LightPilot 2.0™ by Wilson Analytical

Software

User's Manual
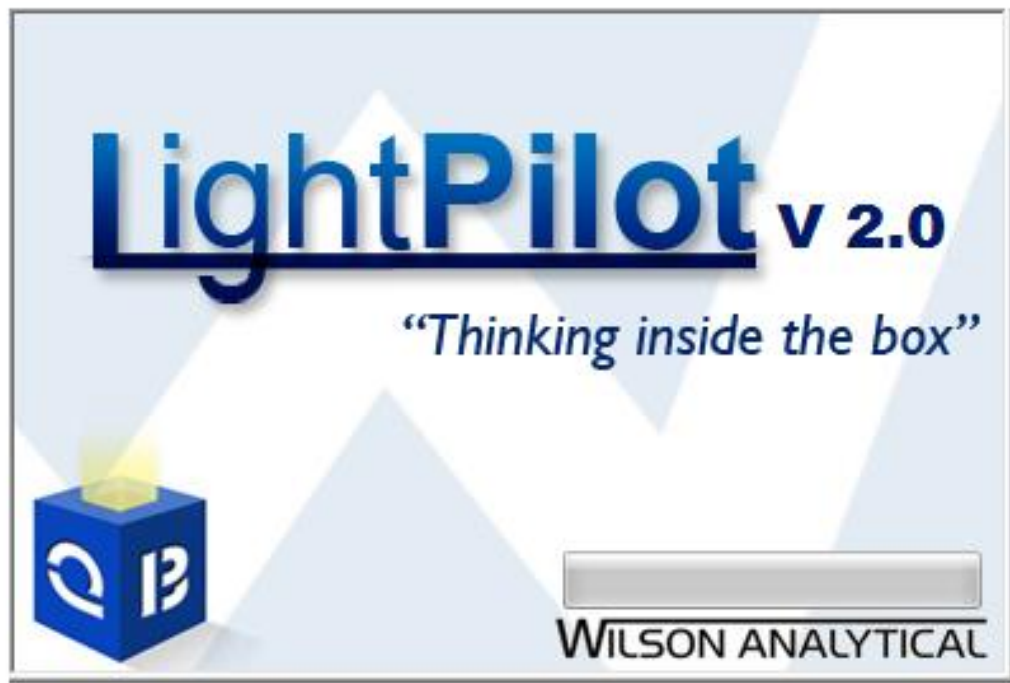
May 2022

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Installing and Running LightPilot Software

If you are using a netbook or other computer supplied by Wilson Analytical, LightPilot will have already been installed and configured. Once the netbook or computer is running, double-click the blue icon to launch LightPilot.

Figure 1. The Wilson Logo used as the LightPilot icon.

If the LightPilot software has not already been installed on your computer, the program can easily be installed, either by inserting the LightPilot USB flash drive that was supplied, or by downloading the latest version of the software from the Wilson Analytical website, at www.wilsonanalytical.com.

To install LightPilot, follow the steps below:

Double click on the LightPilot setup exe and follow the prompts to install the software. After installing the software go to Program Files on your computer. In order to do this, open up File Explorer, then click on 'This PC'. You will then be brought to 'Devices and Drives' where you can click on your Windows' Drive, such as Windows (C:). Once you are in your Windows' Drive double click on the folder titled Program Files. In the Program Files folder, find and open the folder that says Wilson Analytical, then double click on the folder 'LightPilot QB'.

Once you are in the LightPilot QB folder ensure 'OmniDriverSPAM-2.46-win64-installer' is installed by double clicking on it and following the prompts. When you must choose which version to install click on 'Redistributable version' then click next. When you are asked if you want to install VCREDIST silently click on 'Yes' then click next. Next ensure 'jdk-8u121-windows-x64' is installed by double clicking on it and following the prompts. When selecting optional features to install make sure 'Development Tools' is highlighted then click next. Finally ensure 'vc_redist.x64 and 'vc_redist.x86' are installed. If you are prompted that 'Setup Failed' because another version is already installed, do not worry. This most likely means the program has already been installed properly so all you need to do is click on close.

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In all cases you will be alerted when the installation is complete, at which point rebooting the system is recommended. Once the system comes back up, you can launch LightPilot by double-clicking the blue icon on the computer desktop. If complications persist do not hesitate to contact us.

Once the software has been successfully installed, the LightPilot Software can be used to control any version of Wilson Analytical instrumentation. If you are not using the WhereBox (GPS receiver) an error message may appear prompting you to connect the GPS device. If you wish to run the software without the Wherebox, simply open the 'Tools' menu on the toolbar and select 'Change Setup'. This will open a dialog box asking you to enter the current password. Once you have entered the password, the setup window will open. Ensure the 'Require GPS' box at the bottom of the setup pop up is deselected and you will be able to run LightPilot without the WhereBox.
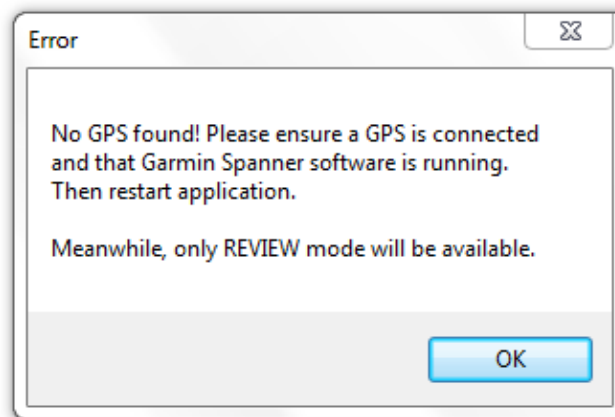

Figure 2. GPS error message that may pop up if the WhereBox is not connected.

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Overview of LightPilot Software

The LightPilot software was designed to make using Wilson Analytical instrumentation a straightforward and enjoyable process, while still ensuring that the data obtained is both accurate and reliable. Scientific jargon and complexity have been kept behind the scenes as much as possible, and the user is guided through the measurement process in a clear and logical fashion. The first thing to know is that the LightPilot software has three modes of operation, each dedicated to a particular task. Each of the modes is discussed in detail in the remainder of this section. The three modes are:

Control mode, used to create or modify a calibration curve for the chemical or chemicals that will be monitored;
Acquisition mode, used to take measurements on the samples of interest; and
Review mode, used to display, review and print data files.

The user can switch between these modes using the 'mode' menu in the toolbar. Note that because the validity and the accuracy of its data hinges on the quality of the calibration curves created by the user, access to the *Control* mode is password protected.
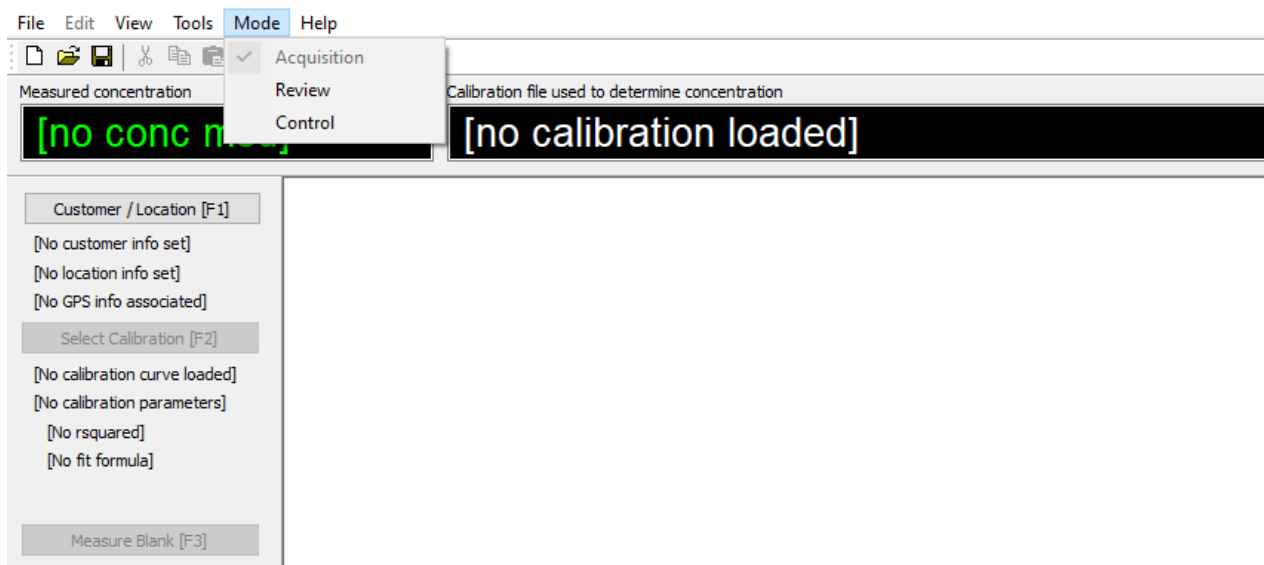

Figure 3. The 'Mode' menu located in the toolbar allows the user to switch between the Acquisition, Review and Control Modes.

In each of the three LightPilot modes, the software has been designed to guide the user through the completion of the appropriate activity (calibration, acquire data, review data) in a logical sequence. For example, Acquisition mode will not allow a chemical sample to be measured until a calibration curve for that chemical has been selected. The software guides the user by having steps grayed out until they are ready to be completed.

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Note that the LightPilot software has been designed so that it can be used without a mouse. Each of the selection buttons in all the modes can be activated by pressing the appropriate function key on the keyboard, for example 'Acquire Baseline [F1]'

LightPilot Setup

Though a default password to access the *Control* mode has been supplied with the instrument, you are not required to keep this password. It can be changed it by simply opening the 'Tools' menu on the toolbar and selecting 'Change Setup'. This will open a dialog box asking you to enter in the current password. Once you have entered the password, the setup window will open (see Figure 4).
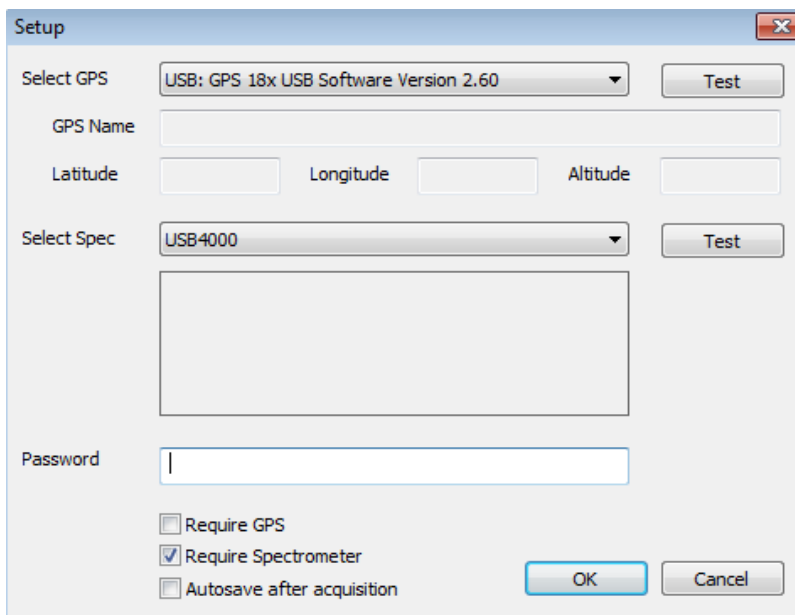

Figure 4. The setup window allows the user to change the password, test the GPS settings, test the Spectrometer settings, change the require GPS option and more.

From the setup window, you can change your password simply by deleting the existing password from the password text box and entering in the new password. The setup window also allows you to test your connections with both the WhereBox (GPS receiver) and the spectrometer, to ensure that both are connected and working properly. To do a check, click 'Test' beside the appropriate device in the dialogue box (see Figure 4).

Finally, the setup window is where you can choose to autosave after each acquisition run and/or set the GPS and the spectrometer requirements. In Figure 4, the 'Require GPS' box at the bottom of the setup window is deselected, allowing the user to run samples without the GPS. To save the changes, click 'OK' at the bottom right hand side of the window, once you have saved the settings, they will remain the same even once you exit LightPilot.

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LightPilot Data Spectrum and Cursor

The default view on the software is to display the baseline, the raw spectrum, and the data, however you do not have keep this setting. To change the data spectrum that is displayed simply open the 'View' menu on the toolbar, hover over 'Data Spectrum' and deselect the datapoints you do not want displayed (see Figure 5). It is important to remember that the default setting will be displayed each time the program is closed and reopened, meaning you will have to change the view each time you open the software.
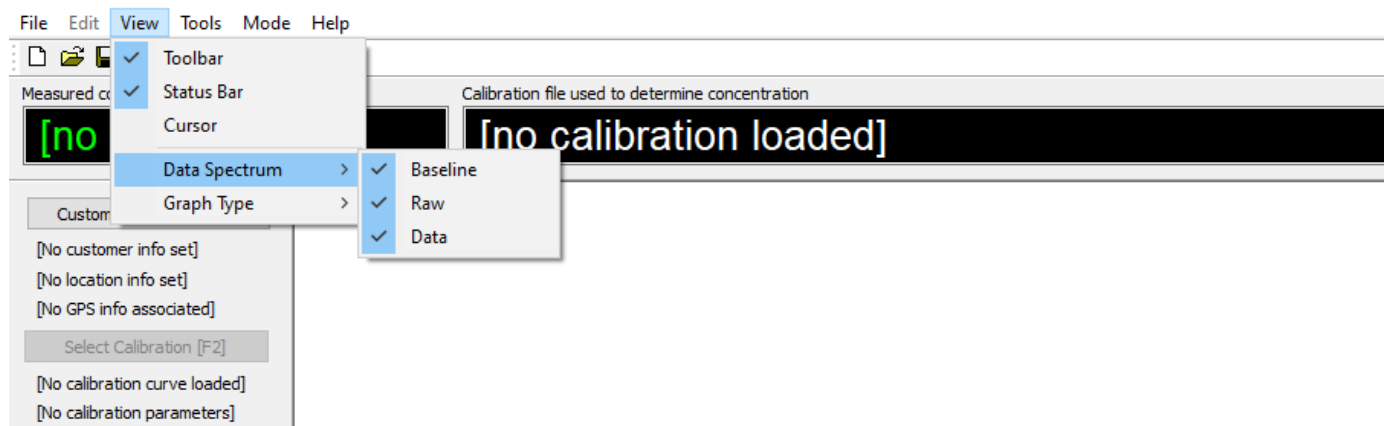

Figure 5. The View menu located in the toolbar allows the user to edit the viewing settings on LightPilot, including the Data Spectrum.

