# 1 Title:

2	Remediating Indoor Pesticide Contamination from Improper Pest Control
3	Treatments: Persistence and Decontamination Studies
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5	Authors and Affiliations:
6	Lukas Oudejans <sup>1,*</sup> , Amy Mysz <sup>2</sup> , Emily Gibb Snyder <sup>1</sup> , Barbara Wyrzykowska-Ceradini <sup>3</sup> , Joshua
7	Nardin <sup>3</sup> , Dennis Tabor <sup>4</sup> , James Starr <sup>5</sup> , Daniel Stout II <sup>5</sup> , and Paul Lemieux <sup>1</sup>
8	
9	<sup>1</sup> U.S. Environmental Protection Agency, Office of Research and Development, Center for

- 10 Environmental Solutions & Emergency Response, Homeland Security & Materials Management
- 11 Division, 109 TW Alexander Dr, Research Triangle Park, NC 27709, United States of America
- 12 <sup>2</sup> U.S. EPA, Region 5, 77 W. Jackson Blvd, Chicago, IL, United States of America
- <sup>3</sup> Jacobs Technology Inc., 600 William Northern Blvd, Tullahoma, TN 37388, United States of
- 14 America
- <sup>4</sup> U.S. EPA, Office of Research and Development, Center for Environmental Measurement &
- 16 Modeling, Air Methods & Characterization Division, 109 TW Alexander Dr, Research Triangle
- 17 Park, NC, United States of America
- <sup>5</sup> U.S. EPA, Office of Research and Development, Center for Environmental Measurement &
- 19 Modeling/Watershed & Ecosystem Characterization Division, 109 TW Alexander Dr, Research
- 20 Triangle Park, NC, United States of America
- 21 \* Corresponding Author:
- 22 Lukas Oudejans,
- 23 E-mail: oudejans.lukas@epa.gov
- 24 Phone: +1 919 541 2973
- 25 Fax: +1 919 541 0496
- 26 Mailing Address: U.S. EPA, Mail Drop E343-06
- 27 109 TW Alexander Dr.
- 28 Research Triangle Park, NC 27711

# 29 Abstract

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31	The improper and excessive use of pesticides in indoor environments can result in adverse
32	human health effects, sometimes necessitating decontamination of residential or commercial
33	buildings. A lack of information on effective approaches to remediate pesticide residues
34	prompted the decontamination and persistence studies described in this study.
35	Decontamination studies evaluated the effectiveness of liquid-based surface decontaminants
36	against pesticides on indoor surfaces. Building materials were contaminated with 25-2,400
37	$\mu$ g/100cm <sup>2</sup> of the pesticides malathion, carbaryl, fipronil, deltamethrin, and permethrin.
38	Decontaminants included both off-the-shelf and specialized solutions representing various
39	chemistries. Pesticides included in this study were found to be highly persistent in a dark indoor
40	environment with surface concentrations virtually unchanged after 140 days. Indoor light
41	conditions degraded some of the pesticides, but estimated half-lives exceeded the study
42	period. Decontamination efficacy results indicated that the application of household bleach or a
43	hydrogen peroxide-based decontaminant offered the highest efficacy, reducing malathion,
44	fipronil, and deltamethrin by >94-99% on some surfaces. Bleach effectively degraded
45	permethrin (>94%), but not carbaryl (<70%) while the hydrogen peroxide containing products
46	degraded carbaryl (>71-99%) but not permethrin (<54%). These results will inform responders,
47	the general public and public health officials on potential decontamination solutions to
48	remediate indoor surfaces.

# 49 Keywords

50 pesticide

- 51 misuse
- 52 decontamination
- 53 cleanup
- 54 persistence

# **1. Introduction**

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56	Under the U.S. Federal Insecticide, Fungicide, and Rodenticide Act (FIFRA), it is a violation to
57	use a pesticide in a manner inconsistent with the product's labeling. Applying pesticides in ways
58	that deviate from the label directions is illegal and considered a misuse or a misapplication.
59	Misusing pesticides can include the application of products not registered by the U.S
60	Environmental Protection Agency (EPA) [1], off-label application in areas where the product is
61	not intended, or at higher rates or concentrations (mass per unit area) than specified on the
62	product label. Misapplications also include disregarding safety instructions and applying
63	restricted-use pesticides without proper applicator certification.
64	Misapplying pesticides in homes, schools, businesses or other indoor environments can lead
65	to adverse health effects and contamination [2-8], often impacting vulnerable populations.
66	Building occupants and federal, state and local agencies responding to pesticide misuse
67	incidents seek information about whether pesticide residues present exposure risk and how to
68	clean treated surfaces to reduce pesticide levels, if necessary [7-10]. This study does not
69	attempt to evaluate whether pesticide levels are unsafe but provides decontamination
70	information should remediation be desired or deemed appropriate.
71	Tools to determine the potential risk to occupants from misused pesticides are limited, and
72	there are few known effective cleaning procedures to reduce pesticide levels in affected
73	structures. Proper cleanup can also be very costly, presenting significant challenges for
74	individuals with limited economic resources. As a result, occupants could continue to inhabit
75	contaminated buildings, could be forced to vacate contaminated properties, or could attempt
76	to remediate on their own, possibly creating toxic byproducts or further spreading pesticides

residues. Science based remediation methods are needed to safely reduce occupant exposure
following pesticide misapplication incidents.

79 Although the fate and transport of pesticides in the environment has been studied 80 extensively including those completed for pesticide registration [11], there is very limited information on the persistence of pesticides in indoor environments [12-15] where degradation 81 82 mechanisms, such as direct sunlight, water, and soil microbes, do not exist. Numerous studies 83 have documented the presence of many different pesticides inside homes and day care centers in indoor air, in dust and on surfaces [16-21] indicating long-term persistence. Few studies 84 85 evaluate indoor pesticide fate for extended periods but results from previous pesticide misuse cases (reference 8 and personal communication) suggest that pesticide residues persist indoors 86 87 due to the absence of the primary degradation factors found outdoors. To confirm that 88 pesticides of interest (malathion, carbaryl, deltamethrin, fipronil, and permethrin) do persist in 89 the indoor environments, the present study included persistence tests conducted in a 90 controlled environment under dark and indoor light conditions, to assess the rate of dissipation of the pesticides from the surface via volatilization and/or degradation. 91 Managing outdoor pesticide spills [22] and remediating pesticide contaminated soil or 92 93 water traditionally focus on control, containment and various cleanup technologies. The cleanup procedures following an outdoor release tend to focus on the physical removal of the 94 95 contamination and, if applicable, leaving pesticide residues associated with normal application conditions in place to breakdown naturally. Leaving residues to degrade naturally indoors may 96 not be a suitable approach due to extended indoor persistence (reference 8 and verbal 97

