

Published in final edited form as: Curr Top Med Chem. 2015; 15(9): 886–893.

Anti-Inflammatory and Antioxidant Properties of *Piper* Species: A Perspective from Screening to Molecular Mechanisms

Sarvesh Kumar^{1,2}, Shashwat Malhotra³, Ashok K. Prasad³, Erik V. Van der Eycken⁴, Marc E. Bracke⁵, William G. Stetler-Stevenson¹, Virinder S. Parmar^{3,4,5,*}, and Balaram Ghosh^{2,*}

¹Radiation Oncology Branch, Center for Cancer Research, National Cancer Institute, National Institutes of Health, 8717 Grovemont Circle, Gaithersburg, MD-20877, USA;

²Molecular Immunogenetics Laboratory, CSIR-Institute of Geno mics and Integrative Biology, Mall Road, Delhi –110 007, India;

³Bioorganic Laboratory, Department of Chemistry, University of Delhi, Delhi-110 007, India;

⁴Laboratory for Organic & Microwave-Assisted Chemistry (LOMAC), Department of Chemistry, University of Leuven (KU Leuven), Celestijnenlaan 200F, B-3001 Leuven, Belgium;

⁵Laboratory of Experimental Cancer Research, University Hospital, University of Gent, DePintelaan 185, B-9000 Gent, Belgium

Abstract

Identifying novel therapeutic agents from natural sources and their possible intervention studies has been one of the major areas in biomedical research in recent years. *Piper* species are highly important - commercially, economically and medicinally. Our groups have been working for more than two decades on the identification and characterization of novel therapeutic lead molecules from *Piper* species. We have extensively studied the biological activities of various extracts of *Piper longum* and *Piper galeatum*, and identified and characterized novel molecules from these species. Using synthetic chemistry, various functional groups of the lead molecules were modified and structure activity relationship (SAR) studies identified synthetic molecules with better efficacy and lower IC₅₀ values. Moreover, the mechanisms of actions of some of these molecules were studied at the molecular level. The objective of this review is to summarize experimental data published from our laboratories and others on antioxidant and anti-inflammatory potentials of *Piper* species and their chemical constituents.

Keywords

Anti-inflammatory; Antioxidant;	Cell adhesion	molecules (CAMs); Endotheli	al cells; <i>Pipe</i>	21
longum; Piper galeatum					

^{*}Address correspondence to these authors at the Molecular Immunogenetics Laboratory, CSIR-Institute of Genomics and Integrative Biology, Mall Road, Delhi 110 007, India; Tel: 91-11-2766-2580; Fax: 91-11-2766-7471; bghosh@igib.res.in; virparmar@gmail.com. CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

INTRODUCTION

Plants have been a great source of medicines since thousands of years. Many medicinal plants have been shown to modulate specific cellular and humoral immune functions[1]. Species of the genus *Piper* are among the important medicinal plants employed in various systems of medicine. *Piper longum* L., *Piper nigrum* L. and *Piper galeatum* L. (Piperaceae), commonly known as "long pepper", "black pepper" and "helmet pepper" respectively, are distributed in the tropical and subtropical regions of the world, throughout the Indian subcontinent, Sri Lanka, Middle Eastern countries and the Americas [2–4]. It is an important component of Indian traditional medicine reported to be used as a remedy for treating respiratory tract infection, chronic gut related pain, gonorrhea, menstrual pain, tuberculosis, and arthritic conditions [5–7].