In each of the three modes, it is possible to view the number of spectrometer counts (intensity) at varying wavelengths by opening the 'View' menu in the toolbar and selecting 'Cursor'. A dialogue box will open, called 'Cursor'. From here, you can select the wavelength at which to display the counts. By default, the cursor is set to the lowest wavelength at the far left of the graph when it is opened (see Figure 6).
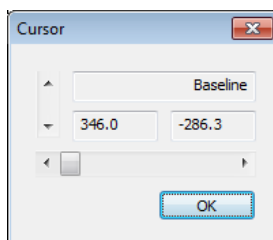
Figure 6. The cursor dialogue box.

There are two ways to adjust the wavelength: to change by large intervals, move the scroll bar side to side; to move by smaller intervals (0.2 nm of a wavelength), click on the left and right arrows to either side of the scroll bar. Note that the cursor line starts at the left of the spectrum (short wavelength end). Using this cursor, you can determine the height of any peaks that appear on the graph. The wavelength being read at any given time is represented on the graph as a vertical black line.

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'Control' Mode Introduction

LightPilot works by taking the measurement of the fluorescence intensity from an unknown sample and comparing it to the expected intensity from the same chemical, measured at a series of standard concentrations. The chart of 'fluorescence intensity *versus* concentration' for any given chemical is called its calibration curve.

Without a calibration curve, the data measured by the instrument is useless, and so it is the purpose of the Control mode to guide the user through the creation and/or modification of calibration curves. The necessary steps are listed on the left-hand side of the Control mode screen. In this section, we will discuss each of these steps in turn.

Choose 'Control' from the 'Mode' menu located on the toolbar, and a password dialog box will open. Type in the password to access the Control Mode (see Figure 7).
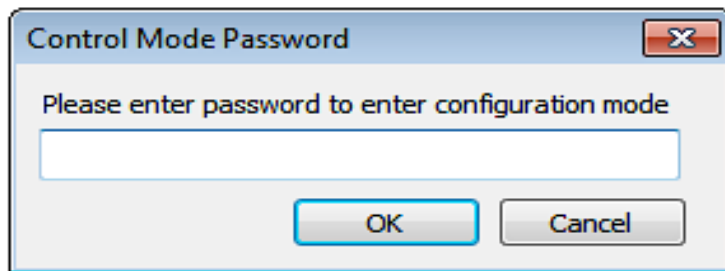

Figure 7. The password dialog box pops up to access the Control Mode.

Once you have accessed the Control Mode follow the appropriate activities from top to bottom starting with the 'Substance Name'. Let us assume that we want to create a calibration curve for a corrosion inhibitor chemical called Corrosion Inhibitor (CI)-A. Type in 'CI-A' (no need to type the quotes) in the 'Substance Name' text box. When the calibration file is saved, at the end of the process, it will be given the name 'CI-A.cal' (again, without the quotes). More information can be added to the name if multiple calibration curves will be needed for the same chemical. For example, 'CI-A' could be 'CI-A 1ppm - 100ppm OPS 07-Aug-2020. If you are re-doing a calibration curve for a chemical, we recommend copying and pasting the previous calibration name into the new curve and then changing the date. This ensures proper consistency and organization.

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Each instrument contains a spectrometer with a unique serial number. The LightPilot software keeps track of the spectrometers used to create calibration and data files, and you may occasionally see a warning alert that data you are using was created with a different unit. This is perfectly normal and there is no reason for the user not to proceed with measurements using calibration curves created on other Wilson instruments. The light output from each instrument has been carefully calibrated to be equivalent, so that clients with more than one unit can work efficiently by sharing data and calibration curves as desired. However, this is assuming the instruments are using the same wavelength for the light source and the same type of spectrometer. Various light sources are available for installation during the construction of the unit.

There is a drop-down menu immediately below the 'Substance Name' text box (see Figure 8), which allows the user to select the instrument that is currently detected by the LightPilot software, or devices that have been detected in the past. In most cases, only one serial number will appear. In the event that two or more serial numbers are displayed, the user should select the unit that will be used for their testing.
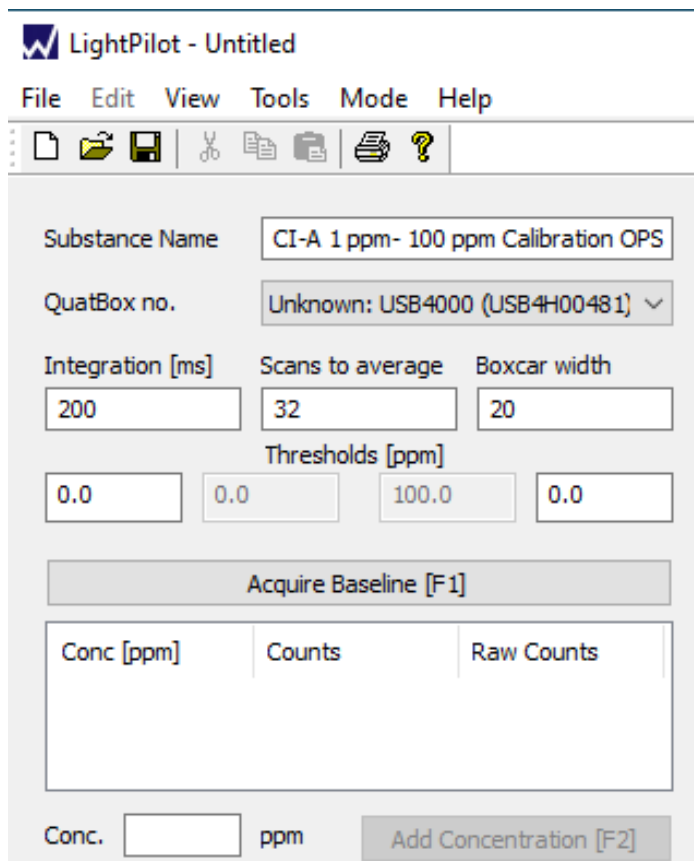

Figure 8. Image of the 'Substance Name' text box and the instrument drop down menu located at the top left-hand corner in the Control Mode.

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Three Spectroscopy Parameters for Calibration in the Control Mode

Wilson Analytical's fluorescence instrumentation exposes a sample to a constant source of light and records the intensity versus wavelength of the light that is re-emitted by the sample. This 'emitted intensity' is recorded as the number of counts at each wavelength between 380nm and 1100nm and is displayed by the LightPilot software as a spectrum on the computer screen. Different chemicals emit light at different characteristic wavelengths, and the normal procedure is to pick a wavelength of maximum intensity (i.e., a peak) as the wavelength for the calibration of a particular chemical.

The next step is to choose values for the three-spectroscopy parameters required to take a measurement. These parameters are:

Parameter 1: Integration Time
Parameter 2: Scans to Average
Parameter 3: Boxcar Width

Now these parameters may not mean much to you, but they are important and will affect the intensity of the fluorescence signal and the quality of the data that you obtain. The parameters have the following effect:

Parameter 1: Integration Time

Think of integration time like the shutter speed on a digital camera. The longer the integration time, the longer the 'shutter' is open, and the more light (signal) reaches the detector. Just like in photography, you want your picture (your data) to have the right exposure. Too much exposure, and you will lose contrast and linearity. Too little, and you will lose sensitivity and be unable to measure low concentrations accurately. And just like in photography, you sometimes have to play around with the settings a little bit to get the exposure just right!

If we are planning to use CI-A at 500ppm, we might want a calibration curve that spans 50ppm (10x lower) to 2500ppm (5x higher). Though this would be ideal we must remember that one of the four secrets to accurate fluorescence measurements is staying in the linear range, and the maximum concentration that we can measure directly in the linear range may be as low as 50ppm. As the dilution factor in this case must be at least x50, we may as well pick x100, as this gives us a margin of safety, plus easier arithmetic. With a dilution factor of x100, the calibration for CI-A will require samples with concentrations ranging from 0.5ppm to 25ppm, for example 1ppm, 5ppm, 10ppm and 25ppm. If necessary, more concentrated samples at 50ppm and 100ppm could be prepared, to see at what point, and to what extent, the calibration curve becomes non-linear.

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The basic principle in selecting the correct integration time is that a doubling of the time will double the amount of light reaching the detector, and hence double the number of counts recorded in the spectrum. The detector will show a maximum of 64,000 counts at any wavelength, so the integration time in this case must be such that at 25ppm, the maximum number of counts is less than 64,000. At the low concentration end of the measurement, we would like to be able to see 0.5ppm reliably, and the minimum number of counts necessary to do this is around 10. In the middle of the range, where we hope to see most of our samples fall, we would like the 5ppm sample to have hundreds or maybe a couple of thousand counts at the chosen wavelength.

In most cases, there will be a range of integration times that are suitable to allow the user to 'see' the concentration range of interest quite comfortably. A good starting point for the integration time is 250ms (250 milliseconds). If there are insufficient counts at the low end of the concentration range, increase the integration time. Conversely, if there are tens of thousands of counts at the high end of the chosen range, decrease the integration time accordingly.

Note that if for any reason a single integration time value that covers the whole concentration range appropriately cannot be found, the user has two choices. The first is to select a value of the integration time that makes the low concentrations 'work', and simply further dilute samples that are off scale. Or secondly, construct two calibration curves, one for higher concentrations, and one for lower.

It is important to note that once an integration time has been chosen, and a measurement taken, all further measurements for that calibration curve will use the same integration time. This feature is hard-wired into the *Control* mode so that the user cannot inadvertently take measurements with different parameters and try to construct a single curve. It only makes sense to take sample measurements with the instrumentation set up in the same configuration that was used to construct the calibration curve. This is one of the things that the LightPilot software looks after "behind the scenes".

Parameter 2: Scans to Average

This parameter is fairly self-explanatory. It is simply the number of fluorescence measurements that the instrument will obtain and average together before displaying on the screen. A big advantage of the fluorescence technique is the speed at which measurements can be taken; a few seconds at most. It is a good idea to take multiple data sets and average them together, as it takes so little time to do so, and the statistical accuracy of the data is thereby improved. A good default value for this parameter is 32, and the user can decide through experimentation if there is a particular advantage in a greater or lesser value.

As with the integration time, once the 'scans to average' parameter has been set, and a measurement taken, all further measurements for that calibration curve will use the same value.

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Parameter 3: Boxcar Width

The boxcar width is a technical parameter that has to do with smoothing of the data. A default value of 20 is acceptable for most applications, and in most cases, the user will see little difference from adjusting its value. Boxcar width has been retained as a parameter for the more expert user, and for those requiring the most extreme precision in their work.

As with the 'integration time' and 'scans to average' settings, once the 'boxcar width' parameter has been set, and a measurement taken, all further measurements for that calibration curve will use the same value.

Thresholds

Beneath the boxes for 'integration time', 'scans to average' and 'boxcar width' are four boxes labeled "Thresholds". Of these boxes, the middle two are 'greyed out', while the leftmost and rightmost ones can have data entered into them.

The two 'greyed out' boxes represent the range of the calibration curve, that is the lowest and highest ppm standard measured in making the curve, respectively. These boxes will fill in automatically as you make your calibration curve.

The leftmost white box allows you to enter the lowest ppm sample that falls outside the range of the curve you want to be reported. The rightmost white box, similarly, lets you enter an upper threshold, or the value that falls above the range of the curve that you want to be reported. Those values that fall into the areas between the threshold values and the range of the curve will be reported in orange, while those that fall outside the threshold values will be reported in red. Values that fall within the range of the calibration will be green.

Note that it is not mandatory to set threshold values; if you leave them blank, they will be assumed to be the range of the curve, and thus any values falling outside the range of the curve will be reported in red.

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Creating the Calibration Curve

The next step is to take a measurement with a 'blank' sample. This gives the instrument a baseline response (from the cuvette, solvent, etc.,) that is subtracted from all subsequent sample measurements. The blank sample should be the same as the calibration standards in all respects other than the chemical you are measuring. For example, if you decide to make the calibration standards for CI-A in a 95% water, 5% isopropyl alcohol (IPA) solution that contains 100ppm of sodium chloride, then this solvent matrix should be used for the blank measurement.