98	communication). Additionally, when pesticide misuse results in an unsafe environment or
99	occupant evacuation, more immediate and proactive residue removal or reduction is required.
100	Information about decontamination approaches for pesticides on indoor surfaces is almost
101	nonexistent and knowledge on complete chemical degradation pathways is limited to general
102	concepts derived from processes observed in water or soils. Initial attempts to remediate
103	pesticide contaminated surfaces through general washing and physical removal are rarely
104	successful [7-9] and physical removal may not be possible for all surfaces. Pesticide
105	manufacturer labels or safety data sheets (SDS) identify decontaminants such as chlorine
106	bleach, caustic soda, or lime without evidence or reference to degradation rates. Use of these
107	products by building occupants may pose health risks and may not be practical for use on
108	common household items or for extensive use throughout grossly contaminated residential or
109	business settings. Further, potentially toxic byproducts may be formed during decontamination,
110	which would require additional chemical analysis and costs in order to verify a successful
111	cleanup.
112	To inform safer and effective application of decontaminants, we conducted a series of
113	decontamination experiments on malathion, carbaryl, deltamethrin, fipronil, and permethrin
114	contaminated materials. Because these studies were the first of their kind, they focused strictly
115	on chemical interactions to degrade pesticides on indoor surfaces to better understand efficacy
116	independent of various scrubbing, rinsing and other physical removal procedures. In addition,
117	physical removal processes introduce numerous variables beyond the scope of this
118	investigation.

119 The efficacies of selected commercially available decontamination solutions (Spic and Span® cleaner, Clorox<sup>®</sup> Bleach, EasyDECON DF200<sup>®</sup>, and Sterilex<sup>®</sup> Ultra-Kleen) were examined for 120 121 their ability to degrade the pesticides under operationally realistic conditions such as 122 application of a representative decontaminant volume per surface area and contact time. These 123 solutions were selected based on their observed effectiveness (Sterilex® Ultra-Kleen) for 124 remediating organophosphate pesticide (methyl parathion) contaminated homes [8], and on 125 the chlorine-based oxidation chemistry (Clorox® Bleach) [23], because they were designed to degrade organophosphate chemical warfare agents (EasyDECON DF200<sup>®</sup>) [24], or because they 126 127 are a conventional, commercially available detergent solution (Spic and Span<sup>®</sup> cleaner). This 128 included a measurement of the efficacy of decontamination solutions on three building 129 materials (stainless-steel, plywood, and vinyl flooring) using representative decontamination 130 solution dwell times on the pesticide contaminated surfaces and contamination levels 131 measured in pesticide misuse incidents (25–2,400 µg/100cm<sup>2</sup>). The measured contamination 132 levels vary due to (1) the amount and concentration of the product applied at the site; (2) the 133 surface types sampled and sampling methods used; (3) the time that passed between the application and the sampling; or (4) previous applications and residues from other sources, 134 135 such as being tracked-in from outdoors or from use of pet products containing the same pesticides. SDS and other health and safety information on any decontamination solution 136 137 should be reviewed as the decontamination solution itself may introduce an additional exposure risk to personnel. This study included a semi-quantitative analysis for known 138 byproducts of noticeable toxicity following the decontamination process which were identified 139 for two pesticides, malathion and fipronil. 140

#### 141 **2. Materials and Methods**

142 2.1 Pesticides

Targeted pesticides were technical grade malathion (CAS Number 121-75-5), carbaryl (CAS 143 144 Number 63-25-2), fipronil (CAS Number 120068-37-3), deltamethrin (CAS Number 52918-63-5), and permethrin (CAS Number 52645-53-1) as well as two commercial pesticide formulations, 145 146 Ortho® MAX® Malathion Inspect Spray Concentrate [Ortho MAX] (The Scotts Company LLC, 147 Huntsville, TX) and Sevin® Carbaryl Insecticide [Sevin] (TechPac, LLC, Atlanta, GA). All pesticides 148 and pesticide containing formulations that were part of this study are tabulated in Table 1 and were procured from either a commercial source (Sigma Aldrich, St. Louis, MO) at 97% or higher 149 150 purity or as commercial formulations from local vendors. Labeled internal standards of 151 pesticides were purchased from Cambridge Isotope Laboratories (Tewksbury, MA, USA), CDN 152 Isotopes (Pointe-Claire, Quebec, Canada) and Dr. Ehrenstorfer® GmbH (Augsburg, Germany). 153 The pesticides selected are common use, registered insecticides that have been misused 154 indoors [2-8]. They also represent a range of pesticide classes and physiochemical properties as 155 to evaluate their behaviors on surfaces and chemical interactions with decontamination 156 solutions. Surface contamination levels, as tabulated in Table 1, were different for each 157 pesticide based on the highest observed surface concentrations reported in several pesticide 158 misuse investigations conducted by state pesticide regulatory agencies and shared with US EPA. 159 This study utilized these pesticide surface concentrations because a high level of surface 160 contamination is likely more difficult to cleanup than a lower level of surface contamination. 161 Technical grade pesticides were dissolved in n-hexane (>98.5%, mixture of hexane isomers, HPLC, GC, pesticide residue analysis grade) or dichloromethane (>99.9% HPLC, GC, pesticide 162

163 residue analysis grade) at stock concentration values that allowed for the application of a single 164 10 µL droplet of the dissolved pesticide on the targeted surfaces to reach the intended surface concentration. After chemical analysis of the concentration of target chemicals in commercial 165 166 pesticide formulations, the Ortho MAX malathion-containing product was mixed with deionized water (1:125 ratio) to create the same malathion concentration on the surface as tested using 167 168 the technical grade. Similarly, the Sevin carbaryl containing product was diluted 8.3-fold with 169 deionized water to create the same carbaryl surface concentration for a direct comparison with the technical grade decontamination tests. 170

171 2.2 Test surface materials

172 Test materials were selected to represent a variety of commonly encountered indoor surfaces with potential different degrees of permeability to pesticides. The materials selection 173 was based on its likely use as a subflooring material (plywood) under carpet or hardwood 174 flooring, or actual flooring material (vinyl) in a residence. Stainless-steel was representative of a 175 nonporous surface and served as a reference material since treatment and removal methods 176 177 are expected to perform more efficiently on this material. Materials were also selected for 178 relative simplicity to minimize potential interactions due to surface material interferences. For example, painted or stained surfaces commonly found in homes may interact with the 179 180 pesticides and decontamination solutions and complicate interpretation of results. Further fate and transport research are needed on more complex surfaces before decontamination 181 182 approaches can be developed. All test substrates were spiked with a pesticide and underwent 183 decontamination treatments. Large sections or panels of each material were obtained from 184 suppliers (stainless-steel, 304 Grade; McMaster-Carr; plywood, untreated pine plywood, Lowe's