Inflammation is the hallmark in the pathogenesis of various inflammatory diseases such as asthma, chronic obstructive pulmonary disease (COPD), cancer and cardiovascular diseases [8]. Cell adhesion molecules play criticalroles in the recruitment and migration of inflammatory cells to the sites of inflammation [9]. Therefore, a promising therapeutic approach to diminish aberrant leukocyte adhesion isto inhibit cytokine-induced expression of cell adhesion molecules [10–12]. TNF induces free radical generation like H₂O₂ which activates inflammatory signaling pathways, including NF-xB in vascular cells [13-15]. An excess production or decreased scavenging of reactive oxygen species (ROS) has been shown to be implicated in the pathogenesis of diverse metabolic disorders such as atherosclerosis, cancer, diabetes and neurodegeneration. Thus the antioxidant therapy has gained an importance in the treatment of such diseases linked to free radicals [16]. Normally the antioxidant property of a compound is contributed to by its (a) oxygen radical scavenging capacity, (b) the ability to inhibit cellular microsomal P-450 linked Mixed Function Oxidases (MFOs), (c) the ability to suppress the formation of reactive oxygen species (ROS) or (d) inducing the expression of antioxidant genes like NQO1, HO1 and GCLM, etc. by activating a major transcription factor, Nrf2 [17]. In search of novel small molecules from natural sources, we have been studying the genus Piper for many years and have reported their antioxidant and anti-inflammatory activities. We have identified many novel compounds from *Piper* species and synthesized their derivatives to study the structure activity relationships [18-20]. Herein, we have summarized the anti-inflammatory and antioxidant activities of Piper species and their constituents from our laboratories and independent groups across the world.

BIOLOGICAL ACTIVITIES OF PIPER SPECIES

Inflammation is caused by live organisms or soluble antigen, and chemical or mechanical stress on tissue, which is necessary to destroy and/or dilute the injurious materials, and remove the injured tissues. It has been established that cell adhesion molecules are the key players in this process [9]. The migration of the leukocytes to the site of inflammation is regulated partly by the expression of cell adhesion molecules, viz. intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and E-selectin [15, 21]. The expression of these cell-adhesion molecules is induced on endothelial cells by various pro-inflammatory cytokines like TNF-α and IL-1, and by bacterial LPS [22–24].

There are studies on many synthetic drugs that have been shown to inhibit the expression of these molecules but they have many side effects. Therefore, there is a need to develop a safer remedy with fewer ornegligible side effects. We have reported the effect of ethanol, chloroform and hexane extracts of P. longum and P. galeatum on TNF-a induced ICAM-1 expression in human umbilical vein endothelial cells [2, 19, 25]. The expression of ICAM-1 was low on unstimulated endothelial cells, and itsexpression was induced over fivefold by stimulation of endothelial cells with TNF-a. It was observed that the chloroform extract of P. longum exhibits 70 % inhibition of TNF-a induced ICAM-1 expression on HUVECs, followed by hexane and ethanol extracts, which showed around 40 % inhibition [2]. The hexane and the chloroform extracts of P. galeatum showed 35 and 65 % inhibition of TNF-a induced ICAM-1 expression on endothelial cells, respectively, which is a little less than the activities of hexane and chloroform extracts of *P. longum*[2]. However, the inhibitory activity of ethanolic extract of P. galeatum extract is little higher than that of P. longum, it might have another active component present in it [25]. We further studied the most active P. longum's chloroform extract (PICE) in detail to elucidate the underlying molecular mechanism accountable for inhibition of TNF-a induced ICAM-1 expression on endothelial cells. We verified its functional consequences at the cellular level and found that Plongum chloroform extract (PICE) significantly inhibited adhesion of neutrophils to endothelial cells monolayer. This reduction is due to the ability of PICE to block the TNF-α induced expression of cell adhesion molecules, i.e. ICAM-1 and VCAM-1 at 17.5 µg/ml concentration and E-selectin at 15 µg/ml concentration on human umbilical vein endothelial cells. By performing a series of *in-vitro* assays, we showed that the inhibition of ICAM-1, VCAM-1 and E-selectin by PICEis mediated through inhibition of Nuclear factor-kappa B $(NF-\kappa B)$ in endothelial cells. NF- κB is known to be one of the major transcription factors involved in the transcriptional regulation of ICAM-1, VCAM-1 and E-selectin expressionby inflammatory cytokines or bacterial LPS. Further, we have demonstrated the antioxidant activity of PICE; we have shown that PICE inhibits the NADPH-catalysed rat liver microsomal lipid peroxidation significantly [2]. These results suggest a possible mechanism of antioxidant as well as anti-inflammatory activity of PICE. Similarly, Kumar et al. studied the antiinflammatory activity of the oil of P. longum dried fruits in rats using the carrageenan-induced right hind paw edema method [26]. The activity was compared with that of commonly used standard drug ibuprofen. The oil of P. longum dried fruits inhibited carrageenan-induced rat paw edema. The results indicated that this oil had significant (p< 0.001) anti-inflammatory activity when compared with the standard or untreated control [26]. Ghosal et al. have reported that the ethanolic extract of fruits of P. longum and piperinefrom this plant material cured 90 % and 40 % of rats with caecal amoebiasis, respectively [6, 27]. These biological activities of the extract of *P. longum* indicate that it has some active chemical constituents in it. Thus, we further studied the phytochemical analysis of *P. longum*"s extracts [19].