To acquire a spectrum from the blank sample, check that the instrument is ready as indicated by the green status LED's, open the hat on the Cuvette holder or take off the lid on the Solid Sampler and insert the blank sample. Note the red LED turns on when the sample holder's hat or lid is off. When you close the hat or put on the lid, the *Interlock* LED should turn green. If it is red, the Cuvette Holder hat is not closed, or the Solid Sampler lid is not on properly. When the blank is in the sampler holder and the LED's are green, select 'Acquire Baseline'. When the measurement is complete (*Power or data* LED on the front changes from blue to green), remove the blank sample. The baseline spectrum will be displayed, as well as the zero concentration counts at the calibration wavelength (see Figure 9).
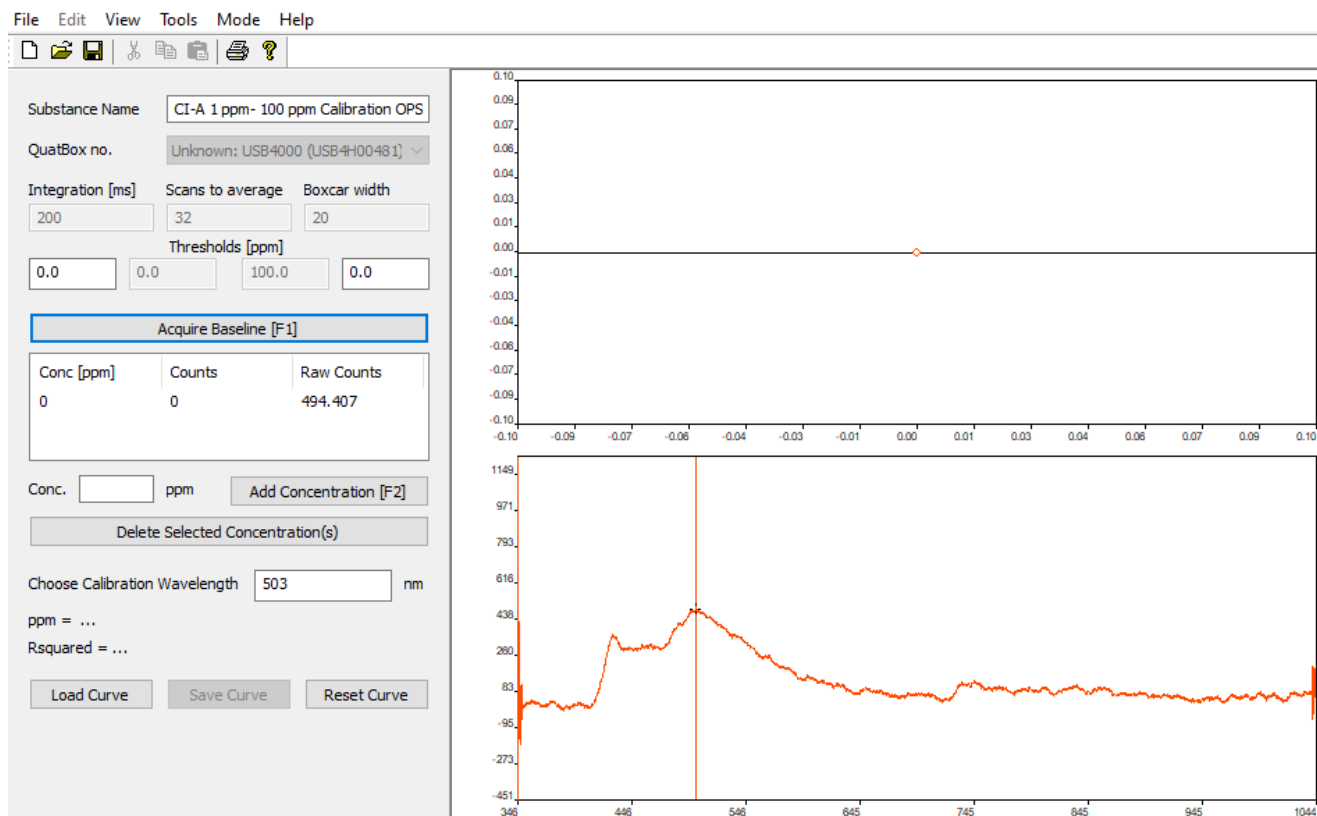

Figure 9. Image of the blank sample, also referred to as the baseline. The written values are shown in the box located under 'Acquire Baseline', while the visual spectrum display box is on the bottom right of the screen.

Now that the three-spectroscopy parameters have been configured, and a baseline measurement has been taken, the actual calibration curve for CI-A can be created.
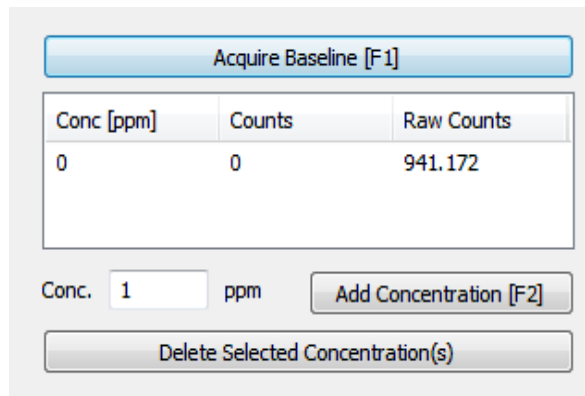
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One way of developing a calibration curve is to create a 'dilution series' of the chemical of interest in distilled or deionized water. For example, a series of samples with 1ppm, 5ppm, 10ppm, 25ppm and 50ppm concentrations of chemical would form a very good calibration sequence. In practice however, chemicals are used in 'real world' water systems that may or may not contain salts, surfactants or other chemicals that could potentially interfere with the fluorescence measurement. Therefore, if possible, it may be a good idea to use the same environment, or sample matrix as found in the field to prepare the calibration samples.

For example, if a corrosion inhibitor is being used at 500ppm in a system where the produced water has a salinity of 5% (50,000ppm), a useful calibration series could be formed by making an accurate 500ppm solution of inhibitor in the produced brine and diluting it by factors of x50, x100, x500, x1000 and x5000, to give solutions of 10ppm, 5ppm, 1ppm, 0.5ppm and 0.1ppm. Alternatively, solutions of 10ppm, 5ppm, 1ppm, 0.5ppm and 0.1ppm could all be prepared in a 100ppm brine (corresponding to the expected strength of the matrix; 500ppm in a 50,000ppm brine, diluted by x500).

Note that it is not necessary to prepare five or six samples to create a calibration curve. The *Control* mode will assume that the fluorescence intensity at zero concentration (0ppm) will be zero, i.e., all calibration curves go through the origin. Therefore, even a single sample, say 10ppm, will result in a straight-line calibration that could be used for subsequent measurement of unknowns. A two-point calibration is however ill-advised, because it will not confirm that you are operating in the linear range and can easily result in large errors between the measured and actual chemical concentrations.
It is advisable to use at least three chemical dilutions to create a calibration, and the concentrations chosen should cover at least an order of magnitude in concentration range. In other words, 1ppm, 10ppm and 50ppm will create a much better calibration curve than 1ppm, 2ppm and 5ppm.

Let us say that we have prepared five calibration samples for CI-A, at 1ppm, 25ppm, 50ppm, 75ppm, and 100ppm. Place a cuvette containing the 1ppm solution into the sample holder and close the sample cover. Type '1' in the white text box that appears between the symbols 'Conc.' and 'ppm', and then select the 'Add Concentration' button (see Figure 10). The fluorescence spectrum from the 1ppm sample will be displayed on the lower right-hand side of the *Control* screen.

Figure 10. 1ppm Concentration sample about to be tested, with 1 typed into the Concentration text Box.

LightPilot will now construct a data table that shows the concentrations run so far, the units, counts and raw counts. This data is the beginning to the calibration curve and displayed graphically on the upper right-hand side of the screen.

Proceed with the creation of the rest of the calibration curve by adding the 25ppm, 50ppm, 75ppm, and 100ppm concentrations in the same way as you did for the 1ppm sample. For each concentration, place the cuvette with the new concentration in the cuvette holder, close the cover, type the new concentration in the text box, and select 'Add Concentration'.

If at any time you make a mistake, or simply wish to remove one or more of the data sets, simply highlight that measurement or measurements by clicking on them and then select 'Delete Selected Concentration(s)'. You can delete a sample if something was typed incorrectly or if you would like to redo a data point.

When you have measured all six calibration samples, you should see six spectra displayed at the bottom right-hand side of the screen in the spectrum display box, each with the same shape (peaks in the same place) but with different intensities (see Figure 11).
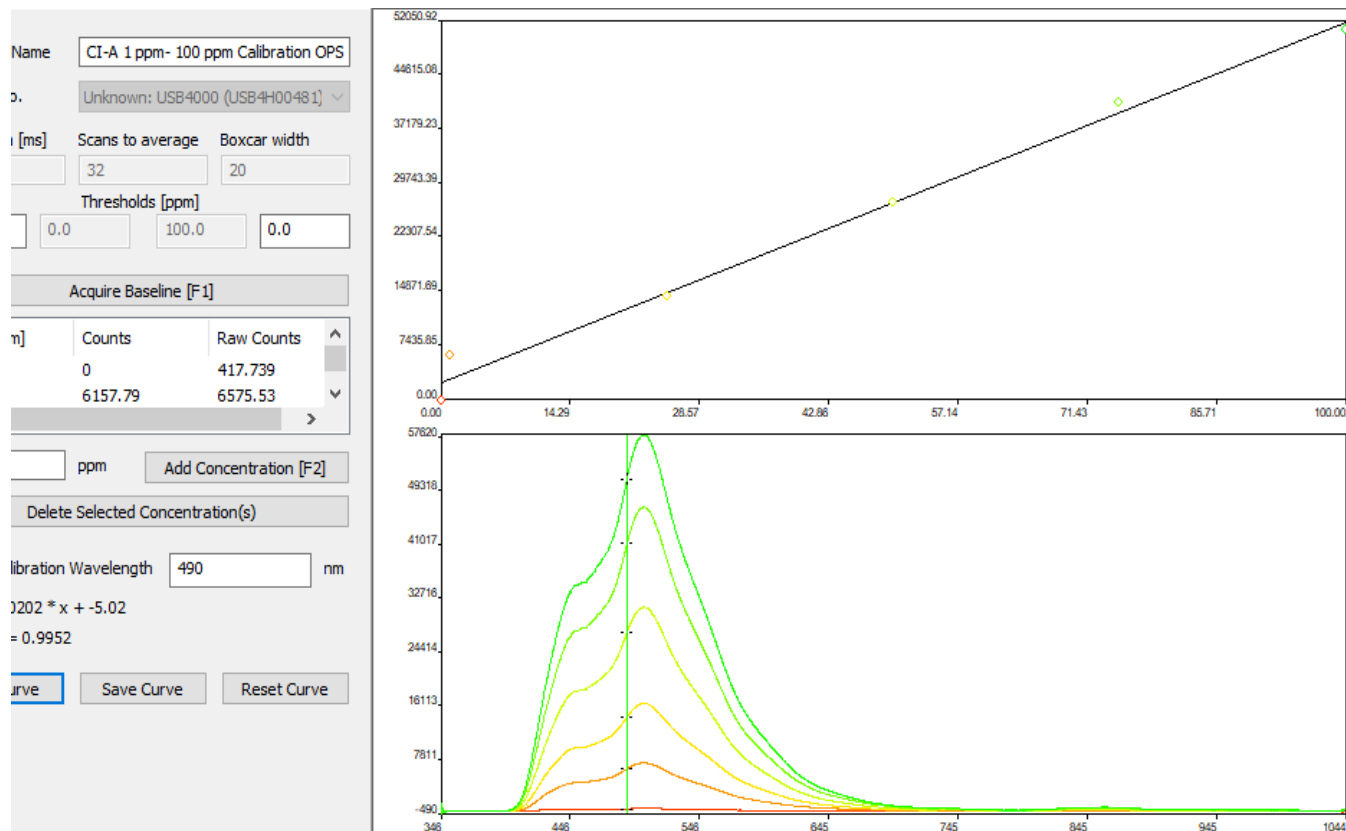
Figure 11. Image of the spectrum display box in the Control Mode with six different concentrations of a sample measured.

In the spectrum display box, there is a single vertical line (See Figure 11). To complete the creation of the calibration curve, you must select a calibration wavelength by moving this vertical line to the proper wavelength. In order to move the vertical line, type in a wavelength into the text box to the right of 'Choose Calibration Wavelength'. So, for example, if you see that your fluorescence spectra have peaks around 503nm, then type "503" into the 'Choose Calibration Wavelength' box (see Figure 12).
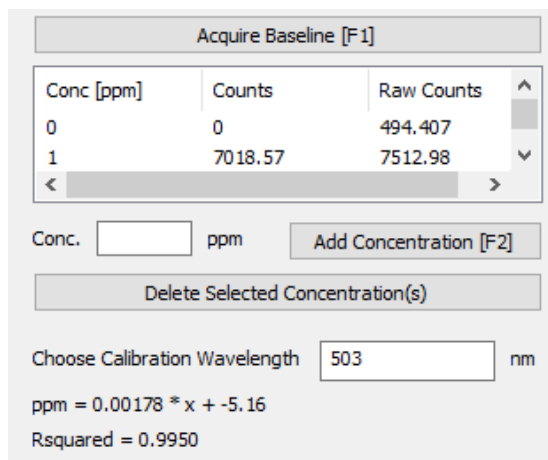

Figure 12. 503 nm Calibration Wavelength has been chosen and typed into the appropriate text box.

