RNA clean up

User manual

NucleoSpin® RNA Clean-up

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1 Components

1.1 Kit contents

	NucleoSpin® RNA Clean-up		
REF	**10 preps 740948.10**	**50 preps 740948.50**	**250 preps 740948.250**
Lysis Buffer RA1	10 mL	25 mL	125 mL
Wash Buffer RA2	15 mL	15 mL	80 mL
Wash Buffer RA3 (Concentrate)*	6 mL	12 mL	3 x 25 mL
RNase-free H_2O	13 mL	13 mL	60 mL
NucleoSpin® RNA Clean-up Columns (light blue rings – plus Collection Tubes)	10	50	250
Collection Tubes (2 mL)	10	50	250
Collection Tubes (1.5 mL)	10	50	250
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1.2 Reagents, consumables, and equipment to be supplied by user

Reagents

- 96–100 % ethanol (to prepare Wash Buffer RA3 and to adjust RNA binding conditions)

Consumables

- 1.5 mL microcentrifuge tubes
- Sterile RNase-free tips

Equipment

- Manual pipettors
- Centrifuge for microcentrifuge tubes
- Personal protection equipment (e.g., lab coat, gloves, goggles)

* For preparation of working solutions and storage conditions see section 3.

2 Product description

2.1 The basic principle

One of the most important aspects in the isolation and handling of RNA is to prevent degradation of the RNA during the isolation procedure. With the **NucleoSpin® RNA Clean-up** kit, RNA containing samples are mixed with a solution containing large amounts of chaotropic ions. This solution immediately inactivates RNases – which are present in virtually all biological materials – and creates appropriate binding conditions which favor adsorption of RNA to the silica membrane. Simple washing steps remove salts, metabolites, organics like phenol, and macromolecular cellular components. Pure RNA is finally eluted under low ionic strength conditions with RNase-free water (supplied).

The RNA clean up preparation using **NucleoSpin® RNA Clean-up** kits can be performed at room temperature. The eluate, however, should be treated with care because RNA is very sensitive to trace contaminations of RNases, often found on general lab ware, fingerprints and dust. To ensure RNA stability keep RNA frozen at -20 °C for short-term or -70 °C for long-term storage.

2.2 Kit specifications

- **NucleoSpin® RNA Clean-up** kits are ideal for the clean up of total RNA from RNA preparations which contain inacceptable amounts of RT-PCR inhibitors (e.g., RNA prepared with phenol-chloroform based methods).
- The kit is further recommended for the isolation of RNA from small amounts of cultured cells whenever copurification of some genomic DNA is acceptable. The kit allows for purification of pure RNA with an A_{260}/A_{280} ratio generally exceeding 1.9 (measured in TE buffer (pH 7.5).
- **NucleoSpin® RNA Clean-up** kits are recommended for the clean up of RNA from enzymatic reactions such as *in vitro* transcribed RNA, amplification reactions, biotinylated RNA, or fluorescent (Cy dye) labeled RNA.
- The purified RNA is ready to use for applications such as enzymatic labeling reactions (e.g., dye incorporation), reverse transcriptase-PCR (RT-PCR), as well as for most other downstream appplications.
- The standard protocol (section 5.1) allows for the cleanup of up to 200 µg of RNA per NucleoSpin® RNA Clean-up Column or the isolation of total RNA from up to 1 x 10^5 cultured cells (section 5.2).

Table 1: Kit specifications at a glance

Parameter	NucleoSpin® RNA Clean-up
Technology	Silica membrane technology
Format	Mini spin columns
Sample material	< 100 µL RNA sample with single column loading containing up to 200 µg RNA < 200 µL RNA sample with double column loading containing up to 200 µg RNA Up to 10^5 cells
Fragment size	> 200 nt
Typical recovery (0.1–200 µg RNA input)	85–95 %
A_{260}/A_{280}	1.9–2.1
Elution volume	40–120 µL
Preparation time	Approx. 20 min/6 preps
Binding capacity	200 µg

2.3 Handling, preparation, and storage of starting materials

RNA intended to be used as sample for the **NucleoSpin® RNA Clean-up** procedure should be handled with the same care as any RNA sample. The stability of prepurified RNA samples (e.g., RNA isolated with phenol based protocols) depends very much on the performed procedure. RNA in biological samples is not protected against digestion until the sample material is flash frozen or disrupted in the presence of RNase inhibiting or denaturing agents. Therefore it is important that biological samples are flash frozen in liquid N_2 immediately and stored at -70 °C or processed as soon as possible. Samples can be stored in lysis buffer after disruption at -70 °C for up to one year, at +4 °C for up to 24 hours or up to several hours at room temperature. Frozen samples are stable up to 6 months. Frozen samples in lysis buffer should be thawed slowly before starting with the isolation of total RNA.

