates along the strip by capillary action and then interacts with reagents on the sample pad, the target antigens will bind to anti-SARS-CoV-2 antibodies on the conjugate pad. Consequently, the antigen-antibody complex will be captured by the anti-SARS-CoV-2 antibodies immobilized at the test region. Excess colored particles will be captured at the control region of the NC membrane.

The presence of a colored band in the test region indicates a positive result for the SARS-CoV-2 viral antigens, while its absence indicates a negative result. A colored band at the control region serves as a procedural control, generally indicating that a proper volume of specimen has been added and membrane wicking is working.

# 3. MATERIALS

#### Materials Provided

- · 20 Individual packaged test
- · 1 Package insert

#### **Optional Materials**

• External Negative and Positive control (Available upon request)

#### **External Negative and Positive Control**

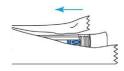
Negative control is lyophilized GCS and positive control is lyophilized SARS-CoV-2 NCP. Each control vial with 50 µL purified water. Insert the sample collector of device into the reconstituted control solution. Rotate the sample collector several times, ensuring that all solution sufficiently absorbed onto the sample collector. Store reconstituted controls at 4°C.

## Materials Required but Not Provided

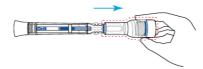
· Clock, timer, or stopwatch

# 4. TEST PROCEDURE

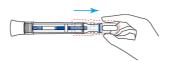
- Bring devices, reagents and specimens and/or controls to room temperature (15~30°C) before use.
- Remove the test device from its packing. Label the device with the patient's identification. For the best results, the assay should be performed within one hours.



3. Take the test device out of the tube with extraction buffer.



Remove the protector.



5. Gently insert the sample collector until resistance is met (about 1-2 cm into the nostril). Rotate the collector five times against the nasal wall and remove from the nostril. Repeat the sample collection procedure for the other nostril to ensure that sufficient specimen be collected from both nasal cavities.

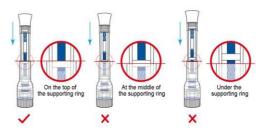


#### NOTE:

- 1). It is important to obtain as much secretion as possible.
- This may feel uncomfortable. Do not insert the collector any deeper if you feel strong resistance. Children aged 2-15 years should be tested by an adult (18+ years old).
- Place the test device vertically into the extraction tube until the top edge of the extraction tube reach the top of the supporting ring.



NOTE: When placing the test device vertically into the extraction tube, the edge of the extraction tube must reach the top of the supporting ring. If not, this may lead to lateral flow failure, resulting in an incorrect result or invalid result.



Read the results at 15 minutes.

# 5. RESULT INTERPRETATION



POSITIVE: Two colored bands appear on the membrane. One band appears in the control region (C), and another band appears in the test region (T)



NEGATIVE: Only one colored band appears in the control region (C). No apparent colored band appears in the test region (T).



INVALID: Control band fails to appear.

Results from any test which has not produced a control band at the specified read time must be discarded. Please review the procedure and repeat with a new test. If the problem persists, discontinue using the kit immediately and contact your local distributor.

## NOTE:

- The color intensity in the test region (T) may vary depending on the concentration of analytes present in the specimen. Note that this is a qualitative test only, and cannot determine the concentration of analytes in the specimen.
- Insufficient specimen volume, incorrect operating procedure or expired tests are the most likely reasons for control band failure.