BIOACTIVE COMPOUNDS OF PIPER LONGUM

Piper longum fruit powder (100 g) was extracted with 50 % aqueous ethanol (150 mL) as described earlier [19]. The supernatant (140 mL) collected by centrifugation at 14,000 rpm was dried under vacuum and designated as "ethanolic extract". This was treated with *n*-

hexane (35 mL) and the hexane layer was dried and designated as "hexane fraction". The residual material was treated with chloroform (40 mL) and the chloroform solution was designated as "chloroform extract". The hexane and chloroform extracts of the fruits of *P. longum* were pooled, as they showed almost similar spots on TLC examination. This combined extract was purified by column chromatography over silica gel using a gradient mixture of ethyl acetate and petroleum ether as the eluent; ethyl 3',4',5'-trimethoxycinnamate (ETMC) and piperine (Fig. 1 & Scheme 1), were eluted with 5 and 17% ethyl acetate/petroleum ether solutions, respectively. The structures of ETMCand piperinewere established on the basis of their spectral analysis (IR, ¹H NMR and ¹³C NMR spectra and HRMS) and confirmed by comparison of melting points and spectral data with those reported in the literature [19].

ICAM-1 INHIBITION AND SAR STUDIES OF IDENTIFIED COMPOUNDS

We have tested the ability of isolated compounds in inhibiting the TNF- α induced expression of cell adhesion molecules on endothelial cells as a criterion of their antiinflammatory activities. To elucidate their structure-function-activity relationship, we synthesized different analogues of ethyl 3',4',5'-trimethoxycinnamate (ETMC), and compared the ICAM-1 inhibitory activity of ETMC with those of its synthetic analogues esters (Scheme 1) as well as of the corresponding acids [19]. The pharmacological, biochemical and therapeutic properties of these cinnamates depend upon the pattern of substitution. Cinnamates have attracted wide interest in recent years due to their diverse pharmacological properties. Among these properties, their antioxidant effects have been extensively studied. The structure-activity studies indicate that the substituents in the aromatic ring, the chain length of the alcohol moietyand the presence of α,β-double bond of the cinnamic acid ester have significant effects on the inhibition of the TNF-a induced expression of ICAM-1 [19]. Previously, we have reported that substitution of oxygen with sulfur enhances the ICAM-1 inhibitory activity of coumarins [28]. To test whether substitution of oxygen with sulfur in ETMC would increase its activity, we designed and synthesized novel analogs of cinnamates, thiocinnamates and thionocinnamates and evaluated the activities of these analogs to inhibit the TNF-a induced ICAM-1 expression on human endothelial cells. The initial screening data demonstrated that ethyl 3',4',5'trimethoxythionocinnamate (ETMTC, Fig. 1 & Scheme 1) is the most potent inhibitor of TNF-α induced, VCAM-1, ICAM-1 and E-selectin expression [29]. Structure-activity relationship studies revealed the critical role of the number of methoxy groups in the aromatic ring of the cinnamoyl moiety, the chain-length of the alkyl group in the alcohol moiety and the presence of α , β - C-C double bond in the thiocinnamates and thionocinnamates. The ICAM-1 inhibitory activity data revealed that activity of the cinnamates significantly decreases with; a) an increase in the length of the alkyl chain in the alcohol part, b) a decrease in the number of methoxy groups present in the benzene ring of cinnamates, c) a decrease in the number of hydroxy groups present in the benzene ring of cinnamates. It has been observed that the dihydroxy derivatives of cinnamates were less effective in inhibiting the TNF-α induced expression of ICAM-1 than trihydroxy derivatives [19, 29–31]. After the structure-activity relationship study, we found that ethyl 3',4',5'-

trimethoxythionocinnamate (ETMTC) is the most potent inhibitor of cell adhesion molecules, thus further preclinical and mechanic studies were focused on ETMTC [28].