Wear gloves at all times during the preparation. Change gloves frequently.

2.4 Elution procedures

It is possible to adjust the elution method and the volume of RNase-free water used for the subsequent application of interest. In addition to the standard method described in the individual protocols (recovery rate about 70–90 %) there are several modifications possible:

- **High yield:** Perform two elution steps with the volume indicated in the individual protocol. About 90–100 % of bound nucleic acid will be eluted.
- **High yield and high concentration:** Elute with the standard elution volume and apply the eluate once more onto the column for reelution.

Eluted RNA should immediately be placed and always kept on ice for optimal stability because almost omnipresent RNases (general lab ware, fingerprints, dust) will degrade RNA. For short term storage freeze at -20 °C, for long term storage freeze at -70 °C.

3 Storage conditions and preparation of working solutions

Attention: Buffers RA1 and RA2 contain chaotropic salt. Wear gloves and goggles!

CAUTION: Buffers RA1 and RA2 contain guanidinium thiocyanate which can form highly reactive compounds when combined with bleach (sodium hypochlorite). DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

- All kit components should be stored at room temperature (18–25 °C) and are stable up to one year. Storage at lower temperatures may cause precipitation of salts.
- Check that 96–100 % ethanol is available as additional solution in the lab.

Before starting any **NucleoSpin® RNA Clean-up** protocol, prepare the following:

- **Wash Buffer RA3**: Add the indicated volume of 96–100 % ethanol (see table below) to Wash Buffer RA3 Concentrate. Mark the label of the bottle to indicate that ethanol was added. Store Wash Buffer RA3 at room temperature (18–25 °C) for up to one year.

	NucleoSpin® RNA Clean-up		
REF	**10 preps 740948.10**	**50 preps 740948.50**	**250 preps 740948.250**
Wash Buffer RA3 (Concentrate)	6 mL Add 24 mL ethanol	12 mL Add 48 mL ethanol	3 x 25 mL Add 100 mL ethanol to each bottle

4 Safety instructions

The following components of the **NucleoSpin® RNA Clean-up** kits contain hazardous contents.

Wear gloves and goggles and follow the safety instructions given in this section.

Only harmful features do not need to be labeled with H and P phrases up to 125 mL or 125 g.
Mindergefährliche Eigenschaften müssen bis 125 mL oder 125 g nicht mit H- und P-Sätzen gekennzeichnet werden.

Component ***Inhalt***	**Hazard contents** ***Gefahrstoff***	**GHS symbol** ***GHS-Symbol***	**Hazard phrases** ***H-Sätze***	**Precaution phrases** ***P-Sätze***
RA1	guanidinium thiocyanate 45–60 % *Guanidinthiocyanat 45–60 %* CAS 593-84-0	 WARNING *ACHTUNG*	302, 412	264W, 273, 301+312, 330
RA2	guanidinium thiocyanate 30–45 % and ethanol 20–35 % *Guanidinhydrochlorid 30–45 % und Ethanol 20–35 %* CAS 50-01-1, 64-17-5	 WARNING *ACHTUNG*	226, 302, 412	210, 264W, 273, 301+312, 330

Hazard phrases

H 226	Flammable liquid and vapour. *Flüssigkeit und Dampf entzündbar.*
H 302	Harmful if swallowed. *Gesundheitsschädlich bei Verschlucken.*
H 412	Harmful to aquatic life with long lasting effects. *Schädlich für Wasserorganismen, mit langfristiger Wirkung.*