# 6. PRECAUTIONS

- 1. For In Vitro Diagnostic Use Only.
- Each device is for single use only and cannot be reused.
- Caution should be taken when inserting the sample collector into the nasal cavity.
- DO NOT ingest.
- 5. Do not use kit or components beyond the expiration date.
- Do not puncture the membrane in the extraction tube before testing.
- Read the instructions for use before use. The instructions for use must be read carefully and followed.
- 8. Do not test or components after their expiration date use.
- The device contains material of animal origin and should be handled as a potential biohazard. Do not use if pouch is damaged or open.
- 10. The test components are packed in foil pouches to protect them from moisture during storage. Check each foil pouch before opening it. Do not use any component that has holes in the film or the pouch has not been completely sealed. Improper storage of test items or components can lead to incorrect results.
- Do not use the kit when any component including test device, protector, extraction buffer, package insert is missing.
- 12. All patient specimens should be handled and discarded as if they are biologically hazardous. All specimens must be mixed thoroughly before testing to ensure a representative sample prior to testing.
- 13. Failure to bring specimens and reagents to room temperature before testing may decrease assay sensitivity. Inaccurate or inappropriate specimen collection, storage, and transport may yield false negative test smalls.
- 14. The buffer components include salts and surfactants, the preservative is sodium azide and water is the solvent. Avoid skin or eyes contact with buffer.
- 15. Avoid eye, skin and mucous membrane contact with the buffer. In the

- event of contact with buffer, rinse with plenty of water.
- 16. Do not use this test on anyone under 2 years of age.
- 17. Keep out of the reach of children. Small test components can pose a choking hazard
- 18. Use only the supplied test components. Do not replace the buffer with any other liquid
- 19. Keep the collector clean. Do not touch the collector and make sure it does not touch any surfaces before use.
- 20. If you have a nose piercing, dab the other nostril. If pierced on both sides, remove the piercing on one side before wiping it off.
- 21. If infection with SARS-CoV-2 is suspected based on current clinical and epidemiological screening criteria recommended by public health authorities, specimens should be collected with appropriate infection control precautions and sent to state or local health departments for testing
- 22. Viral isolation in cell culture and initial characterization of viral agents recovered in cultures of SARS-CoV-2 specimens are NOT recommended, except in a BSL3 laboratory using BSL3 work practices.
- 23. This test is for human use only.
- 24. There may be false negatives due to new unknown variants of SARS-Cov-2 prior to validation data available
- 25. COVID-19 Antigen Nasal Test Kit could detect SARS-CoV-2 variant alpha, beta, gamma, kappa and delta.

# 7. QUALITY CONTROL

#### Internal Procedural Controls

The COVID-19 Antigen Nasal Test Kit has built-in (procedural) controls. Each test device has an internal standard zone to ensure proper sample flow. The user should confirm that the colored band located at the "C" region is present before reading the result.

#### **External Positive and Negative Controls**

Good laboratory practice suggests testing positive and negative external controls to ensure that the test reagents are working and that the test is correctly performed.

# 8. LIMITATIONS OF THE TEST

- 1. The COVID-19 Antigen Nasal Test Kit is for professional in vitro diagnostic use, and should only be used for the qualitative detection of SARS-CoV-2 antigen. The intensity of color in a positive band should not be evaluated as "quantitative or semi-quantitative".
- Both viable and nonviable SARS-CoV-2 viruses are detectable with The COVID-19 Antigen Nasal Test Kit.
- 3. As with all diagnostic tests, a definitive clinical diagnosis should not be based on the results of a single test, but should only be made by the physician after all clinical and laboratory findings have been evaluated.
- Failure to follow the TEST PROCEDURE and RESULT INTERPRETATION may adversely affect test performance and/or invalidate the test result.
- Results obtained with this assay, particularly in the case of weak test lines that are difficult to interpret, should be used in conjunction with other clinical information available to the physician.
- 6. Negative results do not preclude SARS-CoV-2 infection and should be confirmed via molecular assay.

- Store The COVID-19 Antigen Nasal Test Kit at 2~30°Cwhen not in use.
- DO NOT FREEZE.
- · Kit contents are stable until the expiration dates marked on their outer packaging and container

# 10.PERFORMANCE

## Analytical Sensitivity (Limit of Detection):

The limit of detection was determined with a quantified SARS-CoV-2 virus and has been evaluated at 1x102.4TCID50/mL.

The limit of detection was also determined with recombinant SARS-CoV-2 nucleoprotein and has been evaluated at 370 pg/mL.