The vertical line will move to 503nm in the spectrum display box, and you should now see a linear calibration curve in the graph box on the upper right-hand side of the screen that is close to linear (see Figure 13). If the vertical line in the spectrum display box is not at the peak maximum, simply adjust (move) the line by typing a different number into the Calibration Wavelength box. For example, if the line is too far to the right, try typing '500' in the Calibration Wavelength box and the line will move 3 nm to the left from the previous 503 nm Calibration Wavelength.
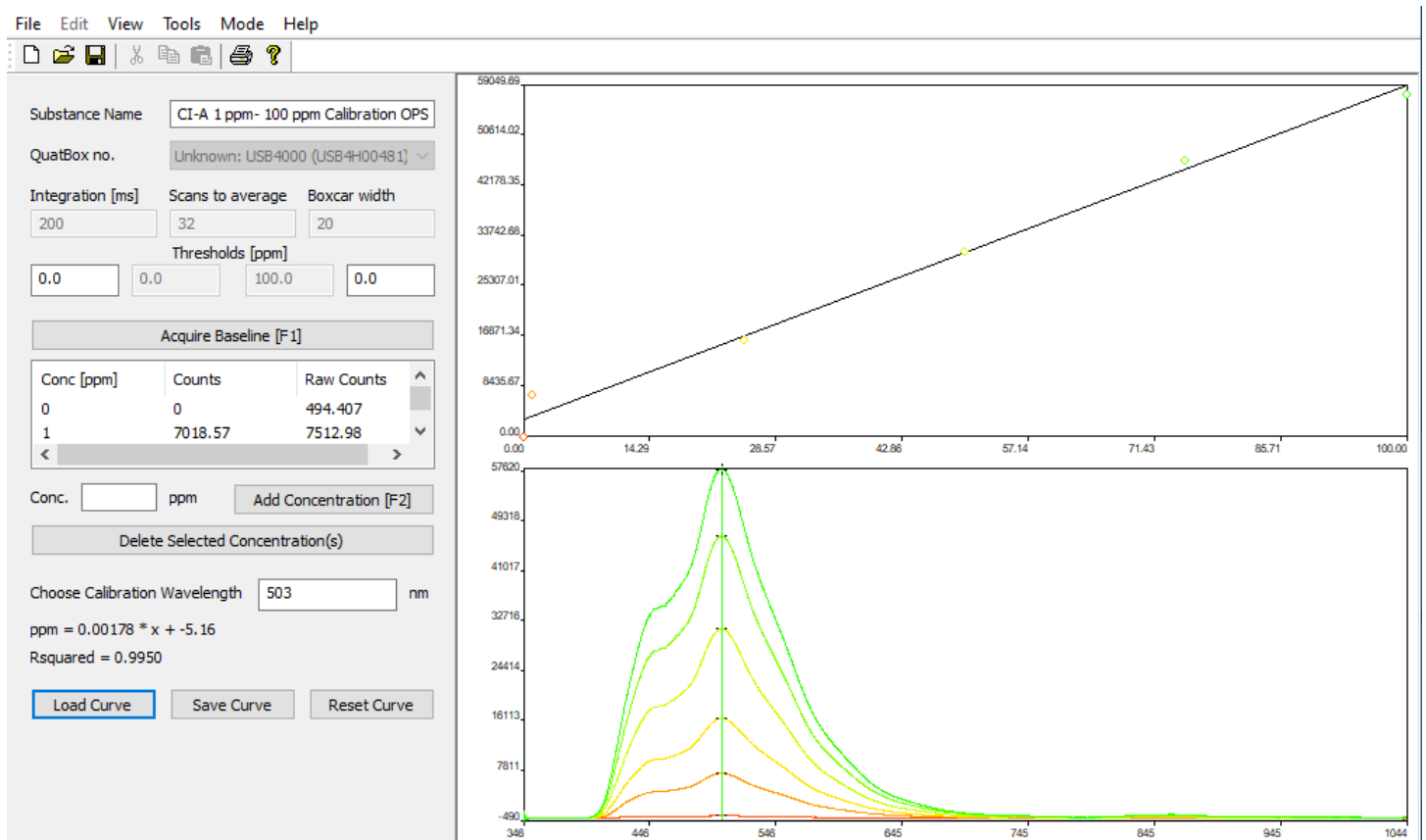

Figure 13. Linear calibration curve in the graph box on the upper right-hand side of the Control Mode, and the spectrum display box on the bottom right-hand side of the Control Mode with the vertical line at the peak maximum.

At this point, the user has complete control over the calibration and can add or subtract samples with different concentrations, or can choose a different wavelength for the calibration, until calibration curve at the top right-hand side of the screen is as linear as possible. The equation of the calibration curve and the R squared value are calculated and displayed immediately underneath the 'Choose Calibration Wavelength' box. This provides the user with more information regarding the linear trend. An R squared value of 0.98 or greater is ideal when making a calibration curve.

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The final step is to save the calibration curve by simply selecting 'Save Curve' at the bottom left-hand side of the screen (see Figure 13). Once you have done this, the 'Save As' dialogue box will pop up which allows you to choose where the calibration curve is saved, and the name of the calibration curve (see Figure 14). Once you click on 'save' the calibration curve has been successfully created and it can be used in the Acquisition mode.
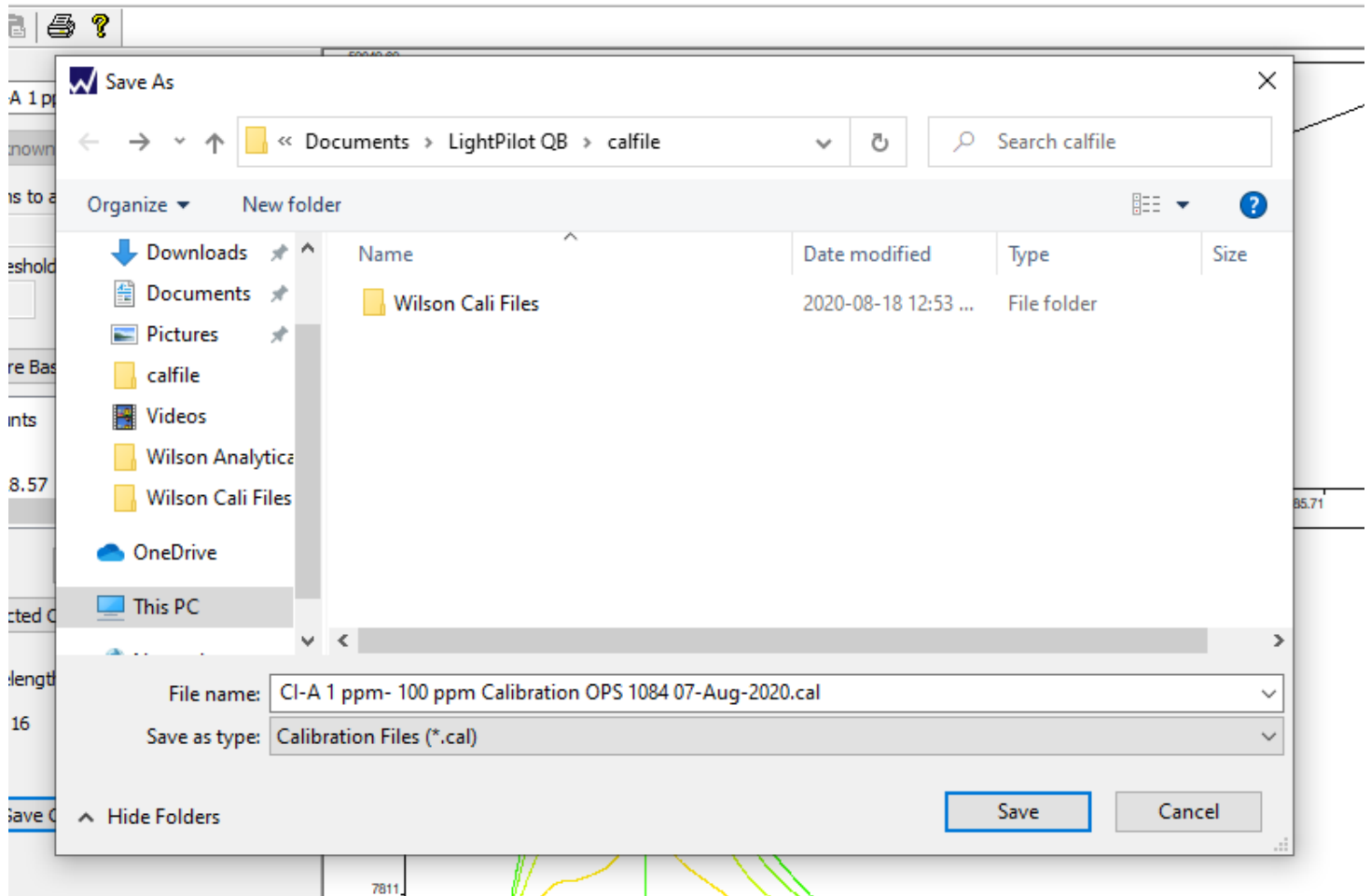

Figure 14. After the user clicks on 'Save Curve' the 'Save As' box pops up allowing the user to choose where the calibration curve will be saved and the name it will be saved under.

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'Acquisition' Mode Introduction

Once a calibration curve has been successfully created, Wilson's instrumentation can be used to determine chemical concentrations in unknown samples. From the toolbar, select the 'Mode' menu, and then Acquisition. No password is required, and the Acquisition screen will be displayed.

Acquisition mode breaks down the measurement of each sample into five sequential steps, and the user is guided through each of the five in turn. The LightPilot software makes it impossible to take the steps out of sequence, although it is possible to go back and change previous steps. Options that are not yet available for a particular sample are 'grayed out' (see Figure 15). For example, once step one has been completed, step two becomes available. Once that has been completed, step three is available, and so on.

The five steps are:
      Entering information about the sample (e.g., name and location)
      Selecting a calibration curve
      Measuring a blank sample
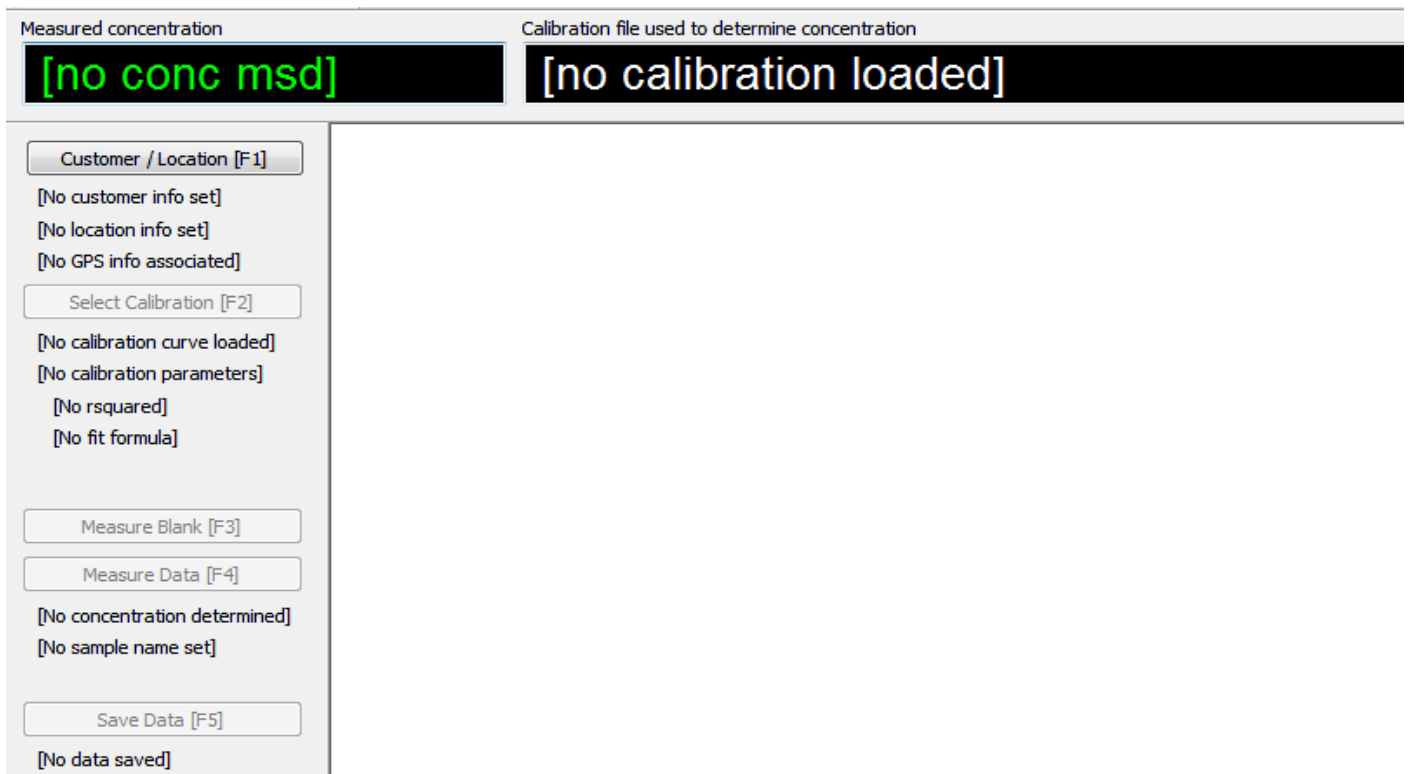   Measuring the unknown sample
      Saving the data file.

Figure 15. The Acquisition Mode with the five steps displayed on the left-hand side of the screen. Note the first step 'Customer/Location' is the only step available for use at this point.