MOLECULAR MECHANISM OF THE INHIBITION OF CELL ADHESION MOLECULES

The functional consequences of inhibition of cell adhesion molecules by ETMTC at the cellular level revealed that it abrogates the TNF- α induced adhesion of neutrophils to the HUVEC monolayer [31]. Further, we have reported the molecular mechanism underlying the observed activity. We determined the status of NF- κ B activation in ETMTC treated human endothelial cells. ETMTC was found to inhibit TNF- α induced nuclear translocation and activation of NF- κ B by inhibiting phosphorylation and degradation of I κ B α . The inhibition of I κ B α phosphorylation and degradation by ETMTC was seen to be due its ability to inhibit I κ B kinase activity in endothelial cells [31].

ANTIOXIDANT ACTIVITY OF ETMTC

Emerging evidence indicates that reactive oxygen species contribute to diverse signaling pathways [32]. TNF-ainduced oxidative stress activates inflammatory signaling pathways, including NF-κB in vascular cells [33]. As ETMTC inhibits TNF-αinduced NF-κB activation, hence we examined the effect of ETMTC on TNF-ainduced reactive oxygen species generation in endothelial cells. We found that ETMTC significantly inhibited TNFa induced reactive oxygen species generation in endothelial cells. To further demonstrate its antioxidant potential, we measured the anti-oxidant gene expression in ETMTC treated bronchial epethelial cells. Nuclear factor (erythroid-derived 2)-like 2, also known as NFE2L2 or Nrf2, is a transcription factor, which is encoded by the NFE2L2 gene in humans [34, 35]. It regulates expression of several detoxifying enzymes and is capable of protecting oxidative stress-related injury and inflammatory diseases in animals [36]. In response to antioxidants, xenobiotics, metals, and UV irradiation, Nrf2 binds strongly to antioxidant response elements (ARE) sequence and modulates ARE-mediated antioxidant enzyme gene expression [37]. Nrf2 is negatively regulated by Kelch-like ECH-associated protein 1 (KEAP1) and positively regulated by DJ-1 [38]. The expression of Nrf2-regulated the genes: GCLM, HO1 and NQO1 in ETMTC treated epithelial cells as analyzed by qRT-PCR as a surrogate marker of antioxidant potential and Nrf2 activation activity in ETMTC treated human epithelial cells $(0.01-20 \,\mu\text{M})$. We have reported that ETMTC induces antioxidant gene expression in a concentration dependent manner and it is more potent than sulphoraphane at 10 µM concentration, further we found ETMTC to induce Nrf2-regulated anti-oxidant gene expression at protein level as confirmed by western blots [31]. ETMTC modulates the expression of Nrf2 regulators, Keap1 and DJ-1. Further, we reported that ETMTC increases the levels of NQO1-ARE mediated luciferase reporter activity. NQO1-ARE luciferase activity was determined by using stably transfected Beas-2B cells after treatment with ETMTC or SFN as positive control or DMSO as solvent [17, 18]. These findings clearly demonstrate the possible molecular mechanism of ETMTC action for inhibiting cell adhesion molecules [31].

ANTI-ASTHMATIC ACTIVITY IN PRECLINICAL MODEL

Asthma is a chronic inflammatory disease of airways and airway epithelial injury is the hallmark of respiratory diseases and therapeutic targeting of epithelial injury has been suggested to be an effective strategy for controlling these diseases. We have clearly demonstrated that ethyl 3',4',5'-trimethoxythionocinnamate (ETMTC) was the most effective among various cinnamate derivatives in inhibiting inflammatory cell adhesion molecules (CAMs) expression [29, 31]. We further investigated the effects of ETMTC on the features of allergic asthma and epithelial injury in a murine model [18]. We reported that ETMTC treatment to ovalbumin sensitized and challenged mice alleviated airway hyper responsiveness, and airway inflammation. This alleviation of asthmatic features was associated with the reduction in the expressions of various CAMs, NF-κB activation, Th2 cytokines, eotaxin and 8-isoprostane [18]. It increased activities of mitochondrial complexes I and IV in lung mitochondria and reduced cytochrome c and caspase 9 activities in lung cytosol. We have also reported that it decreased the levels of oxidative DNA damage marker in bronchoalveolar lavage fluid and DNA fragmentation of bronchial epithelia in lung sections. ETMTC not only increased the levels of 15-(S)-hydroxyeicosatetraenoic acid but also inhibited goblet cell metaplasia and sub-epithelial fibrosis [18]. These results demonstrate that ETMTC reduces airway inflammation and thus shows its antiinflammatory potential and could be useful in treating asthma.

