

## UTid+ In-Clinic Diagnostic System FAQ Sheet

1. Does UTid+ smell while incubating? How bad? What can be done to minimize the odor?
  - a. The plate does not have any smell whatsoever. Bacteria can emit odors, however once the UTid+ test is interpreted (leaving the cover on), the test can be neutralized and discarded; see below.
2. How do you recommend the clinic properly dispose of an incubated test?
  - a. Plates can be inactivated in bleach solution (Clorox 7.5% sodium hypochloride) by soaking the plates directly into the Clorox solution for 30 minutes. Inactivated plates can be disposed in regular trash bins destined for landfill.
3. Is a protective hood, goggles, gloves or any PPE needed or recommended?
  - a. The only required PPE is latex gloves. There is no need for a hood and the plating of samples can be done on a table or counter-top disinfected with diluted alcohol solution.
4. Does the technician require any special handling instructions of the plates?
  - a. UTid+ was design to be a user-friendly culture system and therefore, the technicians are not required any advanced instruction before starting to use our product. The following video has complete instruction of how to plate samples and interpret the results:  
<https://www.youtube.com/watch?v=Pgmf-MeeH2I>  
Technicians are encouraged to contact our 24h-7d tech support at 607-342-8135 if they have any questions about our products.
5. Are there any precautions for handling bacteria?
  - a. Our SDS provides information on our product and not on the bacteria. Culture plates with bacterial growth should be handled with care and technicians should avoid opening the lid of the plates. Plates should be inactivated as described above soon after results interpretation is finished.
6. How do we know if the result is a resistant strain of bacteria? Will a multi-drug resistant (MDR) show the same result as susceptible-(staph)? What do we do if the sample is MDR-staph or other resistant bacteria?
  - a. UTid-plus is diagnostic kit for identification of bacterial pathogens and not an antimicrobial resistant assay. *In vitro* antibiotic sensitivity assay require a higher level of technical knowledge and specialized equipment. If desired, the antibiotic sensitivity profile can be completed at diagnostic laboratories by shipping the UTid+ plate with the bacterial growth to a referral lab.
  - b. What type of applicator is recommended for the plate?  
We recommend the use of a sterile individually wrapped cotton tip swab. E.g.  
[https://www.uline.com/Product/Detail/S-21102/First-Aid/Cotton-Tipped-Applicators-Medical-6?pricode=WB1202&gadtype=pla&id=S-21102&gclid=Cj0KCQjwjo2JBhCRARIsAFG667Woupk3vMkDpEV-OxZtfeoUTTb7Mv3WkL9QbJAZbioNH0heRCBNGt4aAiPhEALw\\_wcB&gclsrc=aw.ds](https://www.uline.com/Product/Detail/S-21102/First-Aid/Cotton-Tipped-Applicators-Medical-6?pricode=WB1202&gadtype=pla&id=S-21102&gclid=Cj0KCQjwjo2JBhCRARIsAFG667Woupk3vMkDpEV-OxZtfeoUTTb7Mv3WkL9QbJAZbioNH0heRCBNGt4aAiPhEALw_wcB&gclsrc=aw.ds)
7. Can you recommend any techniques to capture a sample from ears or skin?
  - a. A sterile cotton tip swab should be used to collect these type of samples.
8. Can you describe what is necessary to conduct a test with a dry sample? (dilution? water? amounts?)
  - a. The cotton swab used to collect the samples should be immersed into 0.5ml of sterile isotonic saline and thoroughly mixed for 20 seconds. Subsequently, the wet cotton swab should be applied thoroughly on the surface of the media of the 4 sections of UTid+ test. If necessary, the swab should be re-immersed into the solution in between sections of the UTid+.