185	Home Improvement; and vinyl, TrafficMaster Allure, Lowe's Home Improvement). Small
186	rectangular coupons (2.5 $ imes$ 4.0 cm dimensions) were cut from the larger materials and were
187	used as substrates for the application of the pesticides and subsequent decontamination steps.
188	All materials were cleaned prior to use by removal of any dust followed by a surface cleaning
189	with methanol. Material coupons were spiked with a single 10 $\mu$ L droplet, using a gas tight
190	micro-syringe (SGE Analytical Science, Melbourne, Australia), of the targeted pesticide at
191	pesticide specific stock concentrations in hexane or dichloromethane.
192	2.3 Decontamination solutions
193	Four decontamination products were initially chosen for testing. EasyDECON <sup>®</sup> DF200
194	(hereafter, DF200) solution was prepared by proportional mixing of DF200 Parts 1-3 (hydrogen
195	peroxide solution as the active ingredient with surfactants; pH = 9.8). DF200 Part 1 is composed
196	of cationic detergents and fatty alcohols; DF200 Part 2 is an 8% hydrogen peroxide stabilized
197	solution; and DF200 Part 3 contains diacetin. Sterilex Ultra Kleen Solution 1&2 (hereafter,
198	Sterilex) was prepared by proportional mixing of Solution 1 and 2 (peroxide solution, pH = 11).
199	Sterilex Solution 1 contains hydrogen peroxide (6-6.6% by weight) as the active ingredient,
200	quaternary ammonium compounds and ammonium salts while Solution 2 contains sodium- and
201	potassium carbonates (approximately 6% by weight). Clorox <sup>®</sup> Concentrated Germicidal Bleach
202	(8% sodium hypochlorite as the active ingredient, pH = 11.4) (hereafter, bleach) was applied as
203	received without dilution. Spic and Span <sup>®</sup> Liquid Multi-Surface and Floor Cleaner (hereafter,
204	Spic&Span) solution was prepared per manufacturer instructions (sodium carbonate as the
205	active ingredient and surfactant, pH = 9.4-9.7). All decontamination solutions were prepared
206	immediately prior to use. The use of full-strength bleach without dilution to clean surfaces is

not recommended as per the manufacturer's label. Previous decontamination testing using tenfold diluted bleach on materials contaminated with organophosphate nerve agents suggested
low efficacy [23]. Therefore, this bench-scale study used full strength bleach to evaluate
whether a concentrated solution would degrade the pesticides more completely without
leaving toxic byproducts.

212 After a 5-min contact time of the pesticide solution with the flooring or nonporous 213 reference material (to allow the solvent to evaporate), 75 µL of the decontamination solution was applied as a single droplet over the contaminant using a gas tight micro-syringe (SGE 214 215 Analytical Science, Melbourne, Australia). Considering the small coupon sizes, the 216 decontamination solution was applied as a single droplet that was large enough to cover the 217 pesticide contaminated area on a coupon. This volume is representative of the volume of 218 decontaminant solution applied to a surface from a back-pack type sprayer [23]. The 219 decontamination solution was allowed to interact with the pesticide on the surface for 18 hours 220 (representing an overnight drying of the decontamination solution). For a second set of test 221 coupons, the same decontaminant with the same volume was reapplied after 90 minutes 222 followed by 16.5 hours contact time. Both sets of test coupons, as well as the positive controls 223 (see below), were extracted at the same time (18 hr after contamination). Each test point 224 consisted of two sets of three replicate test coupons (contaminated and decontaminated 225 material). Each test point also included two positive controls that were contaminated with a 226 pesticide but not decontaminated; one procedural blank that was not contaminated with a 227 pesticide, but the decontamination solution was applied; and one laboratory blank that was not treated with either pesticides or decontamination solutions. 228

The decontamination testing started with all the decontaminants (DF200, Sterilex, bleach, Spic&Span) applied against the malathion contaminated materials (stainless-steel, plywood, and vinyl). Subsequent decontamination testing for other pesticide contaminated materials was limited to the better performing decontaminants and to a lower number of materials (see Table 232).

234 2.4 Pesticide persistence tests

Persistence tests were conducted for the targeted pesticides as applied to stainless-steel 235 coupons (10 cm<sup>2</sup> surface area). Fourteen sets of triplicate stainless-steel coupons plus one 236 triplicate Day 0 set were contaminated with a pesticide mixture containing malathion, carbaryl, 237 238 fipronil, deltamethrin, and permethrin at concentrations to reach the surface concentration in Table 1. All contaminated coupons, as well as single procedural blank coupons (no pesticide 239 240 applied), were placed in two chambers with controlled temperature  $(24 \pm 3^{\circ}C)$  and  $50 \pm 3^{\circ}$ relative humidity (RH). Seven sets of contaminated coupons were placed in a dark chamber. 241 The other seven sets were placed in the second chamber that simulated an indoor light 242 243 environment. The indoor lighting was simulated using seven 4 Watt fluorescent warm white 244 light fixtures (color temperature 3000 K). Light fixtures were present in close distance to the coupons (2-5 inch range). The air exchange rate for both chambers was controlled at one air 245 246 exchange per hour representing a common air change rate for standard residences [25]. At 247 seven intermediate time points up to 140 days post pesticide application, coupon sets including 248 the procedural blank coupon were removed from both chambers and extracted to quantify residual pesticide mass. One set of triplicate coupons was extracted immediately after spiking 249 250 with the pesticide and functioned as the Day 0 starting pesticide mass.

251 2.5 Extraction of pesticides from materials and neutralization of residual decontaminant After a 30-min contact time between the pesticide and surface, residual pesticide amounts 252 were recovered by transfer of the coupon into an extraction vial containing 50 mL of n-hexane 253 254 as the extraction solvent. Vials were then sonicated for 10 min. Following sonication, a 10 mL aliquot of the extract was solvent exchanged with a 10 mL 3:7 water: methanol ratio (v/v) 255 256 solvent mixture for LC-MS/MS analysis. Extracts that were to be analyzed by GC-MS remained in the hexane solvent. Extracts were diluted as needed, spiked with internal standard (See Table 257 A, Supporting Materials) and stored in a refrigerator (4°C) or freezer (-20°C) for interim storage 258 259 until chemical analysis. Extraction efficiencies, defined as the percent ratio between recovered 260 pesticide mass from a coupon material and that from a spike control into the same extraction 261 solvent, were determined for all material / pesticide combinations prior to the decontamination 262 study. Almost all recoveries (see Table A, Supplemental Materials) using this method exceeded the initial targeted recovery value (better than 70%, but not higher than 120%, recovered mass) 263 with a low coefficient of variance among test coupons (less than 30%). In general, extraction 264 efficiency increased for the extraction of pesticides from plywood to vinyl to stainless-steel. 265 266 This can be attributed to the more porous nature of plywood and vinyl versus stainless-steel. 267 Extraction of both permethrin and deltamethrin from vinyl and plywood did not reach the 70% 268 target extraction efficiency but were in the 47-57% range, which were still deemed acceptable as lower recoveries would be associated with both positive controls and test coupons leading to 269 a limited impact in calculated decontamination efficacy. 270

271 It is critical that the extract is neutralized as residual decontaminant that is part of the
272 extract may continue to degrade the pesticide during the extraction and handling of the extract