BIOLOGICAL ACTIVITIES OF PIPERINE

Piperine (Fig. 1) is a major component of black (*Piper nigrum* Linn) and long (*P.longum* Linn) peppers, and is primarily used as a traditional food and medicine [39]. It also exhibits a variety of biological activities, which includes antioxidant, anti-pyretic and anti-tumor properties [40]. Cell adhesion molecules play a critical role in the pathogenesis of inflammatory diseases [41, 42]. To elucidate the anti-inflammatory potential of piperine, we determined its efficacy to inhibit the TNF- α induced expression of cell adhesion molecules on endothelial cells; we found that piperine inhibited the TNF- α induced expression of cell adhesion molecules by inhibiting nuclear factor- κ B (NF- κ B) activation in endothelial cells [20]. Further, we have reported that its ability to inhibit NF- κ B activation is mediated via I κ B kinase inhibition. Recently, independent groups around the globe have reported the pharmacological activities of piperine and shown that it possesses anti-inflammatory, cytoprotective, anticonvulsant, anti-ulcer and antioxidant activities, and analgesic effects [35, 43].

Angiogenesis plays an important role in tumor progression and metastasis. It is regulated by a variety of proangiogenic genes and signaling molecules including epidermal growth factor (EGF), vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF), hypoxiainducible factors, platelet-derived growth factors, angiopoetin-1 and 2, and matrix metalloproteinases [44]. Thus, inhibition of angiogenesis by down regulating pro-angiogenic factor or inducing various anti-metastatic proteins including tissue inhibitor metalloproteinases (TIMPs) could be novel therapy for cancer [45]. *Piper* leaf extract protects against carbon tetrachloride-induced liver fibrosis in rats [46]. Similarly, studies by Doucette *et al.* reported that piperine inhibits HUVEC proliferation and collagen-induced

angiogenesis [47]. Piperine diminishes tumor growth and metastasis in-vitro and in-vivo model of breast cancer. It decreased PMA-induced COX-2 expression and PGE-2 production, as well as COX-2 promoter-driven luciferase activity in a dose dependent manner. Transient transfection studies utilizing COX-2 promoter deletion constructs and COX-2 promoter constructs with specific enhancer elements revealed that the nuclear factorκΒ (NF-κΒ), CCAAT/enhancer binding protein (C/EBP) and activator protein-1 (AP-1), were the predominant contributors to the effects of piperine. Piperine also inhibited PMAinduced NF-κB, C/EBP and c-Jun nuclear translocation [40]. Bae et al. have reported that piperine administration reduced histologic damage and myeloperoxidase (MPO) activity in the pancreas. It ameliorated many of the examined laboratory parameters, including the pancreatic weight (PW) to body weight (BW) ratio, as along with serum levels of amylase and lipase, and trypsin activity. Piperine pretreatment decreased amylase and lipase activity, cell death, and cytokine production in isolated cerulein-treated pancreatic acinar cells via inhibiting the activation of mitogen-activated protein kinases (MAPKs) [48]. Oxygen radical injury and lipid peroxidation have been deemed to be major causes of atherosclerosis, cancer, liver diseases and the ageing processes [49]. Piperine possesses many antiinflammatory properties as reported by our group and others. It has been established in invitro experiments that it protects against oxidative damage by quenching free radicals and reactive oxygen species and hydroxyl radicals. The effect on lipid peroxidation was also reported. Piperine at low concentrations act as a hydroxyl radical scavenger, but at higher concentrations, it activated the Fenton reaction resulting in the generation of hydroxyl radicals [50]. It acts as a powerful superoxide scavenger with IC₅₀ of 1.82 mM and a 52 % inhibition of lipid peroxidation was observed with an IC₅₀ of 1.23 mM. The results showed that piperine possesses direct antioxidant activity against various free radicals [50]. Piperine exhibits antioxidant activity in experimental conditions both *in-vivo* and *in-vitro* conditions through its radical quenching effect and by preventing GSH depletion.