Precaution phrases

P 210	Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking. *Von Hitze, heissen Oberflächen, Funken, offenen Flammen sowie anderen Zündquellenarten fernhalten. Nicht rauchen.*
P 264W	Wash with water thoroughly after handling. *Nach Gebrauch mit Wasser gründlich waschen.*
P 273	Avoid release to the environment. *Freisetzung in die Umwelt vermeiden.*
P 301+312	IF SWALLOWED: Call a POISON CENTER / doctor / … / if you feel unwell. *BEI VERSCHLUCKEN: Bei Unwohlsein GIFTINFORMATIONSZENTRUM/Arzt / … anrufen.*
P 330	Rinse mouth. *Mund ausspülen.*

For further information please see Material Safety Data Sheets (***www.mn-net.com***).
*Weiterführende Informationen finden Sie in den Sicherheitsdatenblättern (**www.mn-net.com**).*

⚠ The symbol shown on labels refers to further safety information in this section.
Das auf Etiketten dargestellte Symbol weist auf weitere Sicherheitsinformationen dieses Kapitels hin.

5 Protocols

5.1 RNA Clean-up

Before starting the preparation:

- Check if Wash Buffer RA3 was prepared according to section 3.

1 Sample preparation

Fill up RNA samples smaller than 100 µL with RNase-free water to **100 µL**.

RNA samples from 100–200 µL should be filled up with RNase-free water to 200 µL.

Fill up RNA sample to 100 µL with water

2 Preparation of lysis-binding buffer premix

Prepare a Buffer RA1-ethanol premix with a ratio of 1:1.

For each **100 µL RNA sample** mix **300 µL Buffer RA1** and **300 µL of ethanol (96–100 %)**.

If multiple samples are processed, the preparation of a master-premix is recommended (e.g., 2 mL Buffer RA1 + 2 mL 98 % ethanol for approximately 6 preparations).

Prepare premix:

Mix 300 µL RA1 with 300 µL ethanol (96–100 %)

3 Adjust RNA binding conditions

To **100 µL RNA sample** add **600 µL (6 volumes) of Buffer RA1-ethanol-premix**. Mix sample with premix by vortexing.

If a 200 µL RNA sample is processed, add 1200 µL Buffer RA1-ethanol premix.

After addition of ethanol, a stringy precipitate may become visible which will not affect the RNA isolation. Be sure to mix thouroughly and apply sample as homogeneous solution onto the column.

+ 6 vol. premix

Mix

4 Bind RNA

For each preparation, take one NucleoSpin® RNA Clean-up Column (light blue ring) placed in a Collection Tube and load the lysate (700 µL).

Centrifuge for **30 s** at **8,000 x *g***. Discard Collection Tube with flowthrough and place the column in a new Collection Tube.

Maximal loading capacity of NucleoSpin® RNA Clean-up Columns is 750 µL. Repeat the procedure if larger volumes are to be processed.

Load 700 µL lysate

8,000 x *g*, 30 s

5 Wash and dry silica membrane

1st wash

Add **700 µL Buffer RA3** to the NucleoSpin® RNA Clean-up Column. Centrifuge for **30 s** at **8,000 x *g***. Discard flowthrough and reuse Collection Tube.

+ 700 µL RA3

8,000 x *g*, 30 s

2nd wash

Add **350 µL Buffer RA3** to the NucleoSpin® RNA Clean-up Column. Centrifuge for **2 min** at **8,000 x *g***.

Transfer the NucleoSpin® RNA Clean-up Column to a nuclease-free Collection Tube (1.5 mL, supplied). Open the lid of the column and let the membrane dry for 3 min.

If for any reason, the liquid level in the Collection Tube has reached the NucleoSpin® RNA Clean-up Column after centrifugation, discard flowthrough and centrifuge again.

The procedure ensures complete removal of ethanol from the column.

+ 350 µL RA3

8,000 x *g*, 2 min

6 Elute RNA

Elute the RNA in **60 µL RNase-free H_2O** (supplied) and centrifuge at **8,000 x *g*** for **1 min**.

If higher RNA concentrations are desired, elution can be done with 40 µL. Overall yield, however, will decrease when using smaller volumes.

For further alternative elution procedures see section 2.4.

+ 60 µL RNase-free H_2O

8,000 x *g*, 1 min

5.2 RNA isolation from up to 10^5 cells

Before starting the preparation:

- Check if Wash Buffer RA3 was prepared according to section 3.

1 Sample preparation

As sample material use **up to 10^5 cells** in a volume of up to **100 µL**.