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Entering Information

After clicking 'Customer/Location [F1]' in the Acquisition Mode, the Customer Information dialogue box appears allowing you to enter in information about the customer and sample. While entering in information you have the option to use the WhereBox unit, or not. If you choose to use the WhereBox, the GPS coordinates, the date and the time will be displayed at the top of the screen in the Customer Information dialogue box. If you choose not to use the WhereBox the GPS field will just contain default letters and question marks that can be safely ignored (seen in Figure 16).
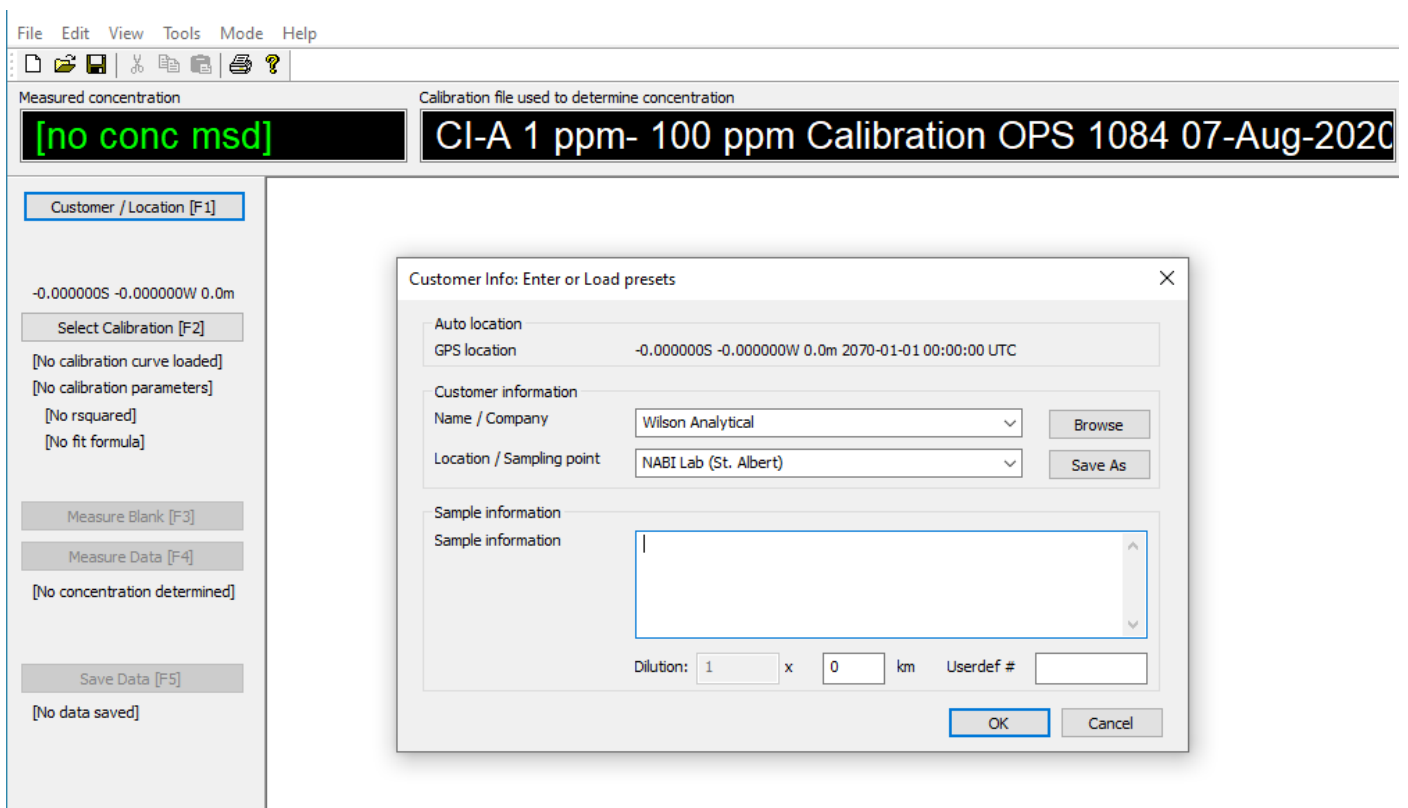

Figure 16. Customer Information dialogue box that allows the user to enter in Customer or Sample information. Note in GPS location the data is unknown because the Wherebox is not in use.

For the WhereBox to work, it must be placed in a location where a GPS satellite radio signal can be reached. It is best to place it out in the open; if indoors, it is best placed as near to the door as possible, especially if you are below ground. The bottom of the WhereBox is also magnetic, allowing it to be stuck to any metal surface, such as the roof of a truck or the wall of a metal shed. This helps to improve the range of places the WhereBox can be set up, allowing for the best GPS satellite radio signal possible.

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If you choose not to use the WhereBox, simply open the 'Tools' menu on the toolbar and select 'Change Setup'. This will open a dialog box asking you to enter in the current password. Once you have entered the password, the setup window will open. Ensure the 'Require GPS' box at the bottom of the setup pop up is deselected and you will be able to run LightPilot without the WhereBox.

Once you are in the Customer Information dialogue box (see Figure 16) you may notice that beside the 'Name/Company' and 'Location/Sampling point' boxes, there are arrows. You can click these arrows to open a dropdown menu of previously entered sample locations and company names. Selecting one of these will save you from having to enter this information multiple times if you are sampling at the same location. To save a new name or location you can click the 'Save As' button to the right of the 'Location/Sampling point' box. In future visits, if all the information remains the same, including the GPS coordinates, if you are using the WhereBox, you can simply click 'Browse' (located above the 'Save As' box) and select the location to fill in the company and location information.

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Selecting a Calibration Curve

As discussed earlier, the user must select a calibration curve that LightPilot can use to calculate the chemical concentration of the unknown sample. This is straightforward to do by selecting 'Select Calibration [F2]'. A list of files with the '.cal' extension will be displayed (see Figure 17). Simply select the calibration file that you wish to use, and LightPilot will use that particular calibration curve to determine the concentration of chemical in the unknown sample(s). Following the earlier example of the calibration curve that was created for CI-A, click on 'Select Calibration [F2]', and then select the file named 'CI-A 1ppm - 100ppm OPS 1084 07-Aug-20" as seen in Figure 17.
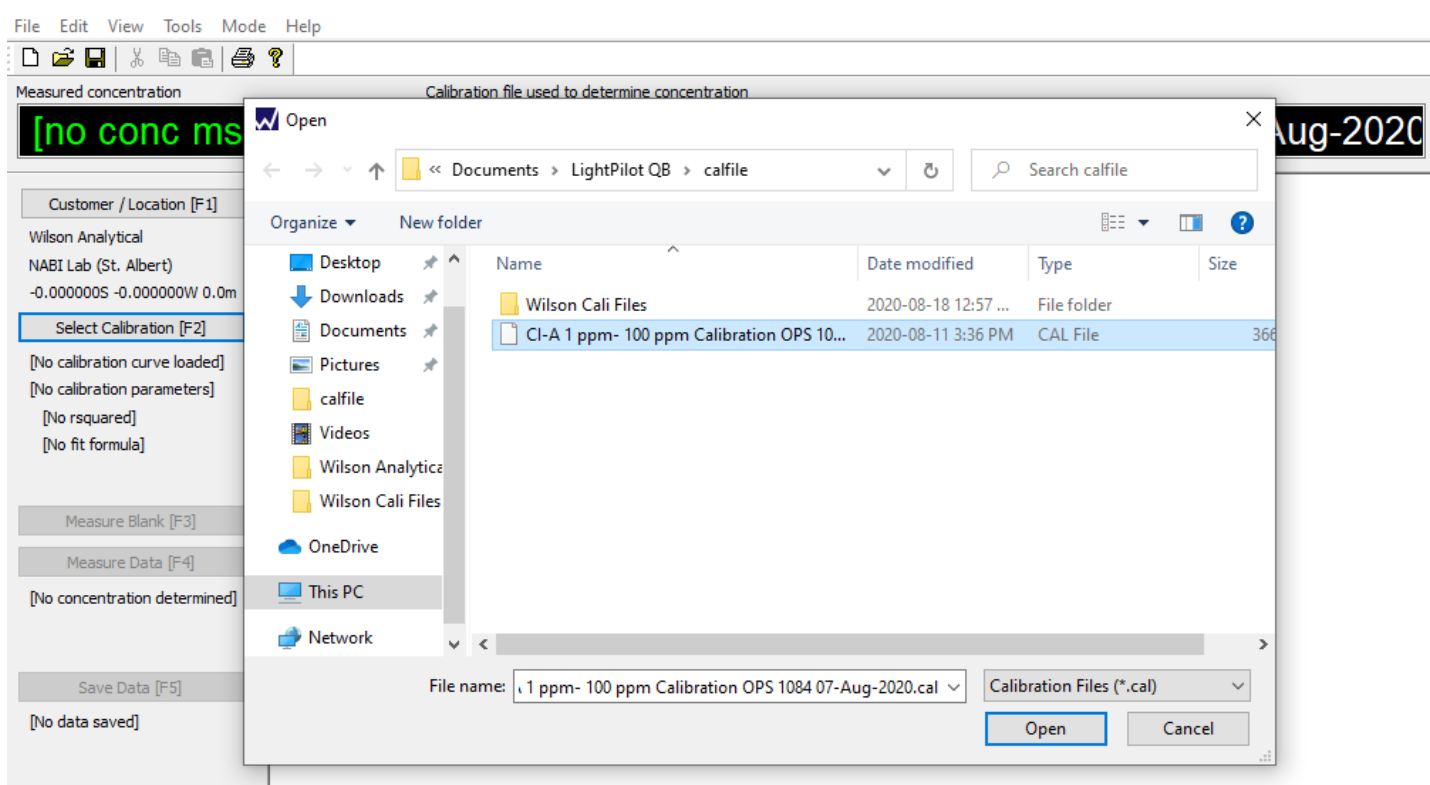

Figure 17. This window pops up after clicking 'Select Calibration [F2]' in the Acquisition Mode. Note there is only one Calibration Curve saved.

Once a calibration file has been selected, LightPilot reads the three-spectroscopy parameters that were used to create that calibration (Integration Time, Scans to Average and Boxcar Width) and uses them for the subsequent measurements of the unknown. The user cannot change the spectroscopy parameters from the Acquisition mode. This is because it makes no sense for measurements on unknowns to be interpreted using a calibration curve obtained with different spectroscopic conditions. As LightPilot forces measurements to be taken under the same conditions as the calibration, the user doesn't have to check back or worry about ensuring there is a match.

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Measuring a Blank Sample

Once the user has entered information about the sample and selected a calibration file, the next step is to take a measurement with a blank sample. This gives the instrument a baseline response (from the cuvette, solvent, etc.,) that is subtracted from the subsequent sample measurement. As discussed earlier, if it is possible the blank sample should use the same solvent matrix as the calibration standards.

To acquire a spectrum from the blank sample, check that the instrument is ready as indicated by the green status LED's. Insert the blank sample into the sample holder and close the cover. The *Interlock* LED should turn red when the sample holder's cover is open and green when the sample holder's cover is closed.

Once the LED's on the front panel are green you are ready to take measurements. Click 'Measure Blank Sample [F3]', and the software prompts you to ensure that the blank sample or the zero-concentration sample, is in place before proceeding (see Figure 18). After clicking on ok wait until the Power or data LED changes from blue back to green, and then remove the sample.
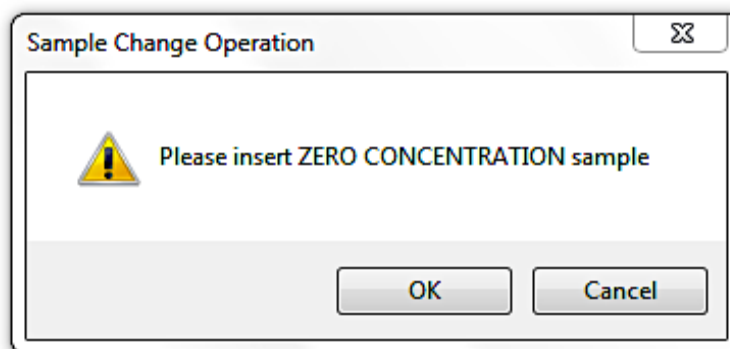

Figure 18. The window that pops up to ensure the blank sample has been placed in the instrument.

The fluorescence spectrum of the blank sample will be displayed on the computer screen. It is likely to be relatively uninteresting as it is a control sample. If the spectrum does contain peaks or features, there is a good chance the blank contains a fluorescent chemical or that the wrong sample has been selected (i.e., the sample isn't really a blank). If the blank contains fluorescent material, the user must be cautious that it does not interfere with the sample measurement.

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Before measuring the unknown sample, it is advisable to run the blank itself as a sample, to ensure the instrumentation is running properly. While it is not a necessary step in running a sample, running the blank as a sample can tell you early on if there is something wrong with your blank, or if the instrumentation is misreading the sample.

When the blank is run as a sample "against itself", the result may be somewhat noisy but should approximate a straight line as this is the same material running as both the blank and the sample and they should therefore cancel each other out. However, if there is a noticeable deviation from a straight line, you should either start over (i.e., redo the blank measurement) with the same blank, or prepare a new blank, to ensure the best results. Doing this serves as a sort of safeguard and ensures the instrument is working properly. It is not uncommon to have to rerun the blank a couple of times when the instrument is first started.

The default view on the software is to display the baseline, the raw spectrum, and the data, however you do not have keep this setting. To change the data spectrum that is displayed simply open the 'view' menu on the toolbar, hover over 'Data Spectrum' and deselect the data points you do not want displayed. It is important to remember that the default setting will be displayed each time the program is closed and reopened, meaning you will have to change the view each time you open the software.

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Measuring an Unknown Sample

Insert the unknown sample into the sample holder and close the cover. Proceed as before until the status LED's on the front panel are all green. Click 'Measure Data [F4]' and the 'Modify sample specifics' dialogue box appears to allow for any corrections to the sample information (see Figure 19). The next step is to type the dilution factor for the sample in the text box provided. For example, if the original sample was diluted by a factor of x100 prior to measurement, the user should type '100' (without the quotes) into the 'Dilution Factor' text box. If no dilution was used, ensure that the dilution factor is set to '1'. Once the information has been entered and the dilution has been recorded, click on 'Measure'. Wait until the Power or data LED changes from blue back to green, and then remove the sample from the instrument.

Figure 19. The 'Modify sample specifics' dialogue box that allows the customer to enter in additional information about the sample being measured and the dilution factor.