273	prior to the analysis. This would bias the recovered pesticide amount leading to artificially
274	higher decontamination efficacy values than those associated with just the degradation of the
275	pesticide on the surface with the decontaminant. The selection of hexane as a nonpolar solvent
276	simplified the needed quenching of the decontamination reaction in the extract as the
277	pesticides partitioned into this solvent layer while the residual decontaminant formed a
278	separate aqueous layer. By taking an aliquot from the solvent phase rather than the reactive
279	ingredient-containing aqueous phase, a further separation of reactant (the residual
280	decontaminant) and pesticide is guaranteed. This was verified prior to the actual
281	decontamination testing through spiking of a pesticide into a simulated extract.
282	2.6 Analytical methods
283	Chemical analysis of the coupon extracts occurred using two analytical methods, gas
284	chromatography/mass spectrometry (GC/MS) for malathion (decontamination tests only) and
285	liquid chromatography/tandem mass spectrometry (LC-MS/MS) for malathion (persistence
286	tests only), carbaryl, fipronil, deltamethrin, and permethrin. Details on the analytical methods
287	are provided in the Supplemental Materials. The lowest recovered pesticide mass is reported at
288	the method quantification level (MQL), which was defined in this study as the lowest calibration
289	curve standard for each pesticide in the study. All malathion extracts were screened for the
290	presence of malaoxon, a toxic oxidation byproduct of malathion [26,27]. For the fipronil
291	decontamination tests, extracts were screened for the presence of fipronil sulfone, fipronil
292	amide, and fipronil desulfinyl as fipronil byproducts of noticeable toxicity [28,29].

293 2.7 Decontamination test matrix, calculations, and statistical analysis

294 Table 2 summarizes the overall decontamination test matrix. Not all decontaminantpesticide combinations were evaluated. Results from the first pesticide (malathion) 295 296 decontamination tests were considered in the selection of decontamination solutions for the 297 second pesticide (carbaryl) with two decontaminants being selected for the other three pesticides. Decontaminants that were excluded from the test matrix in this study should not a 298 299 priori be considered ineffective unless explicitly noted. This decontamination study focused on 300 the identification and efficacy measurement of decontamination solutions that were expected to be efficacious based on the limited literature on surface decontamination via chemical 301 degradation of similar pollutants [2,24]. Decontamination efficacy tests were also executed for 302 303 two commercial pesticide products applied (at the same pesticide surface concentration) to 304 address impacts of product formulation on the observed efficacy.

The decontamination efficacy was defined as the percentage decrease in the mean pesticide mass recovered from the decontaminated (test) coupons ( $\overline{M}_{TC}$ ) compared to the mean pesticide mass applied to the test coupons (positive controls, ( $\overline{M}_{PC}$ )). To account for possible losses, e.g., due to volatilization or natural degradation, the positive control coupons (contaminated but not decontaminated) were extracted at the same time as the test coupons (i.e., after 18 hr) and were used to account for such a decrease. For a given test coupon (*TC*), the response variable calculated for this analysis was:

312 
$$Decontamination Efficacy = 100 \times \frac{\overline{M}_{PC} - \overline{M}_{TC}}{\overline{M}_{PC}}$$

The standard deviation in the mean mass recovered from test and positive control coupons was
used to derive the standard deviation in decontamination efficacy through propagation of
error.

316	Two-tailed Student's t-tests were used to compare the means of the residual pesticide mass
317	following the application of the decontamination solution. Unequal variance between the
318	populations was assumed. A p-value is the result of such comparison and results are considered
319	significantly different if p< 0.05. The statistical analysis included comparisons between pesticide
320	residuals following different decontamination solution applications per pesticide; comparisons
321	in pesticide residuals between single and double decontamination solution applications; and
322	comparisons in residuals among the three materials per decontamination solution.
323	3. Results
324	3.1 Persistence of pesticides in simulated indoor environment
325	The mean recovered mass for carbaryl, fipronil, deltamethrin, and permethrin from a
326	stainless-steel material surface as function of time (up to 140 days) is shown in Figure 1.
327	Persistence test results for malathion are not reported due to high inconsistencies in the
328	extracted malathion within the first two weeks of the experiment. This was attributed to the
329	high dilution ratio of the extract prior to analysis leading to concentrations close to the MQL.
330	The malathion persistence test was not repeated. Analysis of the deltamethrin samples was
331	limited to five time points as extracts were lost due to a solvent extraction step error for two
332	time points.
333	None of the pesticides were detected on any of the procedural and laboratory blanks. In a
334	dark environment, all pesticides were found to be highly persistent with near equal amounts of
335	pesticide recovered after 140 days compared to the contamination level at the start. Initial
336	contamination levels were based on observed surface concentrations from various pesticide

337 misuse investigations. In the presence of simulated indoor light conditions, some degradation of most of the pesticides was observed. Estimated half-life values on a nonporous stainless-338 339 steel surface in an indoor light environment were derived from a single first-order kinetics model which specifies that the rate of concentration decline is proportional to the 340 concentration in the system. Fitted half-life values are tabulated in Table 3. Half-life values 341 342 range from 166 days for fipronil and deltamethrin to 286 days for permethrin. A linear fit to the 343 dark indoor condition persistence data for carbaryl, fipronil and deltamethrin resulted in slope values that were not statistically significantly different from zero (p<0.05). Hence, half-life 344 345 values in dark environments could not be calculated based on these data but are expected to exceed 500 days. No appreciable degradation was observed for carbaryl on stainless-steel over 346 347 the 140-day period in both dark and indoor light environments. The initial increase (Day 1 to 348 Day 21) in recovered amounts of carbaryl from the stainless-steel coupons in both the dark and 349 indoor light environment is probably due to an analytical bias in the highly diluted carbaryl 350 samples. For comparison, Table 3 includes reported half-life values for water and soil. Reported ranges in water and soil half-life values are mainly due to their dependence on pH in water or 351 352 the soil type and depth [30-36]. The analysis of the extracts by LC-MS/MS did not include 353 identification of possible degradation products except for the fipronil persistence test where fipronil sulfone was included in the analysis. Fipronil sulfone (data provided in Supplemental 354 355 Materials) was present in each sample (approximately 200 ng/coupon or less than 2% of 356 applied fipronil mass) from Day 0 to Day 140 with no noticeable change in mass recovered as 357 function of time and independent on dark/indoor light conditions.