Fill up sample to 100 µL (e.g. with PBS)

2 Cell lysis

Add **300 µL Buffer RA1** and vortex vigorously in order to lyse the cells.

+ 300 µL RA1

Vortex

3 Adjust RNA binding conditions

Add **300 µL ethanol (96–100 %)** to the lysate and mix by vortexing or pipetting up and down.

After addition of ethanol, a stringy precipitate may become visible which will not affect the RNA isolation. Be sure to mix thoroughly and apply sample as homogeneous solution onto the column.

+ 300 µL ethanol (96–100 %)

Mix

4 Bind RNA

For each preparation, take one NucleoSpin® RNA Clean-up Column (light blue) placed in a Collection Tube and load the lysate (700 µL).

Centrifuge for **30 s** at **8,000 x *g***. Discard Collection Tube with flowthrough and place the column in a new Collection Tube.

Maximal loading capacity of NucleoSpin® RNA Clean-up Columns is 750 µL. Repeat the procedure if larger volumes are to be processed.

Load lysate

8,000 x *g*, 30 s

5 Wash and dry silica membrane

1st wash

Add **250 µL Buffer RA2** to the NucleoSpin® RNA Clean-up Column. Centrifuge for **30 s** at **8,000 x *g***. Discard flowthrough and reuse Collection Tube.

+ 250 µL RA2

8,000 x *g*, 30 s

2nd wash

Add **700 µL Buffer RA3** to the NucleoSpin® RNA Clean-up Column. Centrifuge for **30 s** at **8,000 x *g***. Discard flowthrough and reuse Collection Tube.

+ 700 µL RA3

8,000 x *g*, 30 s

3rd wash

Add **350 µL Buffer RA3** to the NucleoSpin® RNA Binding Column. Centrifuge for **2 min** at **8,000 x *g***.

Transfer the NucleoSpin® RNA Clean-up Column to a nuclease-free Collection Tube (1.5 mL, supplied). Open the lid of the column and let the membrane dry for 3 min.

If for any reason, the liquid level in the Collection Tube has reached the NucleoSpin® RNA Clean-up Column after centrifugation, discard flowthrough and centrifuge again.

The procedure ensures complete removal of ethanol from the column.

+ 350 µL RA3

8,000 x *g*, 2 min

6 Elute RNA

Elute the RNA in **60 µL RNase-free H_2O,** (supplied) and immediately centrifuge at **8,000 x *g*** for **1 min**.

If higher RNA concentrations are desired, elution can be done with 40 µL. Overall yield, however, will decrease when using smaller volumes.

For further alternative elution procedures see section 2.4.

+ 60 µL RNase-free H_2O

8,000 x *g*, 1 min

6 Appendix

6.1 Troubleshooting

Problem	Possible cause and suggestions
RNA is degraded/no RNA obtained	*RNase contamination* • Create an RNase-free working environment. Wear gloves during all steps of the procedure. Change gloves frequently. Use of sterile, disposable polypropylene tubes is recommended. Keep tubes closed whenever possible during the preparation. Glassware should be oven baked for at least 2 hours at 250 °C before use.
Poor RNA quality or yield	*Reagents not applied or restored properly* • Sample and reagents have not been mixed completely. Always vortex vigorously after each reagent has been added. • No ethanol has been added. Binding of RNA to the silica membrane is only effective in the presence of ethanol. *Kit storage* • Store kit components at room temperature. Storage at low temperatures may cause salt precipitation. If salt precipitates are visible, warm up to 37 °C until all salt precipitates are dissolved. • Keep bottles tightly closed in order to prevent evaporation or contamination. *Sample material* • Sample material not stored properly. Whenever possible, use fresh material. If this is not possible, flash freeze the samples in liquid N_2. Samples should always be kept at -70 °C. Never allow tissues to thaw before addition of lysis buffer. Perform disruption of samples in liquid N_2.
Contamination of RNA with genomic DNA	• The NucleoSpin® RNA Clean-up procedure does not comprise a DNA digestion step. Therefore, the extent of DNA contamination mainly depends on the sample material. If lowest level of DNA contamination is desired, use one of the rDNase containing NucleoSpin® RNA kits (see ordering information).