The fluorescence spectrum of the unknown sample will be displayed on the computer screen. The spectrum display will show a vertical line at the wavelength selected for the calibration calculation (see Figure 20). Typically, this will correspond to a peak in the fluorescence spectrum from the test sample. Note that the calibration wavelength cannot be changed at this point; it is displayed for reference only.

LightPilot will now calculate the actual concentration of the target chemical in the original sample, based on the dilution factor and the calibration curve that has been selected. For example, if the result of the measurement is 3.27ppm, and the dilution factor is x100, the result written at the top of the screen will be 320ppm. The result displayed at the top left-hand side of the screen will be green if it is within the range of the calibration curve as seen in Figure 20, and red if it is out of the calibration range.

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File Edit View Tools Mode Help

Measured concentration	Calibration file used to determine concentration
50.5 ppm	**CI-A 1 ppm- 100 ppm Calibration OPS 1084 07-Aug-2020**

Customer / Location [F1]

Wilson Analytical
NABI Lab (St. Albert)
-0.000000S -0.000000W 0.0m

Select Calibration [F2]

CI-A 1 ppm- 100 ppm
200ms / 32x / [20]
   Rsquared = 0.9950
   ppm = 0.00178 * x + -5.16

Measure Blank [F3]

Measure Data [F4]

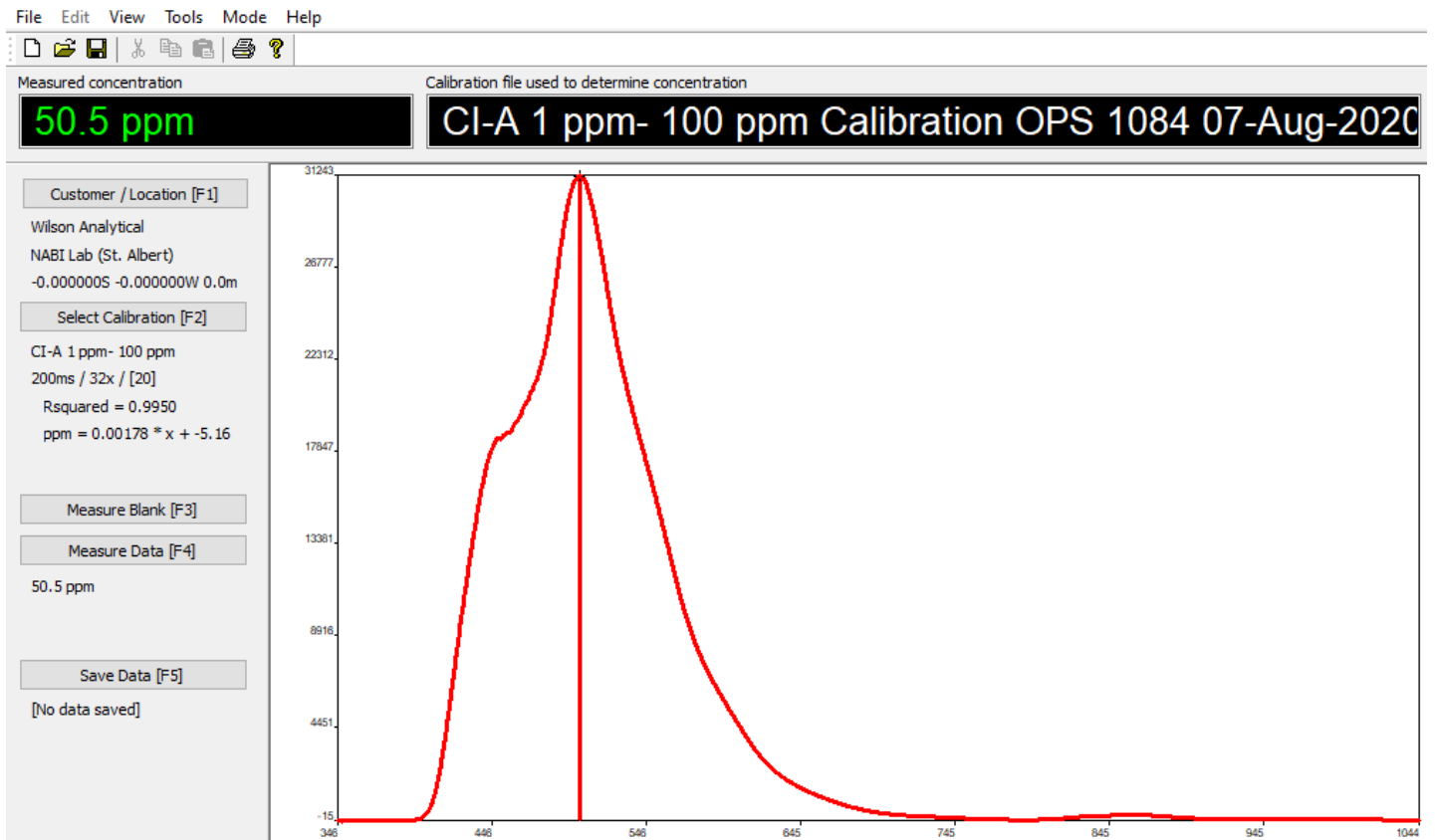50.5 ppm

Save Data [F5]

[No data saved]

Figure 20. The measured spectrum displayed on the bottom right of the screen, with the vertical line indicating the calibration wavelength. Note the calibration file is displayed above the spectrum and the measured concentration to the left has been adjusted for the dilution.

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Saving a Data File

The final step in acquiring a new measurement is to save the information in a data file. This is done very easily by clicking on the 'Save Data File [F5]' button at the bottom left-hand side of the screen. Saving the file is not automatic. However, if the user tries to leave the Acquisition mode without saving the data, or overwrites the current data without saving, he or she will be prompted by the software to save the new result. If you wish to make saving the file automatic refer to LightPilot Setup.

After clicking on the 'Save Data File [F5]' button at the bottom left-hand side of the screen, the 'Save As' dialogue box appears allowing the customer to choose where the file will be saved and what it will be named (see Figure 21).
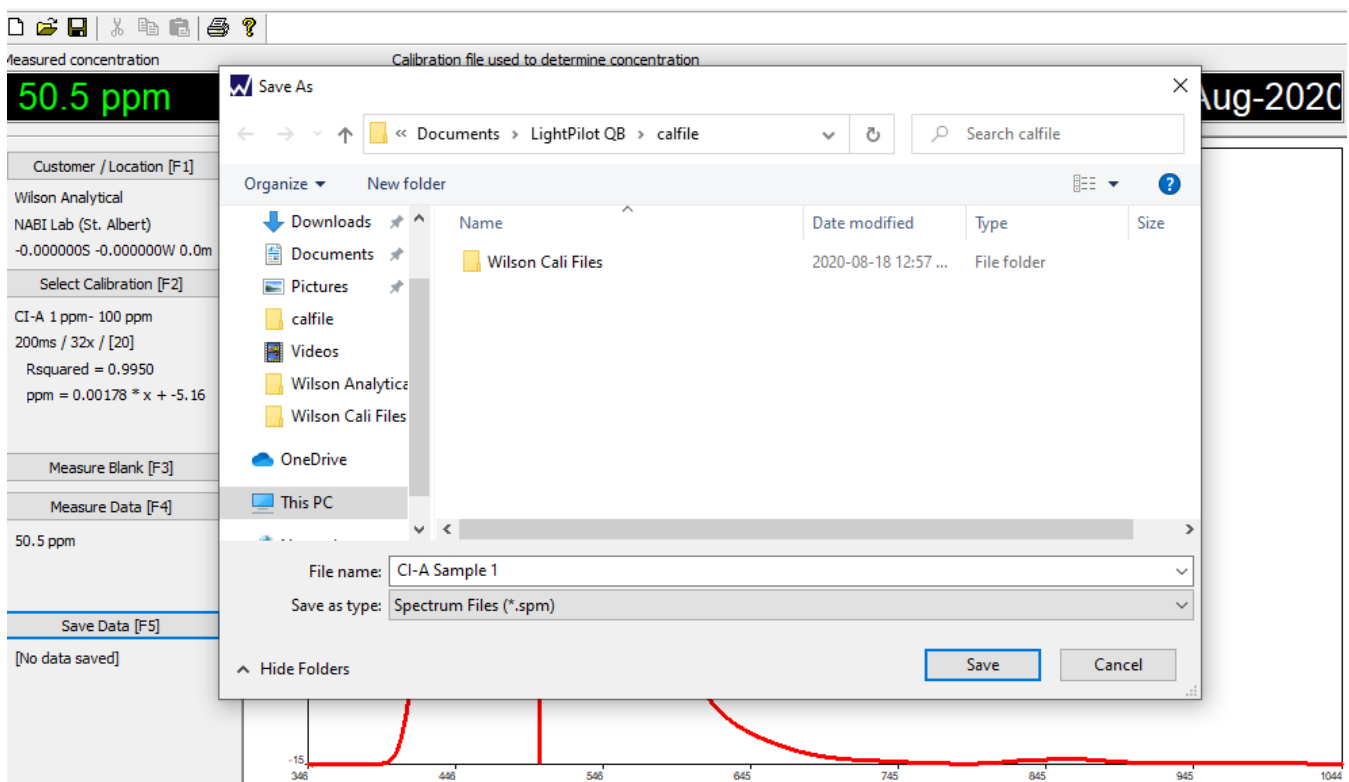

Figure 21. The Save As dialogue box which allows the customer to choose where the file will be saved and what it will be named.

Data files are saved with a '.spm' extension as seen in Figure 21. The files include all sample information entered by the user and the complete spectrum obtained from the unknown sample, plus information about the calibration file used, the GPS locators, a date and time stamp, and the unit number.

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'Review' Mode Display and Print Data Files

The third mode of the LightPilot software is Review, and is used to display, examine, and print data files obtained from the various test samples. From the toolbar select the 'Mode' menu and then click on 'Review'. No password is required, and the Review screen will be displayed.

Click on 'Add Data Sets', and a dialogue box appears with a list of all the available '.spm' files (see Figure 22). Double click the desired spectrum file or highlight the desired file and click open at the bottom of the dialogue box.
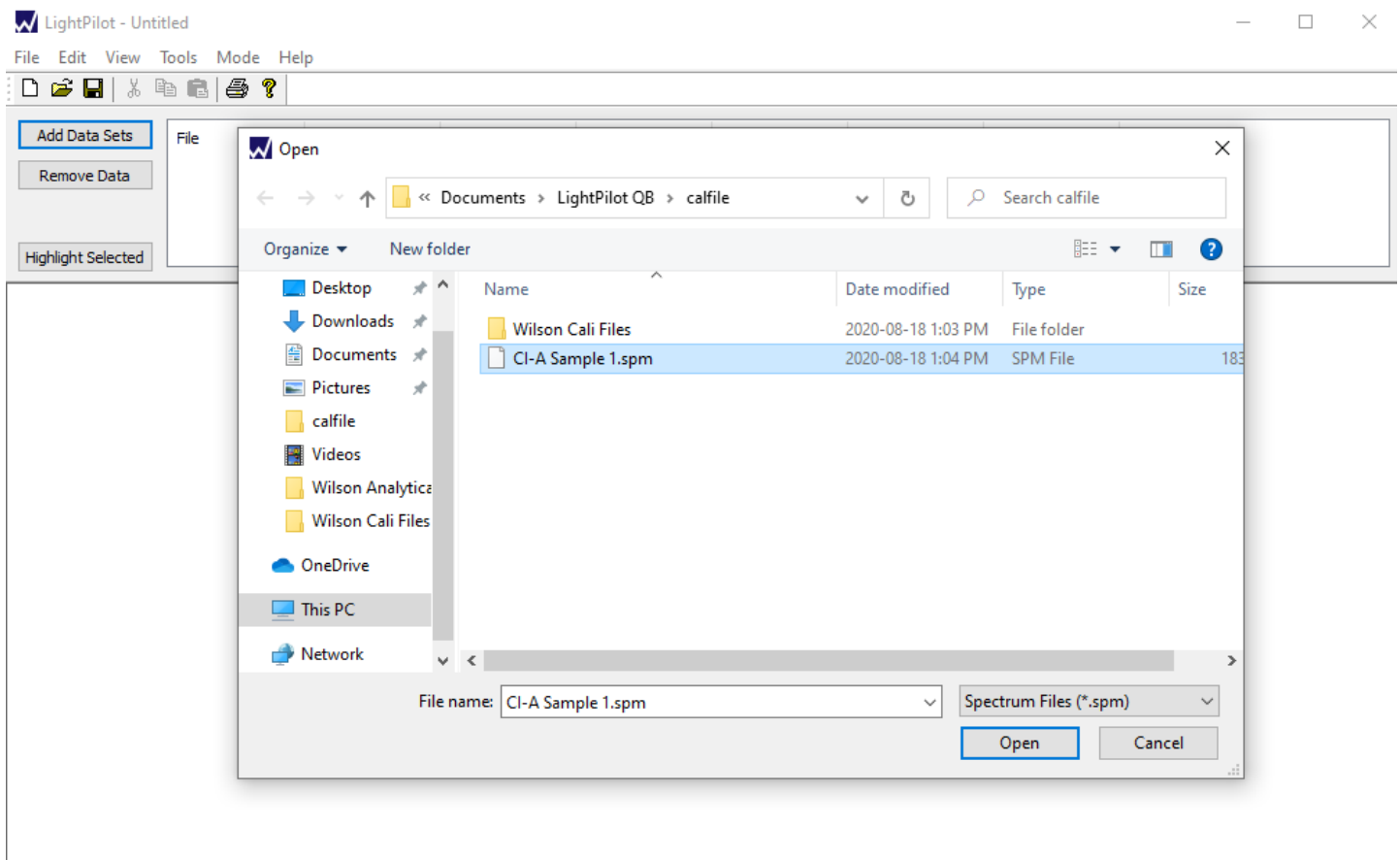

Figure 22. The dialogue box that appears after clicking on 'Add Data Sets' in the Review Mode. This window allows the user to select the data file they would like to review.