BIOACTIVE COMPOUNDS OF P. GALEATUM

P. galeatum is native to India and grows wild in the forests of Wayanad and Kerala regions. Ten sesquiterpenes and monoterpenes have been reported to occur in the extracts of its fruits, namely β-elemene, δ-elemene, α-humulene, β-caryophyllene, α-copaene, α-ionone, 10-(acetylmethyl)-3-carene, dihydrocarvyl acetate, 1-p-menthen-8-yl acetate and linalyl acetate. We have isolated five compounds from the stems of *P. galeatum*, namely β-sitosterol (Fig. 1), cyclostachine-A, piperine, piperolein-B and a novel amide, viz. 1-(3'-hydroxy-5'-methoxycinnamoyl)-piperidine. The crude extracts as well as the isolated pure compounds were screened for their activity to inhibit the TNF-α induced expression of cell adhesion molecule ICAM-1. Among all, β-sitosterol was found to be the most active compound, which was taken for further studies [25].

ANTI-INFLAMMATORY ACTIVITY OF β-SITOSTEROL

Cell adhesion molecules are expressed on various immune cells and play pivotal role during any inflammatory event. Inhibition of these molecules by any pharmacological molecule is being considered as surrogative marker of the anti-inflammatory property of the molecule [42, 51, 52]. We have reported that β -sitosterol significantly inhibits the TNF- α induced

expression of ICAM-1, VCAM-1 and E-selectin. Cell adhesion assay using human neutrophils has been used to measure the functional correlation of cell adhesion molecules inhibition. We have also reported that β -sitosterol significantly blocks the adhesion of neutrophils to endothelial monolayer. We investigated the status of nuclear transcription factor- κB (NF- κB) and were able to establish that β -sitosterol significantly blocks the TNFα induced activation of NF-κB[25]. In an independent study, Loizou et al. reported similar activity of β -sitosterol in human arterial endothelial cells (HAECs) [53]. They reported that β-sitosterol inhibits significantly intracellular adhesion molecule 1 and vascular adhesion molecule 1 in TNF-a-stimulated HAEC as well as the binding of U937 cells to TNF-astimulated HAEC and attenuates the phosphorylation of nuclear factor-κB p65 [53]. β-Sitosterol isolated from leaves of Nyctanthe-sarbortristis Linn. (Oleaceae) has been reported to posess analgesic activity and anti-inflammatory activities. Analgesic activity has been shown by hot plate test and acetic acid-induced writhings and anti-inflammatory activity by carrageenan-induced hind paw edema method [54]. Recently Han et al. have investigated the effect of β-sitosterol in 2,4-dinitrofluorobenzene (DNFB)-induced AD-like skin lesions in NC/Nga mice. They reported that infiltration of inflammatory cells and the number of scratchings were clearly reduced in the β-sitosterol treated group compared with the DNFBtreated group. β-Sitosterol significantly reduced the levels of inflammation-related mRNA and protein in the AD skin lesions. It significantly reduced the levels of histamine, IgE, and interleukin-4 in the serum of DNFB-treated NC/Nga mice [55]. They further reported that the activation of mast cell-derived caspase-1 was decreased by treatment with β-sitosterol in the AD skin lesions. β-Sitosterol also significantly decreased the production of tumor necrosis factor-alpha from stimulated splenocytes, it inhibited the production and mRNA expression of TSLP through blocking of caspase-1 and nuclear factor-kappaB signal pathways in the stimulated HMC-1 cells [55].