A total of 508 clinical specimens were collected to verify the performance of COVID-19 Antigen Nasal Test Kit. There were 106 positive specimens from the individuals who were suspected of COVID-19 within 7 days of symptom and 402 negative clinical specimens confirmed by RT-PCR. The results were summarized below:

Table: COVID-19 Antigen Nasal Test Kit vs. RT-PCR

		RT-PCR		
		Positive	Negative	Total
COVID-19 Antigen	Positive	104	1	105
Nasal Test Kit	Negative	2	401	403
Total		106	402	508

Relative Sensitivity: 98.1 % (93.4% ~ 99.5%)\*

Relative Specificity: 99.8 % (98.6% ~ 100.0%)\*

Overall Agreement: 99.4 % (98.3% ~ 99.8%)\*

\*95% Confidence Interval

The table below shows the positive results broken down by days since

Days Since Symptom Onset	Cumulative RT-PCR Positive (+)	Cumulative COVID-19 Antigen Rapid Test Device Positive(+)	PPA	95%CI
0	5	5	100.0%	56.6%-100.0%
1	15	15	100.0%	79.6%-100.0%
2	28	28	100.0%	87.9% - 100.0%
3	38	38	100.0%	90.8% - 100.0%
4	50	49	98.0%	89.5%-99.6%
5	62	60	96.8%	89.0%-99.1%
6	69	67	97.1%	90.0%-99.2%
7	76	74	97.4	90.9%-99.3%
NA	106	104	98.1%	93.4%-99.5%

# Cross Reactivity:

Cross reactivity with the following organisms has been studied. Samples positive for the following organisms were found negative when tested with

the COVID-19 Antigen Nasal Test Kit

Viruses	Con.	Viruses	Con.
Adenovirus 1	1.0×10 <sup>5</sup> TCID50/mL	HCoV-229E	1.0×10 <sup>5</sup> TCID50/mL
Adenovirus 2	1.0×10 <sup>5</sup> TCID50/mL	HCoV-OC43	1.0×10 <sup>5</sup> TCID50/mL
Adenovirus 3	1.0×10 <sup>5</sup> TCID50/mL	HCoV-NL63	1.0×10 <sup>5</sup> TCID50/mL
Adenovirus 4	1.0×10 <sup>5</sup> TCID <sub>50</sub> /mL	MERS-coronavirus	ol.0e×10mî cpis/L
Adenovirus 5	1.0×10 <sup>5</sup> TCID <sub>50</sub> /mL	SARS-coronavirus	col.0ex1/0m2î pis L
Adenovirus 7	1.0×10 <sup>5</sup> TCID₅₀/mL	Human metapneumovirus	1.0×10 <sup>5</sup> TCID <sub>50</sub> /mL
Adenovirus 55	1.0×10 <sup>5</sup> TCID50/mL	Influenza A (H1N1)pdm09	1.0×10 <sup>5</sup> TCID50/mL
Epstein-Barr virus	5.0×10 <sup>3</sup> copies/mL	Influenza A (H3N2)	1.0×10 <sup>5</sup> TCID50/mL