Once the file is selected the spectrum will be displayed in the middle of the screen while the file name is displayed above the spectrum as seen in Figure 23.
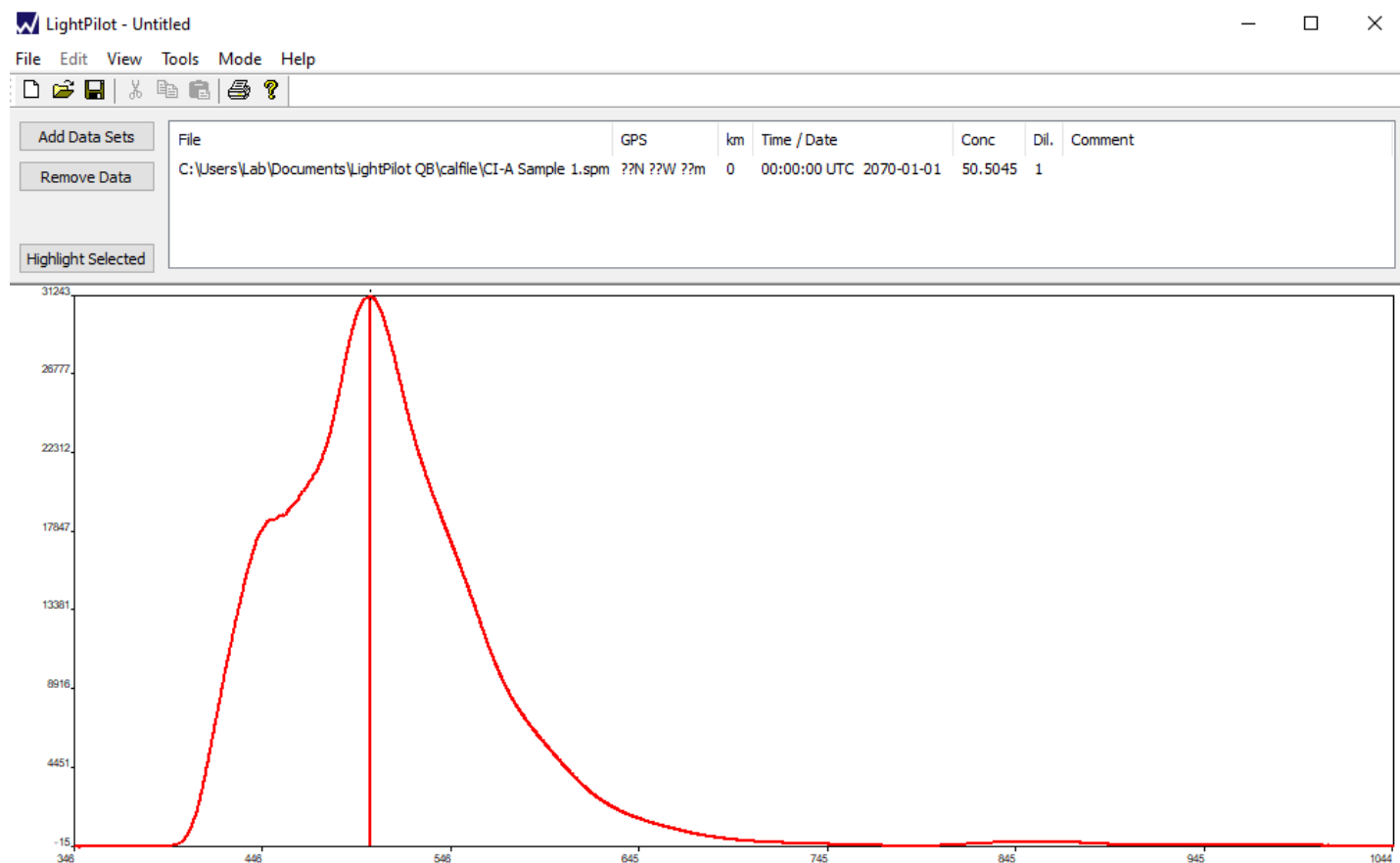
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Figure 23. The data file that was selected is now displayed in the Review Mode.

Repeat the process to review and compare multiple spectra files at once. Finally, a report can be produced, previewed, and printed. To view your report, click on 'View', 'Graph Type', then 'Report'.

If you wish to print, click on 'File' then 'Print' or 'Print Preview' (see Figure 24). Note that you can print as pdf to save the report as a pdf file (see Figure 25). If you have multiple spectra displayed, a report will be printed for all the spectra at once. If you wish to only print one report, you must first remove the undesired spectra by selecting them and then clicking on 'Remove Data' until only the desired spectrum remains.

The data report includes a copy of the spectrum, the calibration curve, results, and sample information. (see Figure 25).
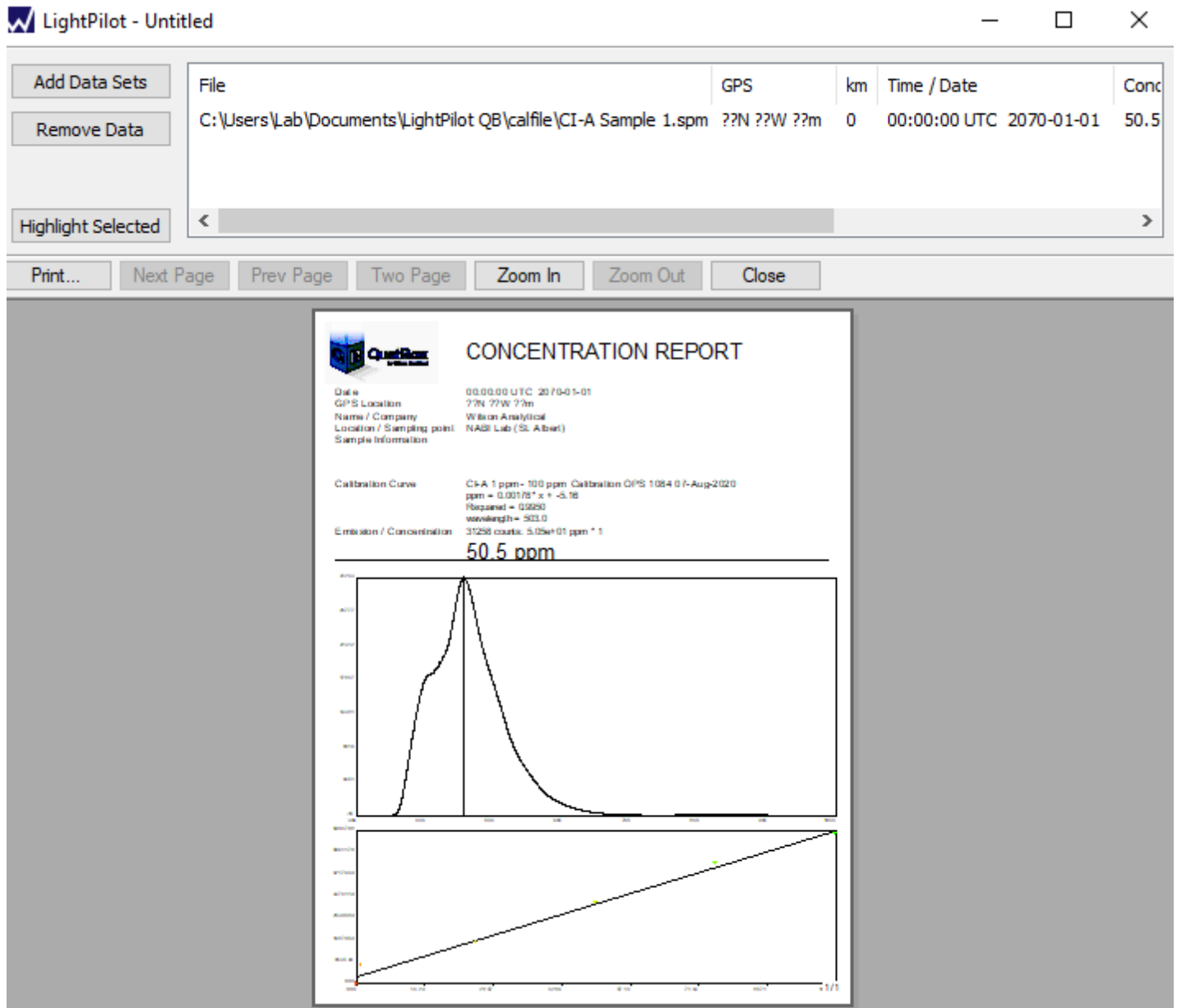
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Figure 24. The Print Preview of the Report generated by LightPilot.

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CONCENTRATION REPORT

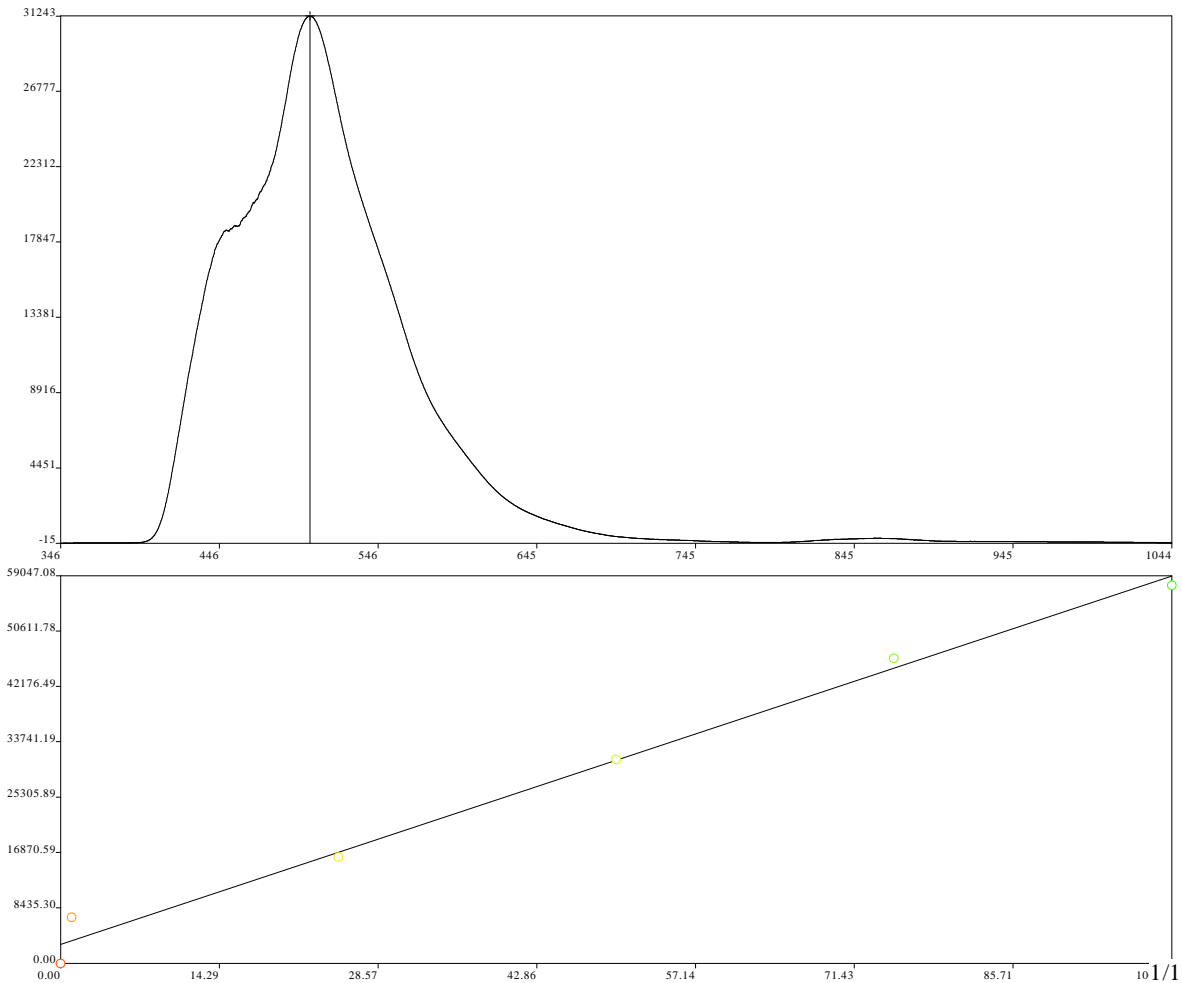|	
Date	00:00:00 UTC 2070-01-01
GPS Location	??N ??W ??m
Name / Company	Wilson Analytical
Location / Sampling point	NABI Lab (St. Albert)
Sample Information	
Calibration Curve	CI-A 1 ppm- 100 ppm Calibration OPS 1084 07-Aug-2020
	ppm = 0.00178 * x + -5.16 Rsquared = 0.9950 wavelength = 503.0
Emission / Concentration	31258 counts: 5.05e+01 ppm * 1
	50.5 ppm

Figure 25. The Print PDF of the Report generated by LightPilot.

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Appendix – Wilson Analytical LightPilot

Importing LightPilot Data into Excel

When using a Windows PC, you may open your .spm files or .cal files using Microsoft Excel. From the Excel window, click File, Open, then Browse to the data file you want to import. By default, these files are stored in the LightPilot subfolder of the user's Documents directory. If you are unable to see the file, make sure you have selected "All Files (*.*)" in the dialog box.

If you are using a Mac, open your .spm files or .cal files by first opening Excel. On the toolbar, click on Data, Get External Data, From Text (Legacy), then select the data you want to import.