ANTI-OXIDANT ACTIVITY OF β-SITOSTEROL

Uncontrolled production of reactive oxygen species contributes to the pathogenesis of various diseases, such as cancer and cardiovascular disorders. β-Sitosterol has been shown to be associated with cardiovascular protection, exerting its effect mainly through increasing the antioxidant defense system and effectively lowering the serum cholesterol levels in humans. Study results showed that β-sitosterol reverts the impairment of the glutathione/ oxidized glutathione ratio induced by phorbol esters in RAW 264.7 macrophage cultures [56]. Further, it has been reported that β-sitosterol increases manganese superoxide dismutase and glutathione peroxidase activities and decrease in the catalase activities, possibly associated with estrogen receptor (ER)-mediated phosphatidyl inositol 3-kinase (PL3K)/glycogen synthase kinase 3 (GSK3beta) signaling [57]. Antidiabetic and antioxidant potential of β-sitosterol has also been reported in streptozotocin-induced experimental hyperglycemia [58]. In another study, β-sitosterol has been reported to protect rats from 1,2-dimethylhydrazine-induced colon cancer by preventing lipid peroxidation and improving antioxidant status. These data suggested to be associated with the increase in manganese superoxide dismutase and glutathione peroxidase activities and the decrease in catalase activity. They further demonstrated that the effects of β -sitosterol on antioxidant enzymes depend on the estrogen/phosphatidylinositol 3-kinase pathway [58, 59]. These

studies provide further support to the antioxidant potential of β -sitosterol and give new insights into the understanding of the molecular mechanism underlying the beneficial effects of β -sitosterol, and hence those of using *Piper* species in the diet.

CONCLUSION

This review opens new avenues for researchers to identify and characterize novel therapeutic molecules from *Piper* species and investigate their efficacy in preclinical animal models. The *in-vitro* and *in-vivo* studies from our group and others have clearly demonstrated the anti-inflammatory and antioxidant potential of *P. longum* and *P. galeatum* extracts and their constituents. Thus, these findings led us to propose that the use of *Piper* extracts or their constituents as dietary supplements may be useful in the prevention of many diseases like asthma, COPD, cancer and cardiovascular diseases.

ACKNOWLEDGEMENTS

BG received grants (NWP 0033 and BSC 0116) from Council of Scientific and Industrial Research, Govt. of India; VSP was supported by a grant from the EU under its Erasmus Mundus EMA-2 Program.

Biography



LIST OF ABBREVIATIONS

AP-1	Activator protein		

ARE Antioxidant response elements

AD Alzheimer disease

bFGF Basic fibroblast growth factor

CAMs Cell adhesion molecules

C/EBP CCAAT/enhancer binding protein

COPD Chronic obstructive pulmonary disease

DMSO Dimethyl sulfoxide

DNFB 2,4-Dinitrofluorobenzene

EGF Epidermal growth factor

ETMC Ethyl 3',4',5'-trimethoxycinnamate

ETMTC Ethyl 3',4',5'-trimethoxythionocinnamate

GSH Glutathione

GCLM Glutamate-cysteine ligase, modifier subunit

HO1 Hemeoxygenase 1

H₂O₂ Hydrogen peroxide

HAECs Human arterial endothelial cells

HUVEC Human umbilical cord vein endothelial cells

ICAM-1 Intercellular adhesion molecule-1

IL-1β Interleukin 1 beta

KEAP1 Kelch-like ECH-associated protein 1

ΙκΒα Inhibitory kappa B alpha

LPS Lipopolysaccharides

μM Micromolar

mM Millimolar

MFOS Mixed function oxidases

MPO Myeloperoxidase

NF-κB Nuclear factor-kappa B

Nrf2 Nuclear factor (erythroid-derived 2)-like 2

NQO1 NAD(P)H dehydrogenase, quinone 1

PICE P. longum's chloroform extract

SAR Structure activity relationship

TIMPs Tissue inhibitor metalloproteinases

TNF-α Tumor necrosis factor-alpha

TSLP Thymic stromal lymphopoietin

VCAM-1 Vascular cell adhesion molecule-1

VEGF Vascular endothelial growth factor

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OMe MeO 4' 2' MeO
$$\frac{2}{6' 3}$$
 $\frac{2}{0}$ $\frac{2}{1}$ $\frac{2$

Fig. (1). Structures of identified potent inhibitors of cell adhesion molecules (CAMs).

$$\begin{array}{c} \text{OMe} \\ \text{MeO} \\ \text{MeO} \\ \text{O} \\ \text{O$$

Scheme 1.

Synthesis of ETMC and ETMTC(source: *Biochemistry*, 2005. 44(48): p. 15944–52; *Eur J Med Chem*, 2011. 46(11): p. 5498–511).