358 3.2 Decontamination results for indoor building materials contaminated with malathion

359 Decontamination efficacy values for a single or double application of the decontamination solution followed by an 18 hr contact time (simulating an overnight drying) are shown in Figure 360 2 for indoor materials contaminated with 400  $\mu$ g/100cm<sup>2</sup> malathion. For malathion, 361 362 decontamination using Spic&Span left more than 50% of the initial malathion applied on the 363 surface after an 18 hr contact time. All other tested decontaminants (DF200, Sterilex, and 364 bleach) yielded greater than 96% reduction in malathion on stainless-steel and vinyl. 365 Decontamination of plywood was noticeably less efficacious than that of stainless-steel or vinyl except when bleach was used. A second application of the same decontamination solution 90 366 367 min after the first application improved the efficacy for plywood. For the other two materials, residual malathion levels were already at or just above the MQL (equivalent to 2.5  $\mu$ g/100cm<sup>2</sup> 368 369 malathion) after one application of DF200, Sterilex, and bleach. Hence, the impact of a second 370 application for these two materials could not be established. Residual malathion levels after a 371 single or double application are tabulated in Table 4. Malathion was not detected on any of the 372 procedural and laboratory blanks. 373 Residual malathion amounts were highest when using Spic&Span followed by Sterilex,

DF200 and bleach for the stainless-steel plywood, and vinyl materials. A statistical comparison of residual malathion amounts following decontamination showed that residue levels were only significantly different (p<0.05) between those following the Spic&Span application and any of the other three decontaminants, while residuals among the three decontaminants were not significantly different statistically (p>0.05). The observed reduction in residual malathion associated with two applications of decontamination solution were not significantly different from the results following a single application of decontamination solution (p>0.05). When

comparing malathion residuals after a double decontamination application, plywood generally
 had the highest amount of malathion remaining followed by vinyl and stainless-steel. However,
 the residual malathion on plywood was not significantly different statistically (p>0.05) from that
 on stainless-steel or vinyl. The only exception was for the Spic&Span product where the
 residual amount of malathion on plywood was significantly different from that on vinyl or
 stainless-steel.

387 All malathion extracts were screened for the presence of malaoxon, a toxic oxidation

388 byproduct of malathion. One extract out of three replicates associated with the bleach

decontamination of the plywood contained malaoxon. The recovered malaoxon mass from this

390 single coupon was not quantified. No malaoxon was observed in extracts following

decontamination with the other decontaminants DF200, Sterilex, and Spic&Span.

The Spic&Span product was not evaluated further against the other pesticides based on its poor decontamination performance against malathion and the lack of an active oxidative ingredient.

3.3 Decontamination results for indoor building materials contaminated with carbaryl 395 Figure 3 summarizes the decontamination efficacies as observed when applying DF200, 396 Sterilex, and bleach for single and double applications onto the three materials contaminated 397 with carbaryl (at a 2400 µg/100cm<sup>2</sup> surface concentration). Efficacies were highest when using 398 399 the DF200 product followed by Sterilex and bleach. For carbaryl, efficacies exceeding 99% were 400 only observed for the DF200 solution when applied to stainless-steel and vinyl. This material dependence was not observed with bleach although bleach efficacies never exceeded 70% 401 402 across all materials. Residual carbaryl levels after one or two decontaminant applications are

403	tabulated in Table 4. Carbaryl was not detected on any of the procedural and laboratory blanks.
404	As was the case for malathion contaminated materials, a double application of
405	decontamination solution did not yield significantly different (p>0.05) residual carbaryl levels
406	except for bleach on stainless-steel. Here, a significant reduction (p=0.02) in residual carbaryl
407	was observed when bleach was applied twice. Although residuals on plywood after any of the
408	decontamination tests were higher than those on other materials, these differences were not
409	significantly different statistically (p>0.05) except when comparing carbaryl residuals on
410	plywood against those on vinyl after decontamination with Sterilex.
411	3.4 Decontamination results for indoor building materials contaminated with fipronil
412	The decontamination efficacy results for fipronil contaminated materials are shown in
413	Figure 4. These decontamination tests were limited to the DF200 and bleach solutions. Both
414	solutions accomplished a better than 99% reduction in fipronil mass (applied at 150
415	$\mu$ g/100cm <sup>2</sup> ), even after a single application except when applied to plywood. The repeated
416	application did not improve decontamination efficacy appreciably for both decontamination
417	solutions. Residual fipronil amounts after decontamination are tabulated in Table 4. Fipronil
418	was not detected on any of the procedural and laboratory blanks. Residual amounts of fipronil
419	after one application were at or just above the MQL (equivalent to 0.5 $\mu$ g/100cm <sup>2</sup> for fipronil)
420	for both decontaminants on stainless-steel and vinyl. In decontamination tests in which residual
421	fipronil was above the MQL, none of the residuals were found to be significantly different
422	(p>0.05) in a direct comparison between the two decontaminants. The changes in residual
423	amount of fipronil after a repeated application of DF200 and bleach were not statistically
424	significant (p>0.05). Plywood was more difficult to decontaminate with bleach with statistically

significant higher amounts of fipronil recovered after a single application. Residuals on plywood
after a second application of bleach were not significantly different compared to the residuals
on the other two materials.

All extracts were screened for the presence of fipronil amide, fipronil desulfinyl, fipronil 428 sulfone as three possible fipronil degradation byproducts [28,29]. All three byproducts were 429 430 detected in small quantities (up to 0.015 µg fipronil amide; up to 0.05 µg for fipronil desulfinyl and up to 0.25  $\mu$ g fipronil sulfone per extract / 10 cm<sup>2</sup> size coupon) in the test coupon extracts 431 of all materials and across both decontaminants. They were also detected in the positive 432 433 control extracts at the same levels/concentrations. Hence, it was impossible to attribute the detection of these three byproducts to their formation during the decontamination process 434 435 with either decontaminant. Further, these byproduct concentrations were in the same order of 436 magnitude to a factor 10 lower than residual fipronil following decontamination of the 437 materials.

438 3.5 Decontamination results for indoor building materials contaminated with pyrethroids 439 The efficacy testing for materials contaminated with the pyrethroids deltamethrin and permethrin (at 25 and 500  $\mu$ g/100cm<sup>2</sup>, respectively) are summarized in Figure 5. 440 Decontamination testing for bleach was limited to stainless-steel material only. For 441 deltamethrin, stainless-steel was the easiest material to decontaminate (better than 94% 442 efficacy) with both DF200 and bleach leading to residuals near or below the MQL of 0.5 443 444  $\mu g/100 \text{ cm}^2$ . Where low efficacies were observed, a second application of the same decontaminant resulted in a slightly improved efficacy. The results for permethrin are different 445 446 from deltamethrin as efficacy values with the DF200 product never reached 55%. Residual

447 contamination levels for these two pyrethroid pesticides after decontamination with DF200 and 448 bleach are summarized in Table 4. Both pyrethroids were not detected on any of the procedural 449 and laboratory blanks. Deltamethrin residuals following decontamination with DF200 were 450 below the MQL (0.05  $\mu$ g/coupon) for stainless-steel while higher residuals were detected on vinyl and plywood. These higher residuals were not significantly different (p>0.05) than those 451 452 for stainless-steel. Deltamethrin residuals following decontamination with bleach were limited 453 to the stainless-steel material. The repeated application of DF200 or bleach did not result in statistically significantly lower residuals of deltamethrin. This statement is biased since the MQL 454 455 for residual deltamethrin was already reached for stainless-steel after a single application of both DF200 and bleach. The differences in residual deltamethrin levels across the three 456 457 materials were not significantly different statistically (p>0.05). Permethrin residuals following 458 decontamination with DF200 were noticeably higher than those following bleach 459 decontamination although there is no direct overlap in the tested materials. The repeated application did not significantly reduce residuals (p>0.05) for either decontaminant on tested 460 461 materials. Residuals pyrethroids on the plywood and vinyl surfaces following DF200 decontamination were not significantly different statistically (p>0.05). 462 463 3.6 Decontamination results for indoor building materials contaminated with commercial pesticide formulations 464 Decontamination efficacy results shown so far were based on the use of a technical grade 465 466 pesticide product. In most pesticide misuse cases, a commercial formulation would have been applied. Figure 6 shows the observed efficacies for the DF200 and bleach products when 467

