

## Non-Porous High Speed Columns

The high-speed ion exchange columns are designed for high throughput of quality control, process monitoring and other applications, which need rapid scanning of complex samples.

After biocompatible coating, the highly rigid non-porous polystyrene-divinylbenzene (PS-DVB) beads have unique mass transfer characteristics for ultra high-speed chromatography. The Major Advantages of Hydrocell Non-Porous Speed

The Major Advantages of HYDROCELL Non-Porous High Speed Columns are:

- High Speed Throughput Separation
- Higher Sensitivity
- Better Resolution & Efficiency
- Significantly Less Mobile Phase Consumption

### **Ion Exchange:**

HYDROCELL high speed non-porous ion exchange media DEAE NP10, QA NP10, CM NP10, and SP NP10 are prepared from 10  $\mu\text{m}$  non-porous PS-DVB polymeric beads with hydrophilic coating following by immobilizing diethylaminoethyl (DEAE), diethylmethylaminoethyl (QA), carboxymethyl (CM), and sulfopropyl (SP) bonded phases, respectively.

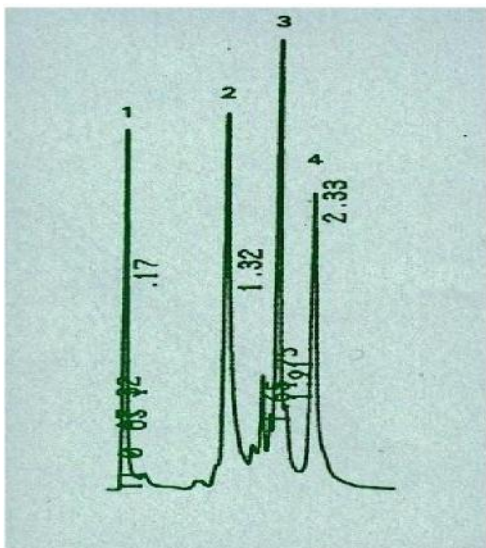
### **Hydrophobic Interaction:**

Similarly, HYDROCELL high speed non-porous hydrophobic interaction media C3 NP10, C4 NP10 and Phenyl NP10 are prepared from 10  $\mu\text{m}$  non-porous PS-DVB polymeric beads with hydrophilic coating following by immobilizing allyl (C3), butyl (C4) and phenyl (PH) bonded phases, respectively.

Both types of non-porous media are compatible with aqueous solutions in the pH range of 1 to 14 and with most polar organic solvents. High-resolution separation of proteins and enzymes can be achieved at high flow rates with steep gradient within a few minutes.