Enterovirus	1.0×10⁵	Influenza B	1.0×10⁵
EV70	TCID50/mL	Victoria lineage	TCID50/mL
Enterovirus	1.0×10 <sup>5</sup>	Influenza B	1.0×10 <sup>5</sup>
EV71	TCID50/mL	Yamagata lineage	TCID50/mL
Enterovirus	1.4×10 <sup>5</sup>	Norovirus	1.0×10 <sup>5</sup>
A16	TCID50/mL		copies/mL
Enterovirus	1.0×10 <sup>5</sup>	Parainfluenza virus	1.0×10 <sup>5</sup>
A24	TCID50/mL	1	TCID50/mL
Enterovirus B1	1.1×10⁵	Parainfluenza virus	1.0×10 <sup>5</sup>
Eliterovirus B1	TCID50/mL	2	TCID50/mL
Echovirus 6	1.0×105	Parainfluenza virus	1.0×10 <sup>5</sup>
	TCID50/mL	3	TCID50/mL
Respiratory	1.0×10 <sup>5</sup>	Parainfluenza virus	1.0×10 <sup>5</sup>
syncytial virus	TCID <sub>50</sub> /mL	1 arammuchza virus 4	TCID <sub>50</sub> /mL
A	I CID50/IIIL	4	I CID50/IIIL
Respiratory	1.0 ⋈ 0⁵		1.0 ⋈ 05
syncytial virus		Rhinovirus B52	
В	TCID50/mL		TCID50/mL
Rhinovirus	1.0×10 <sup>5</sup>	Human coronavirus	1.0×105
A30	TCID50/mL	HKU1	TCID50/mL
Bacterium	Con.	Bacterium	Con.
Bacterium Bordetella	1.8×10 <sup>6</sup>	Mycobacterium	6.32×10 <sup>6</sup>
Bordetella	1.8×10 <sup>6</sup>	Mycobacterium	6.32×10 <sup>6</sup>
Bordetella parapertussis	1.8×10 <sup>6</sup> cfu/mL	Mycobacterium tuberculosis	6.32×10 <sup>6</sup> cfu/mL
Bordetella parapertussis Bordetella	1.8×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup>	Mycobacterium tuberculosis Staphylococcus	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup>
Bordetella parapertussis Bordetella pertussis	1.8×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL	Mycobacterium tuberculosis Staphylococcus aureus	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL
Bordetella parapertussis Bordetella pertussis Candida	1.8×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 1.0×10 <sup>6</sup> cfu/mL	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup>
Bordetella parapertussis Bordetella pertussis Candida albicans	1.8×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 1.0×10 <sup>6</sup>	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL cfu/mL
Bordetella parapertussis Bordetella pertussis Candida albicans Chlamydia	1.8×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 1.0×10 <sup>6</sup> cfu/mL	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup>
Bordetella parapertussis Bordetella pertussis Candida albicans Chlamydia pneumoniae	1.8×10° cfu/mL 2.0×10° cfu/mL 1.0×10° cfu/mL 2.0×10° EB/ml	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup> cfu/mL
Bordetella parapertussis Bordetella pertussis Candida albicans Chlamydia pneumoniae Group C	1.8×10° cftt/mL 2.0×10° cftt/mL 1.0×10° cftt/mL 2.0×10° Eft/mL 2.0×10° EB/ml	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae Streptococcus	6.32×10 <sup>6</sup> cfu/mL 2.0×10 <sup>6</sup>
Bordetella parapertussis Bordetella pertussis Candida albicans Chlamydia pneumoniae Group C Streptococcus Haemophilus	1.8×10° cfu/mL 2.0×310° cfu/mL 1.0×10° cfu/mL 2.0×10° EB/ml 2.0×10° cfu/mL	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae Streptococcus pneumoniae	6.32×10° cfu/mL 2.0×10° cfu/mL 2.0×10° cfu/mL 2.0×10° cfu/mL 2.0×10° cfu/mL 2.0×10°
Bordetella parapertussis Bordetella pertussis Candida albicans Chlamydia pneumoniae Group C Streptococcus Haemophulus influenzae	1.8×10° cft/mL 2.0×10° cft/mL 1.0×10° cft/mL 2.0×10° EB/ml 2.0×10° cft/mL 1.0×10°	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae Streptococcus pneumoniae	6.32×10° cft/mL 2.0×10° cft/mL 2.0×10° cft/mL 2.0×10° cft/mL 2.0×10° cft/mL 2.0×10°
Bordetella parapertussis Bordetella pertussis Candida albicans Chiamydia pneumoniae Group C Streptococcus Haemophilus influenzae Legionella	1.8×10° cfu/mL 2.0×10° cfu/mL 1.0×10° cfu/mL 2.0×10° EB/ml 2.0×10° EM/mL 1.0×10° cfu/mL 2.0×10°	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae Streptococcus pneumoniae Streptococcus pyogenes Pseudomonas	6.32×10° cft/mL 2.0×10°
Bordetella parapertussis Bordetella parapertussis Candida albicans Chlamydia pneumoniae Group C Streptococcus Haemophilus influenzae Legionella pneumophila	1.8×10° cft/mL 2.0×10° cft/mL 1.0×10° cft/mL 2.0×10° EB/ml 2.0×10° cft/mL 1.0×10° cft/mL cft/mL 1.0×10°	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae Streptococcus pneumoniae Streptococcus preumoniae Pseudomonas aeruginosa	6.32×10° cft/mL 2.0×10° cft/mL 2.0×10° cft/mL 2.0×10° cft/mL 2.0×10° cft/mL 2.0×106 cft/mL 2.0×106 cft/mL
Bordetella parapertussis Bordetella pertussis Candida albicans Chiamydia pneumoniae Group C Streptococus Haemophilus influenzae Legionella	1.8×10° cft/mL 2.0×10° cft/mL 1.0×10° cft/mL 2.0×10° EB/ml 2.0×10° cft/mL 1.0×10° cft/mL 2.0×10° cft/mL cft/mL 2.0×10°	Mycobacterium tuberculosis Staphylococcus aureus Staphylococcus epidermidis Streptococcus agalactiae Streptococcus pneumoniae Streptococcus pyogenes Pseudomonas	6.32×10° cft/mL 2.0×10° cft/mL