468 decontaminating a stainless-steel surface contaminated with the formulated pesticide products

469 Ortho MAX (containing 25-50% malathion) and Sevin (containing 43% carbaryl by weight) at the same pesticide loading used for the other decontamination studies. Decontamination efficacies 470 were better than 99.2% with no detectable residual malathion after DF200 or bleach 471 applications on stainless steel surfaces that were contaminated with either technical grade 472 473 malathion or Ortho Max. Efficacies using the DF200 product were somewhat lower when 474 decontaminating stainless steel contaminated with Sevin (88%) than with technical grade 475 carbaryl (>99.9%). Efficacies when using bleach to decontaminate stainless steel contaminated 476 with Sevin was 74% which is slightly higher than when contaminated with technical grade carbaryl (70%). 477

#### 478 **4. Discussion**

479 The persistence test results show that pesticide mass on the tested surfaces does not 480 readily dissipate in indoor environments. Calculated half-life values from a first order 481 exponential decay of the recovered mass data are noticeably longer than half-life in water and 482 comparable to those in soil. The lack of liquid water, sunlight, or only low levels of (different) 483 microbes in an indoor environment will limit the natural degradation of these pesticides. This is 484 consistent with field observation of pesticide residues from applications that were made several years earlier [15]. Further research should consider whether any toxic byproducts are 485 486 formed during the prolonged presence of these pesticides on these indoor materials. 487 The decontamination efficacy tests show that effective decontamination approaches exist 488 that can reduce the initial pesticide mass by more than 95%. The hydrogen peroxide chemistry existing in DF200 and Sterilex products was found to be highly effective for degrading 489 malathion, carbaryl, fipronil, deltamethrin, and permethrin on stainless-steel, plywood, and 490

- 491 vinyl. Hydrogen peroxide is a strong oxidant with a pH dependent electrochemical oxidation
- 492 potential ranging between 0.87 and 1.80 V [37]. Similarly, the hypochlorite oxidation (oxidation
- 493 potential 0.95 V at pH 9.5 [38]) in bleach effectively reduced the mass of all tested pesticides
- 494 except for permethrin. A comprehensive understanding of the decontamination
- 495 mechanisms/reactions cannot be derived from this study alone. Such effort requires liquid
- 496 reactor chemistry experiments that utilize a comprehensive suite of chemical analysis
- 497 instrumentation that were beyond the scope of this study.
- 498 Based on the observed degradation of malathion in the environment, malathion is expected
- 499 to have been initially oxidized at the P=S bond to its P=O oxon analog, malaoxon, which is more
- 500 toxic than the parent compound but would have likely degraded further to O,S-Dimethyl
- 501 phosphorothioate and diethyl succinate [39]. Malaoxon was detected as a malathion
- 502 degradation product in one of the seventy-two decontamination test samples. The very limited
- 503 detection of malaoxon as an oxidation byproduct can be attributed to the continued
- 504 degradation of malaoxon by the same decontaminant, especially under higher pH conditions
- 505 [40]. Similarly, malathion can also rapidly hydrolyze without malaoxon formation at more
- 506 alkaline pH levels as present in bleach (pH >11) [41].
- 507 Degradation of carbaryl is likely to have involved the hydrolysis to 1-naphthol and
- 508 methylamine [42] followed by further degradation into more benign byproducts. The current
- 509 toxicological profile for carbaryl [43,44] does not identify degradation byproducts of significant
- 510 toxicity that should be avoided or minimized in the carbaryl degradation process.
- 511 Degradation of fipronil in the presence of hydrogen peroxide or hypochlorite is expected to
- 512 lead to the intermediate formation of fipronil sulfone and fipronil-desulfinyl (oxidation) and

513 fipronil-amide (hydrolysis) [28,29]. These byproducts were detected in extracts following decontamination of a fipronil-contaminated surface but also in the not-decontaminated 514 positive control extracts. There was no noticeable difference in byproduct concentration 515 between test coupon and positive control coupon extracts which suggests that no additional 516 517 byproducts were created during the degradation by bleach or DF200 of fipronil on these 518 surfaces. 519 The degradation of the pyrethroids deltamethrin and permethrin is expected to follow a 520 combined hydrolysis and/or oxidation process [45]. The hydrolysis is enhanced under alkaline 521 conditions such as observed when using bleach. The current toxicological profile for these pyrethroids [45] does not identify degradation byproducts of significant toxicity that should be 522 523 avoided or minimized in the permethrin or deltamethrin degradation process. 524 The use of full-strength bleach without any dilution to clean surfaces is not recommended 525 under normal conditions. Further research is needed to address whether a diluted bleach product can still be as efficacious as observed in this study. Such effort would also need to 526 527 verify whether the degradation reaction may become incomplete leaving malaoxon as a persistent toxic degradation byproduct on the surface. 528 529 The limited improvement in efficacy and associated limited reduction in residual pesticides 530 on these surfaces by including a second application suggests that the amount of decontaminant 531 on a molar basis in a single application is enough to reach high efficacy. Efficacy for bleach was not significantly affected by the presence of other ingredients that are included in the tested 532 commercial formulations Ortho Max and Sevin. When DF200 was used to decontaminate the 533 Sevin product on stainless steel, more unreacted carbaryl remained on the surface in 534

535 comparison to the decontamination of technical grade carbaryl on the same surface. This

536 suggests that other ingredients in the Sevin product provide demand for the DF200 active

537 ingredients which prevents a full degradation of carbaryl on the surface. Such loss may be

538 overcome through reapplication of the decontamination product,

539 Investigations of the efficacy of surface decontaminants, against VX, a highly persistent OP 540 compound, produced comparable results. Efficacy values for bleach against most of the tested 541 pesticides were similar or higher than those obtained for VX [24]. In that study, the reported efficacies were 42-67% after a 24 hr contact time, depending on the material. A 10x diluted 542 543 bleach solution with an added surface wetting agent (trisodiumphosphate) was used and the starting concentration of VX was higher (10 mg/100cm<sup>2</sup> range) in comparison to the pesticide 544 545 surface concentrations. This suggests that the use of diluted bleach in the presence of high 546 levels of contamination may not be as efficacious, however, such decontamination efficacy is 547 chemical specific and can be material dependent. The VX study also evaluated DF200 and the 548 efficacy results are similar to those observed in this pesticide study. In the VX study, efficacies were 75-99% after a 24-hour contact time depending on the material, with one exception of 549 550 noticeably lower efficacy for vinyl tile material (20%). Consistent high efficacy values on vinyl 551 material were observed in the pesticide study.