The Text Import Wizard guides you through the steps required to load the data into Excel. Select the options as shown in the images below.
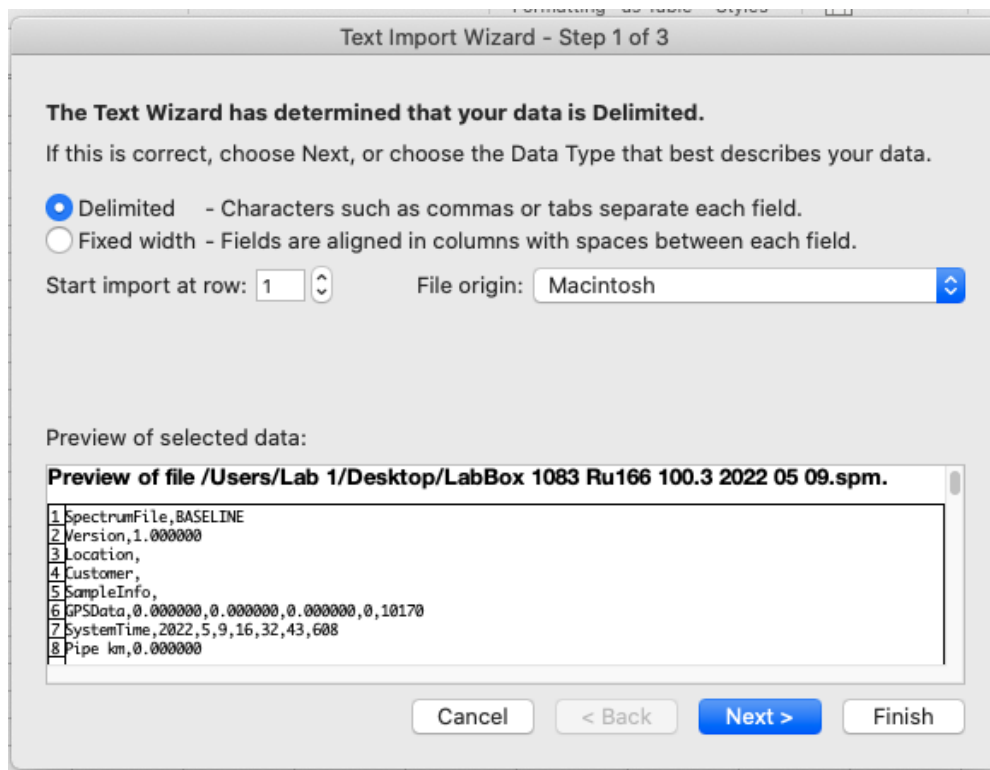

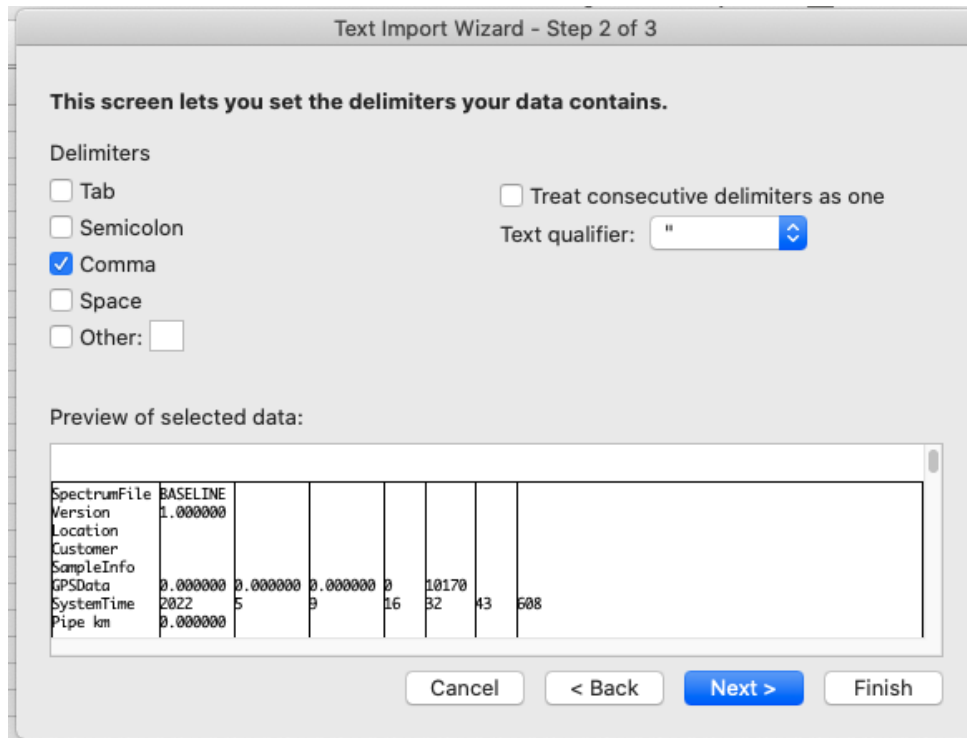**Figure 26**. Step one of three of the Text Import Wizard.

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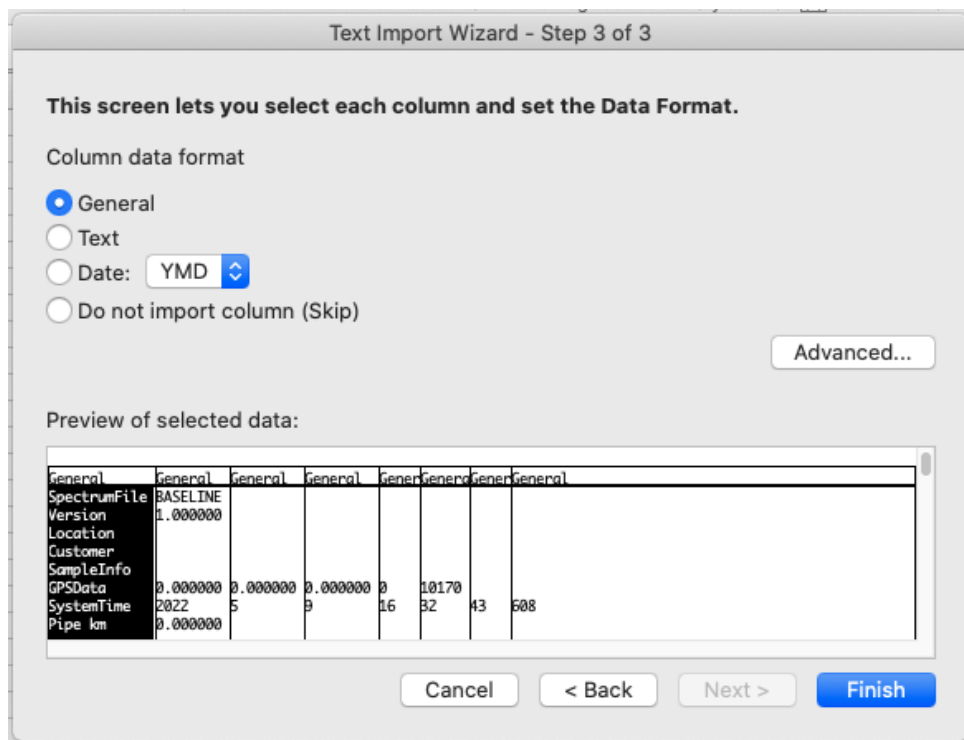**Figure 27**. Step two of three of the Text Import Wizard.

Figure 28. Step three of three of the Text Import Wizard.

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Here are some things to remember when you are looking at the data:

For .spm files:
- You will see your baseline data (after the line "SpectrumFile,BASELINE"), followed by raw data (after the line "SpectrumFile,RAW DATA"), then difference data (after the line "SpectrumFile,DIFF DATA").
- Difference data intensity (counts) = raw data intensity (counts) - baseline intensity (counts).
    - If you want to look at the baseline corrected data, refer to the difference data.
    - If you want the raw count data that does not account for the baseline, use the raw data.

For .cal files:
- You will see all your summary calibration data at the beginning of the file. These data typically appear in the dialog bar on the left-hand side of LightPilot when in Control Mode.
- After the summary data, you will see data defining the spectrum for each of the calibration points. The first set of data will have "Concentration,0", which is the baseline.
- The spectrum data will appear in the same order that you used when acquiring your calibration samples. For example, if you measured 75ppm, then 10ppm, then 50ppm, then 25ppm, then 100ppm, the spectrum data will appear in the same order. If in doubt, check the "Concentration" line that precedes each spectrum data series for the concentration value.
    - The intensity (counts) values displayed for your concentrations are the raw values.
    - If you would like difference values, you must subtract the baseline intensity (counts) value (for concentration 0) from the raw intensity values for the desired concentrations. LightPilot uses the difference values to calculate the calibration curve.

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Wilson File Formats: CAL

The .cal files contain calibration information and the spectra it was derived from. In order to allow later error analysis, these files contain
- a header with calibration information
- the individual raw spectra
  - if a zero concentration spectrum is present, it will be subtracted from all spectra to generate the calibration curve
  - if a zero concentration spectrum is not present, zero concentration will be extrapolated

Format Header

Apart from the beginning and end marker, all fields in this block can occur in arbitrary order. However, if multiple definitions for one line exist, only the last line will be honored by LightPilot.
Some text fields can contain problematic characters such as newlines, commas etc. These are saved using escaped strings which replace problematic characters with \xxx sequences using the standard letter notations for common escapes and an octal code otherwise.

- CalFile
  - Beginning of calibration info block
- EndCalFile
  - End of calibration info block
- Version, *version (double)*
  - a file format version identifier
- Unique File ID, *id (string)*
  - to make sure a calibration file can be found again in the future. Currently these IDs are created and written to the calibration files but may *not* be propagated to the spectra acquired with them!
- Requires Spectrometer, *name (string), serial (string)*
  - spectrometer name / serial in plain text
- Requires Spectrometer iString, *name (string), serial (string)*
  - spectrometer name /serial in obfuscated format
- Calibration Wavelength [nm], *wavelen (double)*
  - calibration wavelength
- Calibration Integration Time [ms], *time (double)*
  - integration time [ms]
- Calibration Spectra to Average, *number (int)*
  - number of spectra to average
- Calibration Boxcar Width, *number (int)*
  - boxcar width (hamming window?)
- Substance, *name (string)*
  - substance / quat / curve name displayed in acquisition screen
- Calibration Points Wavelength [nm], *conc (double), ... (double)*
  - NOTE: misnomer ... these are the concentration values given for the individual cal spectra

- Calibration Points Intensity [counts], *conc (double), ... (double)*

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- o these are the intensity values given for the individual cal spectra. Note that these are floating point numbers because of averaging
- Calibration Points Intensity (raw) [counts], *conc (double), ... (double)*
  - o these are the raw intensity values (without subtracting baseline) given for the individual cal spectra. Note that these are floating point numbers because of averaging
- Calibration Points Baseline [counts], *conc (double)*
  - o this is supposed to be the number of counts (baseline) to subtract. In some older spectrum files this may be a very large integer value due to a bug in the writing code.
- Calibration Type, *number (int)*
  - o fit type of calibration curve. Powers 2-4 are supported internally but LightPilot will only allow setting "2" = linear.
- Calibration Parameters, *c0 (double), c1 (double), ... (double)*
  - o The subsequent paramters are the constants for powers of x (0,1,2, ...)
- Calibration Rsquare, *wavelength (double)*
  - o R-square value calculated for calibration curve
- Conc. Threshold Low Warning [ppm], *bounds (double)*
  - o lower warning bounds [ppm] ... the displayed number will turn orange if the calculated concentration drops below this point
- Conc. Threshold High Warning [ppm], *bounds (double)*
  - o upper warning bounds [ppm] ... the displayed number will turn orange if the calculated concentration rises above this point
- Conc. Threshold Low Error [ppm], *bounds (double)*
  - o lower error bounds [ppm] ... the displayed number will turn red if the calculated concentration drops below this point
- Conc. Threshold High Error [ppm], *bounds (double)*
  - o upper error bounds [ppm] ... the displayed number will turn red if the calculated concentration rises above this point

Format Spectra

Individual spectrum data blocks will follow the initial header.

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Wilson File Formats: CFG

The .cfg files contain configuration parameters for the program. By default, spec4.cnf will be loaded but any other file may be loaded manually from control mode.

Format

As opposed to all other files, the CNF file does not have a begin and marker or a version number.
- GPS, *port (string)*
  - port on which to look for GPS
- Spectrometer, *name (string)*
  - name / serial of GPS to use by default. If this GPS is not found a warning may be issued
- Mode, *number (string)*
  - default mode for LightPilot to start up in:
    - 0 - Acquisition
    - 1 - Control
    - 2 - Review
- Password (clear), *password (string)*
  - password required for control mode in clear text (this is just to prevent accidental tampering)
- Require GPS, *number (boolean)*
  - whether to require GPS to start LightPilot
    - 0 - no (a warning will be issued if GPS is not found)
    - 1 - yes (software will exit if GPS is not found)
- Require Spec, *number (boolean)*
  - whether to require a spectrometer to start up
    - 0 - no (don't use this, as it will allow acquisition modes without spectrometer)
    - 1 - yes (a warning will be issued an only review mode will be available without spectrometer)
- Verbose Acquisition, *number (boolean)*
  - whether to display additional request for sample change before displaying customer info dialog for the second time
    - 0 - no (default)
    - 1 - yes
- Autosave Immediate, *number (boolean)*
  - whether to automatically open a save dialog after data has been acquired
    - 0 - no (default)
    - 1 - yes (subsequent saving is still possible)
- Autosave All, *number (boolean)*
  - Not currently implemented. Intended to make secondary secure copy
- Autosave Warnings, *number (boolean)*
  - Should the program warn if unsaved data might be lost
    - 0 - no
    - 1 - yes (default)

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