In silico analysis:

For Pneumocystis jirovecii (PJP), blast results showed none homology exists between the SRAS-COV-2 nucleocapsid protein and Pneumocystis jirovecii

#### **Interfering Substances:**

The following substances, naturally present in respiratory specimens or that may be artificially introduced into the respiratory tract, were evaluated at the concentrations listed below. None of them were found to affect test performance of The COVID-19 Antigen Nasal Test Kit.

			T 6
Substance	Con.	Substance	Con.
3 OTC nasal sprays	10%	Guaiacol glyceryl ether	20mg/mL
3 OTC mouthwashes	10%	Mucin	1%
3 OTC throat drops	10%	Whole blood	4%
4-acetamidophenol	10mg/mL	Mupirocin	250µg/mL
Acetylsalicylic acid	10mg/mL	Oxymetazoline	25μg/mL
Albuterol	10mg/mL	Phenylephrine	10 mg/mL
Chlorpheniramine	5 mg/mL	Phenylpropanolamine	1 mg/mL
Dexamethasone	50μg/mL	Zanamivir	10mg/mL
Dextromethorphan	10μg/mL	Adamantanamine	500 ng/mL
Diphenhydramine	5 mg/mL	Oseltamivir phosphate	10mg/mL
Doxylamine succinate	1 mg/mL	Tobramycin	10mg/mL
Flunisolide	25μg/mL	Triamcinolone	14mg/mL

The COVID-19 Antigen Nasal Test Kit demonstrates the expected test repeatability and reproducibility with three different lots at three different sites in 5 days by three difference operators

#### Hook effect:

The study demonstrated that no false negatives occurred on virus level at 1×106.4TCID50/mL and recombinant SARS-CoV-2 nucleocapsid protein level at 1.48mg/mL.

# 11. LITERATURE REFERENCES

- 1. Forni, D., Cagliani, R., Clerici, M. & Sironi, M. Molecular evolution of human coronavirus genomes. Trends Microbiol. 25, 35-48 (2017).
- 2. Ithete, N. L. et al. Close relative of human Middle East respiratory syndrome coronavirus in bat, South Africa. Emerg. Infect. Dis. 19, 1697-1699 (2013).

# 12. GLOSSARY OF SYMBOLS

re: r	Catalog number	1	Temperature limitation
(II)	Consult instructions for use	LOT	Batch code
[MD]	In vitro diagnostic medical device	8	Use by
-	Manufacturer	ℽ	Contains sufficient for <n> tests</n>
2	Do not reuse	EC PEP	Authorized representative in the European Community
(€	CE marking according to IVI	) Medica	Devices Directive 98/79/EC





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