Problem	Possible cause and suggestions
Suboptimal performance of RNA in downstream experiments	*Carry over of ethanol or salt* • Do not let the flowthrough touch the column outlet after the second wash using Wash Buffer RA3. Be sure to centrifuge at the corresponding speed for the respective time in order to remove ethanolic Wash Buffer RA3 completely. • Check if Wash Buffer RA3 has been equilibrated to room temperature before use. Washing at lower temperatures lowers efficiency of salt removal by Wash Buffer RA3. • A 2 min centrifugation with a subsequent 3 min drying with open lid is sufficent for an extensive removal of ethanol from the column. Residual ethanol will typically be around 1 %. Increasing the drying step with open lid from 3 min to 20 min will decrease the residual ethanol content commonly to below 0.1 %, but also RNA recovery will be reduced 5–20 %. *Store isolated RNA properly* • Eluted RNA should always be kept on ice for optimal stability since trace contaminations of omnipresent RNases (general lab ware, fingerprints, dust) will degrade the isolated RNA. For short term storage freeze at -20 °C, for long term storage freeze at -70 °C. *RNA concentration is too low* • For highest RNA concentration and most sensitive downstream applications, NucleoSpin® RNA Clean-up XS is recommended. NucleoSpin® RNA Clean-up XS allows elution in only 5–20 µL volume (see ordering information).
Higher RNA yield than theoretically possible	• If performing clean-up of samples containing less than approximately 300 ng, RNA subsequent quantification by A_{260} measurement may simulate yields larger than the RNA input. This may be due to absorbance of silica abrasion. In order to prevent incorrect A_{260} quantification of small RNA amounts, centrifuge the elution tube for 30 s at 8.000–11.000 x *g* and withdraw an aliquot for measurement without disturbing any sediment or use a silica abrasion insensitive RNA quantification method (e.g., RiboGreen® fluorescent dye).

6.2 Ordering information

Product	REF	Pack of
NucleoSpin® RNA Clean-up	740948.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA Clean-up XS	740903.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA	740955.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA Plus	740984.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA XS	740902.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA Plus XS	740990.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA Midi	740962.20	20 preps
NucleoSpin® miRNA	740971.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA Blood	740200.10 / 50	10 / 50 preps
NucleoSpin® totalRNA FFPE	740982.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® totalRNA FFPE XS	740969.10 / .50 / .250	10 / 50 / 250 preps
NucleoSpin® RNA Plant and Fungi	740120.10 / .50 / .250	10 / 50 / 250 preps
NucleoBond® RNA Soil	740140.20	20 preps
NucleoBond® RNA Soil Mini	740142.10 / .50	10 / 50 preps
NucleoSpin® RNA Stool	740130.10 / .50	10 / 50 preps

Visit ***www.mn-net.com*** for more detailed product information.

6.3 Product use restriction / warranty

NucleoSpin® RNA Clean-up kit components are intended, developed, designed, and sold FOR RESEARCH PURPOSES ONLY, except, however, any other function of the product being expressly described in original MACHEREY-NAGEL product leaflets.

MACHEREY-NAGEL products are intended for GENERAL LABORATORY USE ONLY! MACHEREY-NAGEL products are suited for QUALIFIED PERSONNEL ONLY! MACHEREY-NAGEL products shall in any event only be used wearing adequate PROTECTIVE CLOTHING. For detailed information please refer to the respective Material Safety Data Sheet of the product! MACHEREY-NAGEL products shall exclusively be used in an ADEQUATE TEST ENVIRONMENT. MACHEREY-NAGEL does not assume any responsibility for damages due to improper application of our products in other fields of application. Application on the human body is STRICTLY FORBIDDEN. The respective user is liable for any and all damages resulting from such application.

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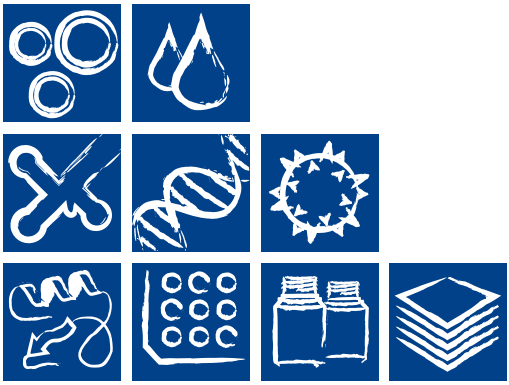

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