

Openly Glowing Plant #1 SynMemo

by Andreas Stürmer, Justin Atkin and Sebastian Cocioba

This is a follow up to Openly Glowing Plant #1 memo.

We propose synthesis of 13 DNA products in the range of 0.5kb-5kb. All pass Twist's gene checks.

The DNA will be transformed into plants by a true master in the field. Sebastian Cocioba runs the educational non-profit Binomica labs, as well as New York Botanics. In his work, he has done nuclear and plastid transformation of various plants such as tobacco, petunia, arabidopsis and wheat. Most frequently he employs agrobacterium, but he also possesses a functioning gene gun. He also has skills in protoplast transformation.

The research builds upon the recent discovery of the fungal metabolic pathway for light emission through the caffeic acid cycle.

7 different DNA fragments are luciferases. 3 are wild type to compare function between source species. 2 are rationally designed luciferase mutants to attempt to shift the color. 1 is a closely related (found through BLAST) fungal enzyme that will be checked for luciferase activity and finally one is an RFP-tagged Luciferase again to attempt a color shift [1]. One of the two luciferin synthesis enzymes is too big to be synthesized as a 5000 bp brick so it will be split up into two parts and will be ligated by BglII digestion.

3 of the fragments are expression cassettes that contain a promoter and terminator and will fit into the pCambia2300 multiple cloning site. The promoter and terminator in these fragments will be AtTCT Promoter and HSP-Terminator of Arabidopsis. [2]

The wt luciferase and the two luciferin biosynthesis enzymes will be cloned into the respective expression cassette to produce the final plasmids before transformation into a selection of plants (primarily tobacco to start) to assay for expression, function, and glow.

Once the wild type and rationally designed variants are assayed, new alternative luciferases will be produced by mutating the wt enzymes (using manganese PCR) and then re-subcloned into the vectors and checked for kinetic properties and shifted colour emission.

We designed mutants versions of the luciferase, which change the putative binding pocket and believe they can shift the light emission wavelength. The pockets were found by aligning the known fungal luciferases from the paper and looking for conserved domains. We adapted the reasoning from a firefly luciferase color-shift paper, where Y->F [3] is a relatively conservative mutation and causes the enzyme to alter its structure by just a little bit. Besides side-chain size

Omphalotus olearius	random mutation	<pre> TTTTGCTTTGTTAGACGAAGCAAAAGGATGTTGCTATTATGATGATTGCTGA AAAGAGAAGATGGTTGGGAGATTGGCCTTTAGAGAGGACCTAGACCTCTATTACT TCTCATATTATTCAAAGCAAGAACTcagTGGCTGATGCTGAATTTGCTACTAAAGGA ATTGATGGAAAGATATCTTAGAGTTCAGCTAGACACTAATACTACTTTTTGT CTACTCTAAAGTTGAATTCGATGCTCAAGCTATTTTTTGTGCTCTCTACTCTATT AATGATCCTCAAAATATCTCTCATGATACTGTTAGAAAGAACTAAGAGAAATGG CTcagATGCAAGATATCATGATTTGACTTTGCTATTGCTTTGGCTGCTCAAGATGGA AAGGAGCTTTTGGAAAGGATGGGACAGACACTCTTTGCTGCTGGACCTGGAGTT CCTGGACCTCTACTGAATGGACTTTTTTGTATGCTGccagTCTGAAAGAAAGATTAG AGTTGTTGAAATGATTTGTTgggGCTTCTGTTGTTATATGACTAATGATCCTGGCTGATA AGATTGTTGAAGCTACTGTTCAAGGAACTGAAGAAaagGAAGCTCagatgaagat </pre>	
Fungal Luciferase of <i>Mycena citricolor</i>	different luciferase for random mutation	<pre> AATAAGCGACCATGGCTTATCAmhaACTTGGATTCAAACTTTGGTTTTGGAGCTTGG GTTGCTATGGCTGTTGCTTTTCTTTTAAAGAGGATATGAAACTTTTTTGAAGGG AGGACCTCTATGCTCTCAAAATGTTAGAGGATATATTATTTGGTTTTGGCTT TGTTAGACAAGAacagTGGGATTTGAAATTTATGATAGAATGCTGAAAGAGAGAG ATGTTGGTAATTTGGCTCAAGAGAGAGGACCTAGACCTAGAGACTCTCTCATATT ATTCAAGAcagTGTCTCAACATACTGATCCTGCTTTGGAGCTGCTATTGAAAGGA TACTGTTATTCCTAGAGTTCAAGCTAGACATGCTGCTAATACTCATATTGCTAGAGca CTT TGAATTCAGTCTGCTGCTATTTTTTGAATGCTGATGTTGCTTTGGCTGAAGGA TTGCTGGCTTCTGAAGCTTTAGAGACTAGGGAGAAATTTCTCacATGGATATT ATCATGATTTACTTTGGATTTGGCTTTGGCTGCTGCTGATGGAAGGAGTTGTTGG AAAGGATGGGACAAAGACATCCTTTGGCTGGACCTGGAGTCTGGACCTCTAA TGAATGAGCTTTGTTGTTGCTGTAAGAAAGAGAGAAATGGAGTTGTTGAACAA ATTGTTGAAGCTGCTATTGGATATATGCTATTTGCTGCTTTGGAAaagGAGCTCa gatatgaagat </pre>	771

[1] <https://www.ncbi.nlm.nih.gov/pubmed/30478037> Kotlobay AA, Sarkisyan KS, Mokrushina YA, Marcet-Houben M, Serebrovskaya EO, Markina NM, Gonzalez Somermeyer L, Gorokhovatsky AY, Vvedensky A, Purtov KV, Petushkov VN, Rodionova NS, Chepurnyh TV, Fakhranurova LI, Guglya EB, Ziganshin R, Tsarkova AS, Kaskova ZM, Shender V, Abakumov M, Abakumova TO, Povolotskaya IS, Eroshkin FM, Zaraisky AG, Mishin AS, Dolgov SV, Mitiouchkina TY, Kopantzev EP, Waldenmaier HE, Oliveira AG, Oba Y, Barsova E, Bogdanova EA, Gabaldón T, Stevani CV, Lukyanov S, Smirnov IV, Gitelson JI, Kondrashov FA, Yampolsky IV. Genetically encodable bioluminescent system from fungi. Proceedings of the National Academy of Sciences of the United States of America 2018;115(50):12728-32.

[2] <https://www.ncbi.nlm.nih.gov/pubmed/25410250> Han YJ, Kim YM, Hwang OJ, Kim JI. Characterization of a small constitutive promoter from Arabidopsis translationally controlled tumor protein (AtTCTP) gene for plant transformation. Plant cell reports 2015;34(2):265-75.

[3] <https://www.nature.com/articles/srep02490> Wang, Y., Akiyama, H., Terakado, K. et al. Impact of Site-Directed Mutant Luciferase on Quantitative Green and Orange/Red Emission Intensities in Firefly Bioluminescence. Sci Rep 3, 2490 (2013). <https://doi.org/10.1038/srep02490>

[4] <https://link.springer.com/article/10.1007/s13258-016-0417-3> Ning Li, Yuanyuan Li, Chengchao Zheng, Jinguang Huang & Shizhong Zhang. Genome-wide comparative analysis of the codon usage patterns in plants. Genes & Genomics volume 38, pages723–731(2016)

[5] <https://link.springer.com/article/10.1007/s11103-006-0041-8> Liangjiang Wang & Marilyn J. Roossinck. Comparative analysis of expressed sequences reveals a conserved pattern of optimal codon usage in plants. Plant Molecular Biology volume 61, pages699–710(2006)