## Hydrocell DEAE NP10

### Protein Mixture



**Column:** 35 x 4.6 mm

**Mobile Phase:**

A: 10 mM Tris, pH 8.0

B: Eluent A + 0.5 M NaCl, pH 8.0

**Gradient:** Linear 0-50% B in 5 minutes

**Detection:** UV 280 nm

**Flow Rate:** 2.0 mL/min

**Injection:** 10  $\mu$ L

**Peak Identification:**

1. Myoglobin
2. Conalbumin
3. Ovalbumin
4. Soybean Trypsin Inhibitor

## Hydrocell QA NP10

### Protein Mixture

**Column:** 35 x 4.6 mm

**Mobile Phase:**

A: 10 mM Tris, pH 8.0

B: Eluent A + 0.5 M NaCl, pH 8.0

**Gradient:** Linear 0-50% B in 5 minutes

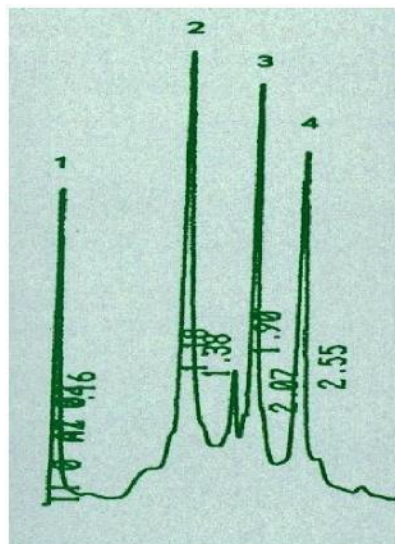
**Detection:** UV 280 nm

**Flow Rate:** 2.0 mL/min

**Injection:** 10 µL

**Peak Identification:**

1. Myoglobin
2. Conalbumin
3. Ovalbumin
4. Soybean Trypsin Inhibitor



## Hydrocell SP NP10

### Protein Mixture

**Column:** 35 x 4.6 mm

**Mobile Phase:**

A: 20 mM Potassium Phosphate, pH 6.0

B: Eluent A + 0.5 M NaCl, pH 6.0

**Gradient:**

Linear 0-100% B in 5 minutes

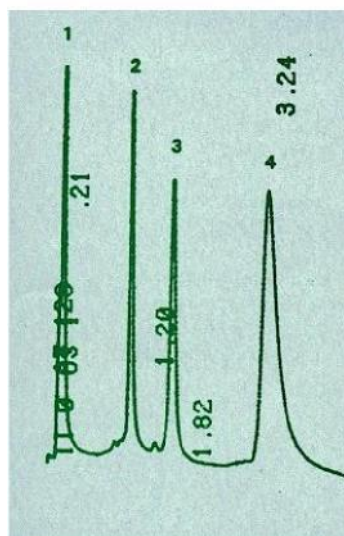
**Detection:** UV 280 nm

**Flow Rate:** 2.0 mL/min

**Injection:** 10 µL

**Peak Identification:**

1. Myoglobin
2. Trypsinogen 3,  
Chymotrypsinogen A
4. Lysozyme