In this study, initial pesticide surface concentrations were representative of the pesticidespecific levels measured during field investigations involving misapplications of pesticides in homes or businesses. These concentrations varied widely from 25  $\mu$ g/100cm<sup>2</sup> (deltamethrin) to 2,400  $\mu$ g/100cm<sup>2</sup> for carbaryl yielding molar ratios of active ingredient in the decontamination solution over the pesticide solution ranging from approximately 40 (Sterilex/carbaryl) to over

557 22,000 (DF200/deltamethrin). Although this study was limited to one surface concentration per

558 pesticide, it is expected that surfaces contaminated with a lower pesticide surface

559 concentration would be equal or easier to clean as this molar ratio would be larger. For higher

560 pesticide surface concentrations, the molar ratio may become close to the stoichiometric ratio

561 likely resulting in lower decontamination efficacy values.

562 The long contact time (18 hr), simulating an overnight drying of the decontamination 563 solution, allows for degradation reactions to continue potentially longer than if the decontaminant was rinsed or neutralized shortly after application. However, it is expected that 564 565 the eventual drying of the decontaminant through evaporation (occurring in time frame of several hours; no information collected) will limit further degradation beyond the time of 566 567 decontamination solution evaporation. Among the materials tested, residual pesticide amounts on plywood were in most cases higher than those on stainless-steel and vinyl. In addition, the 568 569 variability in recovered pesticide mass from plywood was noticeably higher than those from 570 stainless-steel and vinyl. This may be due to the nonhomogeneous nature of plywood that can lead to variations in the degree of pesticide permeation. Water-based decontaminants applied 571 to the surface would be unable to access pesticides that permeate into the plywood. 572

#### 573 **5. Conclusions**

The bench scale decontamination efficacy test results reported here are a first step in identifying and developing decontamination strategies for indoor environments following pesticide misuse. Further research is needed to evaluate the potential effects of decontamination methods used in the field, including material compatibility, safety concerns, how decontaminant solutions are applied, and the use of physical removal processes, such as

579 scrubbing and/or rinsing of surfaces. Also, the decontamination efficacy values are not 580 connected to achieving a particular cleanup level or health-based criteria. As noticed when 581 decontaminating plywood and as observed in other studies [46] for a painted drywall material, 582 the transport of a pesticide into a permeable material or into paint/coating covering a material 583 plays a significant role in whether these decontamination technologies are equally successful in 584 cleaning surfaces that are abundant in an indoor environment. The presence of these 585 permeable materials in an environment where pesticide misuse occurs may drive the decision to forego in situ cleaning and instead remove the material to meet the health-risk based 586 587 cleanup standard. Such clearance goals are pesticide specific and may be depend on the extent 588 of pesticide application and the potential exposure to the affected indoor surfaces. 589 This study was conducted to provide information to federal, state, tribal, and local agencies 590 about decontamination approaches for residential or commercial buildings to reduce occupant

591 exposure following pesticide misuse incidents. The mechanistic details of the degradation

592 chemistries are pesticide specific, although the same degradation approach may be applicable

593 by first approximation for the same class of pesticides such as pyrethroids, organophosphate or

594 carbamates. Decontamination practices should be tailored to the specific pesticide(s) involved

595 to maximize its efficacy and minimize the risk of transferring contamination within and from the

site. Further, formation of toxic decontamination byproducts should be avoided. This may need

597 to be evaluated on a case-by case basis.

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606	endorsement should be inferred. EPA does not endorse the purchase or sale of any commercial
607	products or services.

608 Appendix A. Supplementary Data

Supplementary data associated with this article can be found, in the online version, at
 *<insert link>*

# 611 6. References

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761		

# 762 Tables and Table Captions

#### 763 Table 1. Targeted Pesticides

Pesticide	CAS #	Pesticide Family	Purpose	Target Surface Concentration (µg/100cm <sup>2</sup> ) <sup>1</sup>
Malathion	121-75-5	Organophosphate	Insecticide	400
Carbaryl	63-25-2	Carbamate	Insecticide	2400
Fipronil	120068-37-3	Phenylpyrazole	Insecticide	<mark>150</mark>
Deltamethrin	2918-63-5	Pyrethroid	Insecticide	25
Permethrin	52341-32-9	Pyrethroid	Insecticide	500
Ortho <sup>®</sup> MAX <sup>®</sup> Malathion Inspect Spray Concentrate	N/A	Organophosphate	Commercial insecticide product (50% malathion)	400
Sevin <sup>®</sup> Carbaryl Insecticide	N/A	Carbamate	Commercial insecticide product (43% carbaryl)	<mark>2400</mark>

- 764 N/A: Not Applicable
- 765 <sup>1</sup> Based on highest observed surface concentration from specific pesticide misuse cases
- 766

#### 767 Table 2. Test Matrix

Pesticide	Spic&Span	Bleach	DF200	Sterilex
Malathion	Х	Х	Х	Х
Carbaryl		Х	Х	Х
Fipronil		Х	Х	
Deltamethrin		Х	Х	
Permethrin		Х	Х	

768

#### 769 Table 3. Persistence Half-Life values of Selected Pesticides on Stainless-Steel

Pesticide	Half-Life on SS with Light ± SD (days)	Half-Life on SS in Dark (days)	Half-Life in Water (days)	Half-Life in Soil (days)	Soil/Water Reference
Malathion	Not measured	Not measured	1.7-17 <sup>1</sup>	1-17	27,30,31
Carbaryl	ND; >500	ND; >500	4 <sup>2</sup>	16 (at surface) -72 (in soil)	32
Fipronil	166 ± 62	ND; > 500	0.25-0.5 <sup>3</sup>	125	33,34
Deltamethrin	166 ± 42	ND; > 500		5.7-209	35
Permethrin	286 ± 91	ND; > 500	0.8-1.1 <sup>5</sup>	40	36

- 770 SS: Stainless-Steel
- 771 ND: Not determined due to lack of decay
- 772 SD: Standard deviation in fitted half-life value

Pesticide	Decontaminant	Recovered mass from SS (Mean ± SD, μg) Number of Applications		Recovered mass from plywood (Mean ± SD, μg) Number of Applications		Recovered mass from vinyl (Mean ± SD, μg) Number of Applications	
		1	2	1	2	1	2
	DF200	0.25*	0.25*	3.8±3.2	3.6±2.2	1.1±0.5	0.6±0.1
Malathian	Sterilex	0.25*	0.25*	8.1±3.0	4.7±3.1	1.2±0.1	1.2±0.5
widiathion	Spic&Span	37±1.6	34±5.5	18±5.2	19±6.6	30±0.2	30±3.0
	Bleach	0.25*	0.25*	0.91±0.27	0.63±0.41	0.25*	0.25*
	DF200	5.0±0.0	0.08±0.02	37±20	31±22	0.05±0.02	0.04*
Carbaryl	Sterilex	49±10	25±12	129±38	83±32	36±19	3.3±1.9
	Bleach	158±20	97±21	141±41	103±23	122±25	107±28
Finronil	DF200	0.05*	0.05*	0.97±0.47	1.5±0.95	0.07±0.02	0.056±0.001
FIPTOTIII	Bleach	0.05*	0.05*	3.9±1.5	3.2±2.5	0.05*	0.05*
Deltamethrin	DF200	0.05*	0.05*	0.24±0.12	0.14±0.09	0.15±0.17	0.17±0.18
	Bleach	0.05*	0.05*				
Dormothrin	DF200			16±2.0	14±2.8	20±7.1	15±1.2
Permetnrin	Bleach	4.3±3.4	3.1±1.8				

#### 773 Table 4. Pesticides Residuals after Decontamination of 10 cm<sup>2</sup> Surfaces.

774 SD: Standard deviation to the Mean

\*: Method quantification level (MQL) with no standard deviation; residuals were at or below this value
 and not quantified.

