Sperm Vitality

Concept:

Sperm **membrane integrity** (structural and functional) when questionable or inadequate may compromise sperm fertilizing capacity (by affecting motility, capacitation, acrosome reaction, and binding of sperm on Zona Pellucida).

Membrane integrity is evaluated through two tests:

- 1. Sperm Vitality (Structural Integrity) Supravital Dye Exclusion
- 2. HOS (Hypo-Osmotic Swelling) [Functional Integrity]

Sperm Vitality reflects the number of live **membrane intact spermatozoa**. It is **tested** based on the ability of cell membrane to **exclude vital stains** (Eosin – Nigrosine dye) from entering the **spermatozoa** and **permeate** into its nucleus. When physically **damaged** or **broken**, the eosin Y **dye** is able to **stain sperm**. If membrane is **intact**, the dye is **unable** to do so.

Vitality thus evaluates viability of cell. For example:

- Live (Viable). Unstained
- Dead (Non-Viable). Stained

Note: Sperm Vitality is also an additional quality check for sperm motility evaluation since the number of Live (Viable) sperm, as determined by vitality staining, should be the same or greater than the percent of motile sperm.

Specimen Preparation:

- Semen sample is collected with:
 - Abstinence period of 2 to 7 days.
 - Ideal collection through masturbation in sterile container.
 - Non-spermicidal polyurethane semen collection pouch can be used when required.
- Semen sample should liquefy and then mixed before testing.
- Ideally, the test should be performed within 30 to 60 minutes of the sample collection.

Special Instructions:

- Hyper-viscous semen sample should be processed to bring towards normal viscosity. (BIOSCREEN Viscosity kit can be used to help lower sample viscosity).
- Severe oligospermic semen sample (i.e., sample with Sperm Concentration less than 5 million / mL) should be processed to concentrate the sperm concentration to around 8 to 10 million / mL before performing the test.
- Frozen semen plasma must be thawed at 37°C before performing the test.

Kit Contents: 2.5mL of Eosin-Nigrosin Solution.

REQUIRED BUT NOT PROVIDED IN KIT:

- Distilled Water
- Xylene
- Mounting Solution
- Immersion Oil

Storage Conditions:

- The kit should be stored in dark at 2°C to 8°C after receiving.
- Bring all the reagents to room temperature before use.
- Once opened, store reagents in the fridge protected from light.
- Expiry date is printed on the outside of the box.

Procedure:

- Step 1. Label plasticware and disposable material with appropriate Patient ID and Sample ID.
- Step 2. o Take 10ųL of undiluted, well-mixed liquefied semen.
 - Add 50ųL of Eosin-Nigrosin dye solution in a microtube.
 - Mix the dye and sample thoroughly with the help of pipette tip and wait for 30 seconds.
- Step 3. After 30 seconds, please 10ųL of sample from Step 2 on a clean glass-slide and prepare a smear.
- Step 4. Allow the smear to air dry or use a slide warmer.

Note: Examine the dry smear as soon as possible.

Examination:

- Put a drop of oil immersion on dry smear.
- **Examine** smear under microscope using **100x** lens.
- Unstained / White Sperm (indicative of Live sperm)
 - Red / Dark Pink Sperm (indicative of Dead sperm)

Note: "Leaky necks" sperm stained only in neck region (Heads remain unstained) are considered as live sperm.

Red / dark pink color is more prominent in post acrosomal area of head.

The finding of a high percentage of viable (Live) sperm in the presence of extremely low motility strongly suggests the presence of an ultra-structural (cytoskeleton) sperm defect.

The Supravital Stains are not appropriate for assessing the vitality of the cryo preserved spermatozoa because Glycerol interests with the stain.

The sum of percentage (%) of Dead sperm and motile sperm should not exceed 100%.

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Collection
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Reference Image:



Result:

- Number of Sperm Evaluated: _____. Number Unstained Sperm (Live): Number of Stained Sperm (Dead):
- Normal Value for Live Sperm.
 - 0 Normal. >58%
 - Equivocal. >55% and <63% 0
 - Abnormal. <55% 0

Limitations:

- This test provides presumptive quantitative information of sperm vitality.
- This parameter should be analyzed by a specialist.
- The result should be evaluated considering all clinical and laboratory findings related to the same sample.

Cover Slipping Stained Slides:

- Permanent Stained Slide. •
 - 0 Dip the stained slide in Xylene solution prior to cover slipping.
 - Place the mounting media on the slide. 0
 - Place the coverslip onto the slide as quickly as possible to 0 avoid air drying and air bubbles.

Precautions:

- All patient samples and reagents should be treated as potentially infectious and the user must wear protective gloves, eye protection, and laboratory coats when performing the test.
- The kit should be discarded in a proper biohazard container after testing.
- Do not eat, drink, or smoke in the area where specimens and kit reagents are handled.
- Do not use beyond the expiration date which appears on the package label.
- It is recommended to use gloves and a face mask.

Safety and Environment:

- Do not release the products used into the environment. Follow • guidelines for the storage and disposal of toxic substances.
- Biological samples must be handled as potentially infectious.

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