## Hydrocell C4 NP10

### Protein Mixture

**Column:** 35 x 4.6 mm

**Mobile Phase:**

A: 2.5 M Ammonium Sulfate in Eluent B

B: 0.1 M Potassium Phosphate, pH 7.0

**Gradient:** Linear 0-100% B in 5 minutes

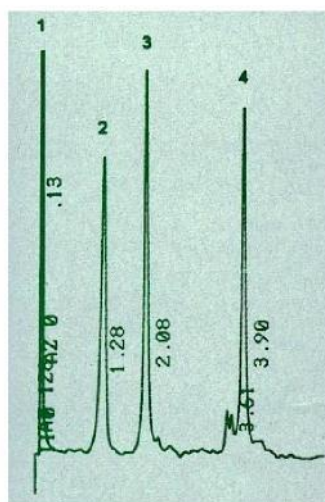
**Detection:** UV 280 nm

**Flow Rate:** 2.0 mL/min

**Injection:** 10 µL

**Peak Identification:**

1. Cytochrome C
2. Ribonuclease A
3. Lysozyme
4. Chymotrypsinogen A



## Hydrocell Phenyl NP10

### Protein Mixture

**Column:** 150 x 4.6 mm

**Mobile Phase:**

A: 2.5 M Ammonium Sulfate in Eluent B

B: 0.1 M Potassium Phosphate, pH 7.0

**Gradient:**

Linear 0-100% B in 20 minutes

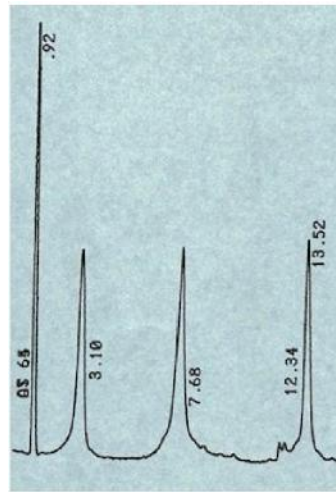
**Detection:** UV 280 nm

**Flow Rate:** 1.0 mL/min.

**Injection:** 20  $\mu$ L

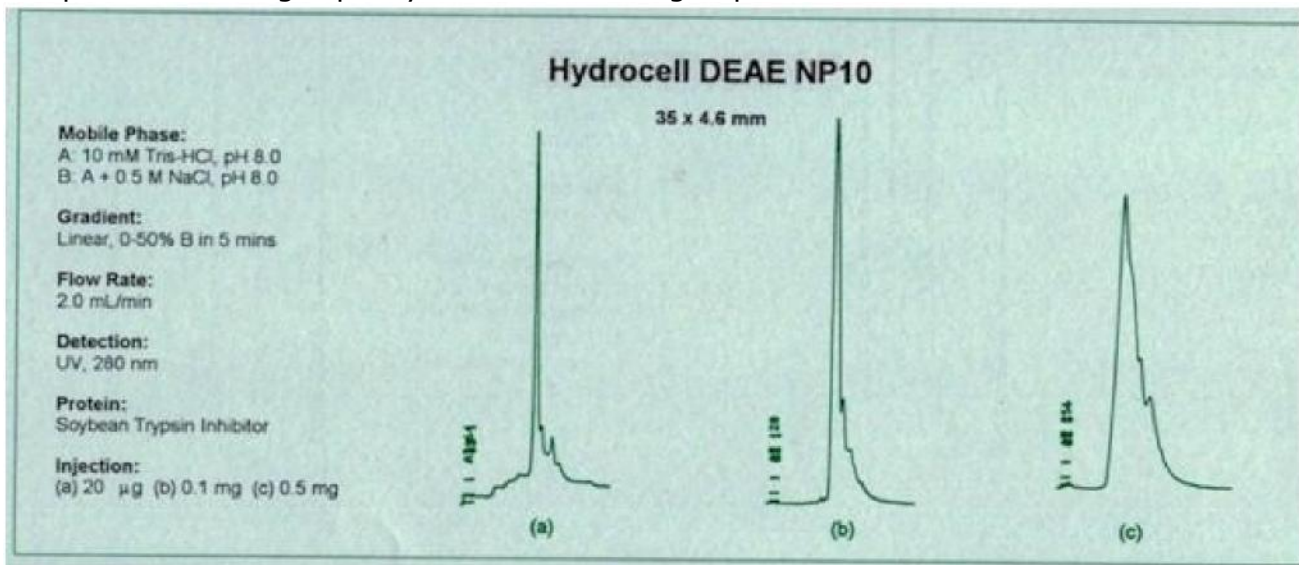
**Peak Identification:**

1. Ribonuclease A
2. Lysozyme
3. Chymotrypsinogen A



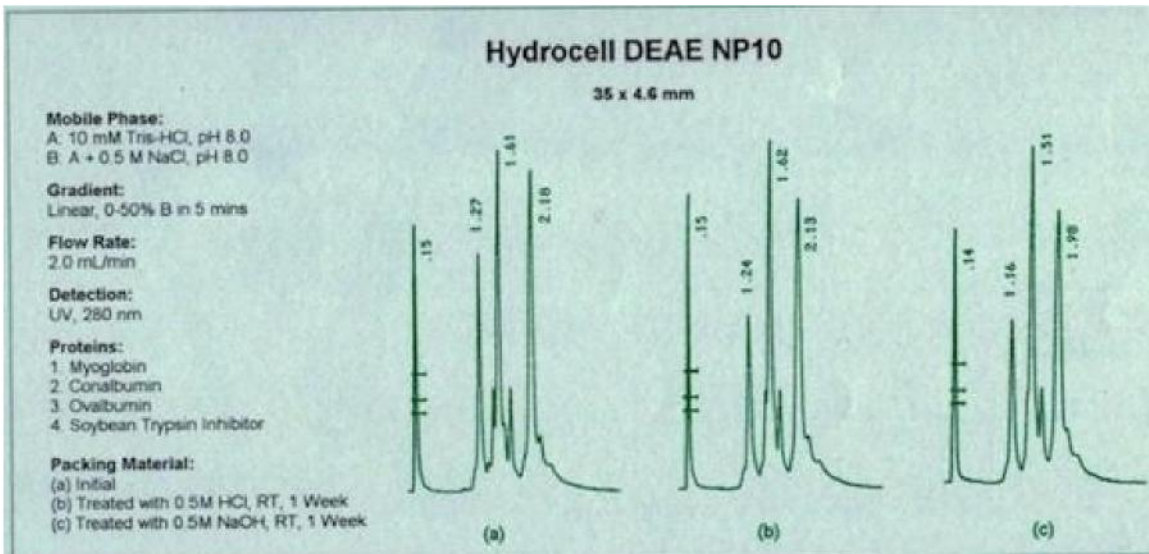
## Loading Capacity

The loading capacity of HYDROCELL NP10 is comparable to many of the porous media available. The dynamic protein loading capacity determined by Frontal Uptake procedure are about 35 mg of ovalbumin, 25 mg of lysozyme and 15 mg of lysozyme per mL of column volume on DEAE NP10, SP NP10 and C3 NP10 columns, respectively. The practical loading capacity is ca. 0.1- 0.2 mg of protein on a 35 x 4.6 mm column.