777

# 778 Figure Captions.

- 779 Figure 1. Recovered pesticide mass as function of time in dark and indoor light conditions on stainless-
- 780 steel. Error bars are one standard deviation from the mean (n=3). Panel A: Carbaryl data. Panel
- 781 B: Fipronil data. Panel C: Deltamethrin data. Panel D: Permethrin data.
- 782 **Figure 2.** Decontamination efficacies for tested decontaminants against malathion. Error bars in efficacy
- 783 are one standard deviation from the mean (n=3).
- 784 Figure 3. Decontamination efficacies for tested decontaminants against carbaryl. Error bars in efficacy
- 785 are one standard deviation from the mean (n=3).
- 786 **Figure 4.** Decontamination efficacies for tested decontaminants against fipronil. Error bars in efficacy

787 are one standard deviation from the mean (n=3).

788	Figure 5. Decontamination efficacies for tested decontaminants against deltamethrin (upper panel) and
789	permethrin (lower panel). Error bars in efficacy are one standard deviation from the mean
790	(n=3).
791	Figure 6: Decontamination efficacies for DF200 and bleach for decontamination of stainless-steel
792	contaminated with malathion and carbaryl using a technical grade standard and commercial

793 products Ortho MAX and Sevin, diluted to equal concentration of the technical grade standard.

# 795 Figures.

796 Figure 1.



797

799 Figure 2.





801 Figure 3.



802

803 Figure 4.







**Decontamination Solution** 

807 Figure 6.



# 810 Supplemental Materials

811

- 812 Table A: Extraction Efficiencies
- 813 Analytical Methods Information
- 814 Persistence data for fipronil sulfone

# 816 Supplemental Materials

817

- 818 Title: Remediating Indoor Pesticide Contamination from Improper Pest Control Treatments:
- 819 Pesticide Persistence Studies and Decontamination Approaches
- 820
- 821 Authors: Lukas Oudejans, Amy Mysz, Emily Gibb Snyder, Barbara Wyrzykowska-Ceradini, Joshua
- 822 Nardin, Dennis Tabor, James Starr, Daniel Stout II, and Paul Lemieux
- 823

### 824 Extraction Efficiencies

- 825 Extraction efficiencies were measured prior to the persistence and decontamination testing as
- to verify whether the extraction method was able to recover the pesticide from the surface.
- 827 Efficiencies are tabulated in Table A.

#### 828 **Table A: Extraction Efficiencies of Targeted Pesticides from Materials**

Pesticide	Mean Percentage of Pesticide Mass Recovered ± SD (n=3)					
	Stainless-Steel	Plywood	Vinyl			
Malathion	103 ± 2	96 ± 16	139 ± 14			
Carbaryl	98 ± 3	113 ± 57	79 ± 8			
Fipronil	93 ± 7	77 ± 9	75 ± 3			
Deltamethrin	75 ± 23	51 ± 14	51 ± 30			
Permethrin	80 ± 21	47 ± 17	57 ± 42			

829

830

#### 831 **Description of Analytical Methods**

# 832 GC/MS analysis of malathion (decontamination study only)

- A 1 μL sample of the extract was analyzed by GC/MS using an Agilent 6980 gas chromatograph
- 834 coupled with a low-resolution Agilent 5973 mass spectrometer. Injection port temperature was
- set at 150 °C. Analytes were chromatographically separated using a DB-5MS column
- $(dimensions, 60 m \times 0.25 mm i.d., 0.25 \ \mu m film thickness)$  with a GC oven temperature profile
- starting at 110 °C (1 min hold) to 250 °C at 15 °C/min (5 min hold), to 300 °C at 15 °C/min (10  $\,$
- 838 min hold). The mass spectrometer was operated in selective ion monitoring (SIM) mode
- collecting chromatograms at masses 127 m/z and 158 m/z (malathion identification), 173
- 840 (malathion quantification), 183 (malathion-d10 internal standard), and 99 m/z (malaoxon).

# LC-MS/MS analysis of malathion (persistence tests only), carbaryl, fipronil, deltamethrin, and permethrin

- 843 Analysis of malathion, carbaryl, permethrin, deltamethrin, fipronil, fipronil sulfone, fipronil
- amide, and fipronil desulfinyl was done using an Agilent (Santa Clara, CA) model 1100 LC with
- an AB Sciex (Foster City, CA) API 4000 tandem mass spectrometer. All compounds were
- separated using an Agilent Zorbax<sup>®</sup> C18 column (3.5 um, 3 x 150 mm) and a mobile phase flow
- rate of 400 uL/min. Compound specific LC-MS/MS settings are provided in Table B:

Pesticide	Ionization	Mobile phase	T <sub>r</sub> <sup>2</sup>	<b>Q1</b> <sup>3</sup>	<b>Q2</b> <sup>3</sup>	Internal
			(min.)			Standard
	Mode <sup>1</sup>	(A:B)1				
malathion	M+18	20:80	3.4	348	127	malathion- $D_{10}$
carbaryl	M+18	20:80	2.5	219	145	<sup>13</sup> C <sub>6</sub> carbaryl
cis-permethrin	M+18	2:98	3.2	408	183	<sup>13</sup> C <sub>6</sub> <i>cis</i> -
trans-permethrin	M+18	2:98	3.5	408	183	permethrin
deltamethrin	M+18	2:98	2.7	521	279	deltamethrin D <sub>6</sub>
fipronil	M-1	15:85	2.9	435	330	
fipronil sulfone	M-1	15:85	3.0	451	282	finranil das E
fipronil amide	M-1	15:85	2.3	453	348	npronii des F <sub>3</sub>
fipronil desulfinyl	M-1	15:85	2.8	387	351	

#### 848 Table B: LC-MS/MS settings

849 1. A = 5 mM ammonium acetate in water. B = methanol.

- 850 2. Retention time.
- 851 3. Q1 and Q2 are the first and second ion mass filters, respectively.
- 852
- 853

#### 854 Persistence of fipronil sulfone

- 855
- 856 Figure 1 shows the amount of fipronil-sulfone recovered from stainless steel coupons as function of time
- under dark and indoor light conditions. Fipronil-sulfone is a degradation product of fipronil and was
- 858 present in the spiked fipronil solution on Day 0.
- 859



860 861 Figure 1: Recovered fipronil sulfone mass as function of time in dark and indoor light conditions on stainless-steel. Error bars are one standard deviation from the mean (n=3). 862