## Chemical Stability

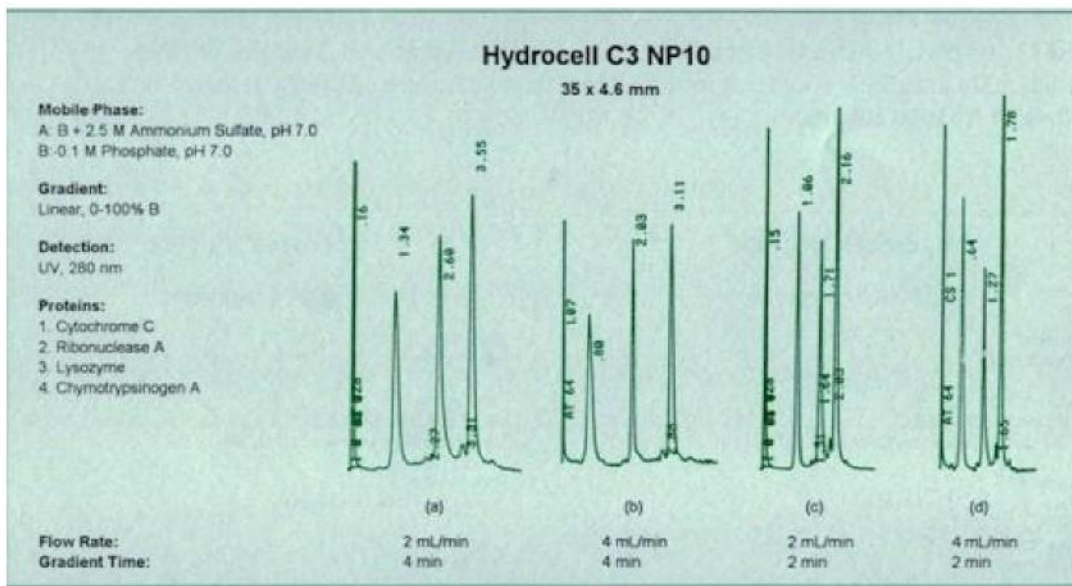
The HYDROCELL NP10 media do not have any ester or amide linkages as do the other commercial products. Their chemical stability allow fast clean up with strong acids (e.g., 0.5M HCl) or bases (e.g., 0.5M NaOH). As shown in the following chromatograms the resolutions among four standard proteins did not show any significant changes when the initial packing material (a) of HYDROCELL DEAE NP10 was treated with 0.5M HCl solution (b) or 0.5M NaOH solution (c) at room temperature for one week.



## Protein Retention Time

HYDROCELL NP10 media do not readily change their physical dimensions under high flow rate or under steep gradient applications. The analysis of proteins and enzymes can be conducted with short run times by either increasing the gradient slope or increasing the flow rate. However, the flow rate does not contribute to the shortness of the analysis time as much as the gradient slope does, especially for high retention macromolecules. The high speed, high resolution separation can be achieved at a moderate flow rate with high gradient slope on a HYDROCELL NP10 column to reduce the solvent consumption.

As shown in the following chromatogram, an increase in flow rate at the same gradient time (a & b, or c & d) sharpens peaks and slightly decreases run time, while a decrease in gradient time at the same flow rate (a & c or b & d) dramatically reduces run time.



## Non-Porous Anion Exchangers

In addition to their use in process monitoring and quality control analysis of proteins and enzymes, the highly efficient DEAE NP10 and QA NP10 high-speed columns are also suitable for the analysis of small nucleotides, oligonucleotides, and DNA fragments. High loading capacity and low back pressure are the unique characteristics of these columns. These characteristics allow the easy scale-up from analytical to preparative separations. The sensitivity of non-porous anion exchangers is better and retention time is shorter than NS 1500. It is recommended for the trace analysis and rapid scanning of complex samples.

Scroll down for Chromatograms.

## **Hydrocell DEAE NP10**

### **DNA Fragments**

**Column:** 35 x 4.6 mm

**Mobile Phase:**

A: 2 M Urea, 20 mM Potassium Phosphate, pH 5.5

B: Eluent A + 1.0 M Ammonium Sulfate, pH 5.5

**Gradient:**

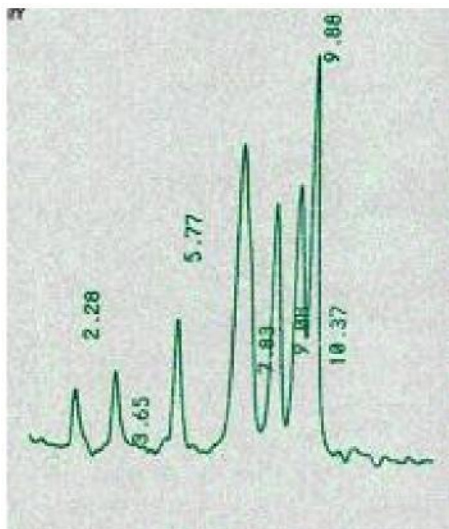
Linear 40-70% B in 15 minutes

**Flow Rate:** 1.0 mL/min.

**Detection:** UV 280 nm

**Sample:** pUC18 DNA Hae III Digest

**Injection:** 10  $\mu$ L



## **Hydrocell DEAE NP10**

### **DNA Fragments**

**Column:** 35 x 4.6 mm

**Mobile Phase:**

A: 2 M Urea, 10 mM Potassium Phosphate, pH 5.5

B: Eluent A + 0.5 M Ammonium Sulfate, pH 5.5

**Gradient:**

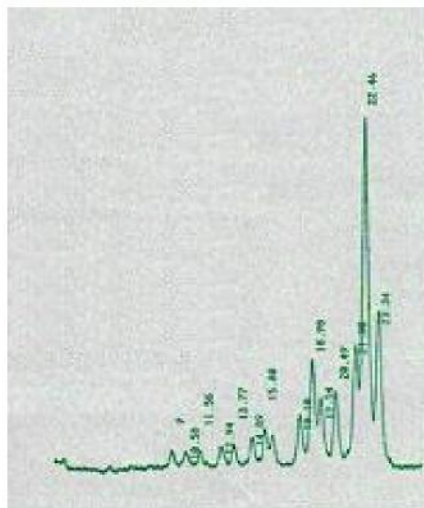
Linear 40-100% B in 30 minutes

**Flow Rate:** 1.0 mL/min

**Detection:** UV 280 nm

**Sample:** pBR322 DNA Hae III Digest

**Injection:** 10  $\mu$ L



## **Hydrocell QA NP10**

### **Oligonucleotides**

**Column:** 150 x 4.6 mm

**Mobile Phase:**

A: 25 mM CHES, pH 8.0

B: Eluent A + 1.2 M Ammonium Sulfate, pH 8.0

**Gradient:**

Linear 10-50% B in 40 minutes

**Flow Rate:** 1.0 mL/min.

**Detection:** UV 260 nm

**Sample:** Oligothymidylic Acid d(pT) 12-18 mer, 5 units in 1 mL of eluent A

**Injection:** 10  $\mu$ L

